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JC962 U.S. PTO

UTILITY PATENT APPLICATION TRANSMITTAL
(Large Entity)*(Only for new nonprovisional applications under 37 CFR 1.53(b))*Docket No.
NEX 10-6Total Pages in this Submission
3**TO THE ASSISTANT COMMISSIONER FOR PATENTS**

Box Patent Application

Washington, D.C. 20231

Transmitted herewith for filing under 35 U.S.C. 111(a) and 37 C.F.R. 1.53(b) is a new utility patent application for an invention entitled:

Systematic Evolution of Ligands by Exponential Enrichment: Photoselection of Nucleic Acid Ligands and Solution SELEX

and invented by:

Larry Gold, Michael Willis, Tad Koch, Steven Ringquist, Kirk Jensen, Brent AtkinsonJC918 U.S. PTO
09/723718
11/28/00If a **CONTINUATION APPLICATION**, check appropriate box and supply the requisite information:☒ **Continuation** ☐ **Divisional** ☐ **Continuation-in-part (CIP)** of prior application No.: 09/459,553

Which is a:

☐ **Continuation** ☐ **Divisional** ☐ **Continuation-in-part (CIP)** of prior application No.: _____

Which is a:

☐ **Continuation** ☐ **Divisional** ☐ **Continuation-in-part (CIP)** of prior application No.: _____

Enclosed are:

Application Elements

1. ☒ Filing fee as calculated and transmitted as described below
2. ☒ Specification having 98 pages and including the following:
 - a. ☒ Descriptive Title of the Invention
 - b. ☒ Cross References to Related Applications *(if applicable)*
 - c. ☐ Statement Regarding Federally-sponsored Research/Development *(if applicable)*
 - d. ☐ Reference to Microfiche Appendix *(if applicable)*
 - e. ☒ Background of the Invention
 - f. ☒ Brief Summary of the Invention
 - g. ☒ Brief Description of the Drawings *(if drawings filed)*
 - h. ☒ Detailed Description
 - i. ☒ Claim(s) as Classified Below
 - j. ☒ Abstract of the Disclosure

UTILITY PATENT APPLICATION TRANSMITTAL
(Large Entity)

(Only for new nonprovisional applications under 37 CFR 1.53(b))

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NEX 10-6

Total Pages in this Submission
3

Application Elements (Continued)

3. ☒ Drawing(s) *(when necessary as prescribed by 35 USC 113)*
- a. ☐ Formal Number of Sheets _____
- b. ☒ Informal Number of Sheets 35
4. ☐ Oath or Declaration
- a. ☐ Newly executed *(original or copy)* ☐ Unexecuted
- b. ☒ Copy from a prior application (37 CFR 1.63(d)) *(for continuation/divisional application only)*
- c. ☒ With Power of Attorney ☐ Without Power of Attorney
- d. ☐ DELETION OF INVENTOR(S)
Signed statement attached deleting inventor(s) named in the prior application,
see 37 C.F.R. 1.63(d)(2) and 1.33(b).
5. ☒ Incorporation By Reference *(usable if Box 4b is checked)*
The entire disclosure of the prior application, from which a copy of the oath or declaration is supplied under
Box 4b, is considered as being part of the disclosure of the accompanying application and is hereby
incorporated by reference therein.
6. ☐ Computer Program in Microfiche *(Appendix)*
7. ☒ Nucleotide and/or Amino Acid Sequence Submission *(if applicable, all must be included)*
- a. ☒ Paper Copy
- b. ☐ Computer Readable Copy *(identical to computer copy)*
- c. ☐ Statement Verifying Identical Paper and Computer Readable Copy

Accompanying Application Parts

8. ☐ Assignment Papers *(cover sheet & document(s))*
9. ☐ 37 CFR 3.73(B) Statement *(when there is an assignee)*
10. ☐ English Translation Document *(if applicable)*
11. ☐ Information Disclosure Statement/PTO-1449 ☐ Copies of IDS Citations
12. ☒ Preliminary Amendment
13. ☒ Acknowledgment postcard
14. ☒ Certificate of Mailing
- ☐ First Class ☒ Express Mail *(Specify Label No.):* EL652339515US

UTILITY PATENT APPLICATION TRANSMITTAL (Large Entity)

(Only for new nonprovisional applications under 37 CFR 1.53(b))

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3

Accompanying Application Parts (Continued)

15. ☐ Certified Copy of Priority Document(s) (if foreign priority is claimed)
16. ☒ Additional Enclosures (please identify below):

Request to Transfer Previously filed Sequence Information

Fee Calculation and Transmittal

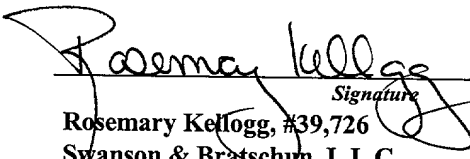
CLAIMS AS FILED

For	#Filed	#Allowed	#Extra	Rate	Fee
Total Claims		- 20 =	0	x \$18.00	\$0.00
Indep. Claims		- 3 =	0	x \$78.00	\$0.00
Multiple Dependent Claims (check if applicable) <input type="checkbox"/>					\$0.00
BASIC FEE					\$690.00
OTHER FEE (specify purpose)					\$0.00
TOTAL FILING FEE					\$690.00

- ☐ A check in the amount of _____ to cover the filing fee is enclosed.
- ☒ The Commissioner is hereby authorized to charge and credit Deposit Account No. 50-1643 as described below. A duplicate copy of this sheet is enclosed.
- ☒ Charge the amount of \$690.00 as filing fee.
- ☒ Credit any overpayment.
- ☐ Charge any additional filing fees required under 37 C.F.R. 1.16 and 1.17.
- ☐ Charge the issue fee set in 37 C.F.R. 1.18 at the mailing of the Notice of Allowance, pursuant to 37 C.F.R. 1.311(b).

Dated: November 28, 2000

cc: Vanessa Appleby


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IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

APPLICANT:	GOLD <i>et al.</i>	}	EXAMINER: ART UNIT:
SERIAL NO.:	09/		
FILED:	NOVEMBER 28, 2000		
FOR:	SYSTEMATIC EVOLUTION OF LIGANDS BY EXPONENTIAL ENRICHMENT: PHOTOSELECTION OF NUCLEIC ACID LIGANDS AND SOLUTION SELEX		

Assistant Commissioner for Patents
Washington, D.C. 20231

PRELIMINARY AMENDMENT

Dear Sir:

This application is a continuation of U.S. Patent Application Serial No. 09/459,553, filed December 13, 1999, entitled "Systematic Evolution of Ligands by Exponential Enrichment: Photoselection of Nucleic Acid Ligands and Solution SELEX."

Please enter the following amendments in the above-identified patent application.

In the Specification

At page 1, delete lines 6-25 and insert the following:

--RELATED APPLICATIONS

This application is a continuation of U.S. Patent Application Serial No. 09/459,553, filed December 13, 1999, entitled "Systematic Evolution of Ligands by Exponential Enrichment: Photoselection of Nucleic Acid Ligands and Solution SELEX."--

At page 2, line 22, after "Ligands," insert --issued as U.S. Patent No. 5,475,096--.

At page 2, line 24, after "entitled" insert --Methods for Identifying--.

At page 5, line 9, delete "Wilbert" and insert --Gilbert--

At page 10, line 26, delete "Eggens" and insert --Eggen--.

At page 11, line 8, after "Structure," insert --now abandoned (See, U.S. Patent No. 5,707,796)--.

09/459,553

At page 15, line 23, delete "SELEXES" and insert --SELEX--.

At page 19, line 15, delete "(A)" and insert --(Figure 18A)--.

At page 19, line 16, delete "(B)" and insert --(Figure 18B)--.

At page 19, line 20, delete "(A)" and insert --(Figure 19A)--.

At page 19, line 20, please delete "(B)" and insert --(Figure 19B)--.

At page 21, line 31, after "1991" insert --, now U.S. Patent No. 5,475,096--.

At page 25, line 4, delete "5'-iodo-uracyl" and insert --5'-iodo-uracil--.

At page 27, line 18, delete "caes" and insert --cases--.

At page 33, line 1, delete "protein" and insert --proteins--.

At page 37, line 18, after "and" delete "a".

At page 38, line 23, delete "Ld-XL" and insert --LD-XL--.

At page 39, line 17, delete "cystein" and insert --cysteine--.

At page 39, line 17, delete "possible" and insert --possibly--.

At page 39, line 24, after "extract" insert --were mixed--.

At page 41, line 31, delete "the" and insert --that--.

At page 42, line 2, delete "35 ul of a 100 ul" and insert --35 μ l of a 100 μ l--.

At page 42, line 3, delete "ul/ml" and insert -- μ g/ml--.

At page 42, line 11, delete "indicted" and insert --indicated--.

At page 42, line 23, delete "min" and insert --min.--.

At page 42, line 27, delete "and" and insert --and--.

At page 42, line 28, delete "or" and insert --of--.

At page 45, lines 18-19, delete "in press" and insert --32:10254--.

At page 50, line 21, delete "KCL" and insert --KCl--.

At page 52, line 12, delete "KCL" and insert --KCl--.

At page 58, line 12, delete the hard return at the end of the line.

At page 62, line 6, delete "radio-labelled" and insert --radiolabelled--.

At page 65, line 26, delete "37°C., phenol/ChCl₃" and insert --37°C, phenol/CHCl₃--.

At page 65, line 28, after "hours" delete "and".

At page 66, line 14, delete "ug" and insert --ug--.

At page 66, line 16, delete "polyacrylamide-/8" and insert --polyacrylamide/8--.

At page 66, line 23, delete "15" and insert --Fifteen--.

At page 66, line 26, delete "um" and insert --μm--.

At page 67, line 4, delete "uM" and insert --μM--.

At page 67, line 13, delete "ug" and insert --μg--.

At page 67, line 13, delete "90 C" and insert --90° C--.

At page 70, line 34, delete "ul" and insert --μl--.

At page 72, line 12, delete "Aigen" and insert --Aiken--.

Please delete originally filed pages 76-89 and replace them with the attached Sequence Listing.

In the Claims

Please cancel claims 1-10 and 13-28.

11. (amended) Nucleic acid ligands able to crosslink [the] a target molecule [produced by the method of claim 1] comprising one or more photoreactive groups.

12. (amended) The nucleic acid ligands of claim 11 further comprising one or more of the photoreactive groups selected from the group [set forth in claim 3] consisting of 5-bromouracil, 5-iodouracil, 5-bromovinyluracil, 5-iodovinyluracil, 5-azidouracil, 4-thiouracil, 5-bromocytosine, 5-iodocytosine, 5-bromovinylcytosine, 5-iodovinylcytosine, 5-azidocytosine, 8-azidoadenine, 8-bromoadenine, 8-iodoadenine, 8-azidoguanine, 8-bromoguanine, 8-iodoguanine, 8-azidohypoxanthine, 8-bromohypoxanthine, 8-iodohypoxanthine, 8-azidoxanthine, 8-bromoxanthine, 8-iodoxanthine, 5-bromodeoxyuracil, 8-bromo-2'-deoxyadenine, 5-iodo-2'-deoxyuracil, 5-iodo-2'-deoxycytosine, 5-[(4-azidophenacyl)thio]cytosine, 5-[(4-azidophenacyl)thio]uracil, 7-deaza-7-iodoadenine, 7-deaza-7-iodoguanine, 7-deaza-7-bromoadenine and 7-deaza-7-bromoguanine.

After entry of this Amendment, claims 11 and 12 are pending. The amendments made are supported by the specification and do not constitute new matter.

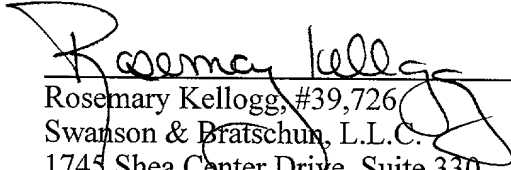
Attached is a copy of the Abstract of the invention printed on a separate sheet of paper.

Favorable consideration is earnestly solicited.

The Examiner is hereby encouraged to telephone the undersigned should there be any questions or concerns regarding this amendment.

Respectfully submitted,

November 28, 2000


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This invention relates, in part, to a method for selecting nucleic acid ligands which bind and/or photocrosslink to and/or photoinactivate a target molecule. The target molecule may be a protein, pathogen or toxic substance, or any biological effector. The nucleic acid ligands of the present invention contain photoreactive or chemically reactive groups and are

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useful, inter alia, for the diagnosis and/or treatment of diseases or pathological or toxic states.

The underlying method utilized in this invention is termed SELEX, an acronym for Systematic Evolution of
5 Ligands by EXponential enrichment. An improvement of the SELEX method herein described, termed Solution SELEX, allows more efficient partitioning between oligonucleotides having high and low affinity for a target molecule. An improvement of the high affinity
10 nucleic acid products of SELEX are useful for any purpose to which a binding reaction may be put, for example in assay methods, diagnostic procedures, cell sorting, as inhibitors of target molecule function, as therapeutic agents, as probes, as sequestering agents and the like.

15

BACKGROUND OF THE INVENTION

The SELEX method (hereinafter termed SELEX), described in U.S. Patent Application Serial No. 07/536,428, filed June 11, 1990, entitled Systematic
20 Evolution of Ligands By Exponential Enrichment, now abandoned, U.S. Patent Application Serial No. 07/714,131, filed June 10, 1991, entitled Nucleic Acid Ligands, and U.S. Patent Application Serial No. 07/931,473, filed August 17, 1992, entitled Nucleic Acid Ligands, issued as
25 U.S. Patent No. 5,270,163, all of which are herein specifically incorporated by reference (referred to herein as the SELEX Patent Applications), provides a class of products which are nucleic acid molecules, each having a unique sequence, each of which has the property
30 of binding specifically to a desired target compound or molecule. Each nucleic acid molecule is a specific ligand of a given target compound or molecule. SELEX is based on the unique insight that nucleic acids have sufficient capacity for forming a variety of two- and
35 three-dimensional structures and sufficient chemical

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versatility available within their monomers to act as ligands (form specific binding pairs) with virtually any chemical compound, whether monomeric or polymeric. Molecules of any size can serve as targets.

5 The SELEX method involves selection from a mixture of candidates and step-wise iterations of structural improvement, using the same general selection theme, to achieve virtually any desired criterion of binding affinity and selectivity. Starting from a mixture of
10 nucleic acids, preferably comprising a segment of randomized sequence, the method includes steps of contacting the mixture with the target under conditions favorable for binding, partitioning unbound nucleic acids from those nucleic acids which have bound to target
15 molecules, dissociating the nucleic acid-target pairs, amplifying the nucleic acids dissociated from the nucleic acid-target pairs to yield a ligand-enriched mixture of nucleic acids, then reiterating the steps of binding, partitioning, dissociating and amplifying through as many
20 cycles as desired.

 While not bound by theory, SELEX is based on the inventors' insight that within a nucleic acid_mixture containing a large number of possible sequences and structures there is a wide range of binding affinities
25 for a given target. A nucleic acid mixture comprising, for example a 20 nucleotide randomized segment can have 4^{20} candidate possibilities. Those which have the higher affinity constants for the target are most likely to bind to the target. After partitioning, dissociation and
30 amplification, a second nucleic acid mixture is generated, enriched for the higher binding affinity candidates. Additional rounds of selection progressively favor the best ligands until the resulting nucleic acid mixture is predominantly composed of only one or a few
35 sequences. These can then be cloned, sequenced and

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individually tested for binding affinity as pure ligands.

Cycles of selection, partition and amplification are repeated until a desired goal is achieved. In the most general case, selection/partition/amplification is continued until no significant improvement in binding strength is achieved on repetition of the cycle. The method may be used to sample as many as about 10^{18} different nucleic acid species. The nucleic acids of the test mixture preferably include a randomized sequence portion as well as conserved sequences necessary for efficient amplification. Nucleic acid sequence variants can be produced in a number of ways including synthesis of randomized nucleic acid sequences and size selection from randomly cleaved cellular nucleic acids. The variable sequence portion may contain fully or partially random sequence; it may also contain subportions of conserved sequence incorporated with randomized sequence. Sequence variation in test nucleic acids can be introduced or increased by mutagenesis before or during the selection/partition/amplification iterations.

Photocrosslinking of nucleic acids to proteins has been achieved through incorporation of photoreactive functional groups in the nucleic acid. Photoreactive groups which have been incorporated into nucleic acids for the purpose of photocrosslinking the nucleic acid to an associated protein include 5-bromouracil, 4-thiouracil, 5-azidouracil, and 8-azidoadenine (see Fig. 1).

Bromouracil has been incorporated into both DNA and RNA by substitution of bromodeoxyuracil (BrdU) and bromouracil (BrU) for thymine and uracil, respectively. BrU-RNA has been prepared with 5-bromouridine triphosphate in place of uracil using T7 RNA polymerase and a DNA template, and both BrU-RNA and BrdU-DNA have been prepared with 5-bromouracil and 5-bromodeoxyuracil

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phosphoramidites, respectively, in standard nucleic acid synthesis (Talbot et al. (1990) Nucleic Acids Res. 18:3521). Some examples of the photocrosslinking of BrdU-substituted DNA to associated proteins are as follows: BrdU-substituted DNA to proteins in intact cells (Weintraub (1973) Cold Spring Harbor Symp. Quant. Biol. 38:247); BrdU-substituted lac operator DNA to lac repressor (Lin and Riggs (1974) Proc. Natl. Acad. Sci. U.S.A. 71:947; Ogata and Wilbert (1977) Proc. Natl. Acad. Sci. U.S.A. 74:4973; Barbier et al. (1984) Biochemistry 23:2933; Wick and Matthews (1991) J. Biol. Chem. 266:6106); BrdU-substituted DNA to EcoRI and EcoRV restriction endonucleases (Wolfes et al. (1986) Eur. J. Biochem. 159:267); *Escherichia coli* BrdU-substituted DNA to cyclic adenosine 3',5'-monophosphate receptor protein (Katouzian-Safadi et al. (1991) Photochem. Photobiol. 53:611); BrdU-substituted DNA oligonucleotide of human polyomavirus to proteins from human fetal brain extract (Khalili et al. (1988) EMBO J. 7:1205); a yeast BrdU-substituted DNA oligonucleotide to GCN4, a yeast transcriptional activator (Blatter et al. (1992) Nature 359:650); and a BrdU-substituted DNA oligonucleotide of Methanosarcina sp CHT155 to the chromosomal protein Mcl (Katouzian-Safadi et al. (1991) Nucleic Acids Res. 19:4937). Photocrosslinking of BrU-substituted RNA to associated proteins has also been reported: BrU-substituted yeast precursor tRNA^{Phe} to yeast tRNA ligase (Tanner et al. (1988) Biochemistry 27:8852) and a BrU-substituted hairpin RNA of the R17 bacteriophage genome to R17 coat protein (Gott et al. (1991) Biochemistry 30:6290).

4-Thiouracil-substituted RNA has been used to photocrosslink, especially, t-RNA's to various associated proteins (Favre (1990) in: *Bioorganic Photochemistry*, Volume 1: Photochemistry and the Nucleic Acids, H.

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Morrison (ed.), John Wiley & Sons: New York, pp. 379-425; Tanner et al. (1988) *supra*). 4-Thiouracil has been incorporated into RNA using 4-thiouridine triphosphate and T7 RNA polymerase or using nucleic acid synthesis with the appropriate phosphoramidite; it has also been incorporated directly into RNA by exchange of the amino group of cytosine for a thiol group with hydrogen sulfide. Yet another method of site specific incorporation of photoreactive groups into nucleic acids involves use of 4-thiouridylyl-(3'-5')-guanosine (Wyatt et al. (1992) *Genes & Development* 6:2542).

Examples of 5-azidouracil-substituted and 8-azidoadenine-substituted nucleic acid photocrosslinking to associated proteins are also known. Associated proteins that have been crosslinked include terminal deoxynucleotidyl transferase (Evans et al. (1989) *Biochemistry* 28:713; Farrar et al. (1991) *Biochemistry* 30:3075); *Xenopus* TFIIIA, a zinc finger protein (Lee et al. (1991) *J. Biol. Chem.* 266:16478); and *E. coli* ribosomal proteins (Wower et al. (1988) *Biochemistry* 27:8114). 5-Azidouracil and 8-azidoadenine have been incorporated into DNA using DNA polymerase or terminal transferase. Proteins have also been photochemically labelled by exciting 8-azidoadenosine 3',5'-biphosphate bound to bovine pancreatic ribonuclease A (Wower et al. (1989) *Biochemistry* 28:1563) and 8-azidoadenosine 5'-triphosphate bound to ribulose-bisphosphate carboxylase/oxygenase (Salvucci and Haley (1990) *Planta* 181:287).

8-Bromo-2'-deoxyadenosine as a potential photoreactive group has been incorporated into DNA via the phosphoramidite (Liu and Verdine (1992) *Tetrahedron Lett.* 33:4265). The photochemical reactivity has yet to be investigated.

Photocrosslinking of 5-iodouracil-substituted

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nucleic acids to associated proteins has not been previously investigated, probably because the size of the iodo group has been thought to preclude specific binding of the nucleic acid to the protein of interest. However, 5-iodo-2'-deoxyuracil and 5-iodo-2'-deoxyuridine triphosphate have been shown to undergo photocoupling to thymidine kinase from *E. coli* (Chen and Prusoff (1977) Biochemistry 16:3310).

Mechanistic studies of the photochemical reactivity of the 5-bromouracil chromophore have been reported including studies with regard to photocrosslinking. Most importantly, BrU shows wavelength dependent photochemistry. Irradiation in the region of 310 nm populates an n, π^* singlet state which decays to ground state and intersystem crosses to the lowest energy triplet state (Dietz et al. (1987) J. Am. Chem. Soc. 109:1793), most likely the π, π^* triplet (Rothman and Kearns (1967) Photochem. Photobiol. 6:775). The triplet state reacts with electron-rich amino acid residues via initial electron transfer followed by covalent bond formation. Photocrosslinking of triplet 5-bromouracil to the electron rich aromatic amino acid residues tyrosine, tryptophan and histidine (Ito et al. (1980) J. Am. Chem. Soc. 102:7535; Dietz and Koch (1987) Photochem. Photobiol. 46:971), and the disulfide bearing amino acid, cystine (Dietz and Koch (1989) Photochem. Photobiol. 49:121), has been demonstrated in model studies. Even the peptide linkage is a potential functional group for photocrosslinking to triplet BrU (Dietz et al. (1987) *supra*). Wavelengths somewhat shorter than 308 nm populate both the n, π^* and π, π^* singlet states. The π, π^* singlet undergoes carbon-bromine bond homolysis as well as intersystem crossing to the triplet manifold (Dietz et al. (1987) *supra*); intersystem crossing may occur in part via internal conversion to the n, π^* singlet state.

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Carbon-bromine bond homolysis likely leads to nucleic acid strand breaks (Hutchinson and Köhnlein (1980) *Prog. Subcell. Biol.* 7:1; Shetlar (1980) *Photochem. Photobiol. Rev.* 5:105; Saito and Sugiyama (1990) in: *Bioorganic Photochemistry, Volume 1: Photochemistry and the Nucleic Acids*, H. Morrison, ed., John Wiley and Sons, New York, pp. 317-378). The wavelength dependent photochemistry is outlined in the Jablonski Diagram in Figure 2 and the model photocrosslinking reactions are shown in Figure 3.

5 The location of photocrosslinks from irradiation of some BrU-substituted nucleoprotein complexes have been investigated. In the lac repressor-BrdU-lac operator complex a crosslink to tyrosine-17 has been established (Allen et al. (1991) *J. Biol. Chem.* 266:6113). In the
10 archaeobacterial chromosomal protein MCl-BrdU-DNA complex a crosslink to tryptophan-74 has been implicated. In yeast BrdU-substituted DNA-GCN4 yeast transcriptional activator a crosslink to alanine-238 was reported (Blatter et al. (1992) *supra*). In this latter example
15 the nucleoprotein complex was irradiated at 254 nm which populated initially the π, π^* singlet state.

The results of some reactivity and mechanistic studies of 5-iodouracil, 5-iodo-2'-deoxyuracil, 5-iodo-2'-deoxyuracil-substituted DNA, and 5-iodo-2'-
20 deoxycytosine-substituted DNA have been reported. 5-Iodouracil and 5-iodo-2'-deoxyuracil couple at the 5-position to allylsilanes upon irradiation in acetonitrile-water bearing excess silane with emission from a medium pressure mercury lamp filtered through
25 Pyrex glass; the mechanism was proposed to proceed through initial carbon-iodine bond homolysis followed by radical addition to the π -bond of the allylsilane (Saito et al. (1986) *J. Org. Chem.* 51:5148).

Aerobic and anaerobic photo-deiodination of 5-iodo-
35 2'-deoxyuracil-substituted DNA has been studied as a

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function of excitation wavelength; the intrinsic quantum yield drops by a factor of 4 with irradiation in the region of 313 nm relative to the quantum yield with irradiation in the region of 240 nm. At all wavelengths the mechanism is proposed to involve initial carbon-iodine bond homolysis (Rahn and Sellin (1982) Photochem. Photobiol. 35:459). Similarly, carbon-iodine bond homolysis is proposed to occur upon irradiation of 5-iodo-2'-deoxycytidine-substituted DNA at 313 nm (Rahn and Stafford (1979) Photochem. Photobiol. 30:449). Strictly monochromatic light was not used in any of these studies. Recently, a 5-iodouracil-substituted duplex DNA was shown to undergo a photochemical single strand break (Sugiyama et al. (1993) J. Am. Chem. Soc. 115:4443).

Also of importance with respect to the present invention is the observed direct population of the triplet states of 5-bromouracil and 5-iodouracil from irradiation of the respective $S_0 \rightarrow T$ absorption bands in the region of 350-400 nm (Rothman and Kearns (1967) *supra*).

Photophysical studies of the 4-thiouracil chromophore implicate the π, π^* triplet state as the reactive state. The intersystem crossing quantum yield is unity or close to unity. Although photocrosslinking within 4-thiouracil-substituted nucleoprotein complexes has been observed, amino acid residues reactive with excited 4-thiouracil have not been established (Favre (1990) *supra*). The addition of the α -amino group of lysine to excited 4-thiouracil at the 6-position has been reported; however, this reaction is not expected to be important in photocrosslinking within nucleoprotein complexes because the α -amino group is involved in a peptide bond (Ito et al. (1980) Photochem. Photobiol. 32:683).

Photocrosslinking of azide-bearing nucleotides or

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nucleic acids to associated proteins is thought to proceed via formation of the singlet and/or triplet nitrene (Bayley and Knowles (1977) *Methods Enzymol.* 46:69; Czarnecki et al. (1979) *Methods Enzymol.* 56:642; Hanna et al. (1993) *Nucleic Acids Res.* 21:2073). Covalent bond formation results from insertion of the nitrene in an O-H, N-H, S-H or C-H bond. Singlet nitrenes preferentially insert in heteroatom-H bonds and triplet nitrenes in C-H bonds. Singlet nitrenes can also rearrange to azirines which are prone to nucleophilic addition reactions. If a nucleophilic site of a protein is adjacent, crosslinking can also occur via this pathway. A potential problem with the use of an azide functional group results if it resides ortho to a ring nitrogen; the azide will exist in equilibrium with a tetrazole which is much less photoreactive.

The coat protein-RNA hairpin complex of the R17 bacteriophage is an ideal system for the study of nucleic acid-protein photocrosslinking because of the simplicity of the system *in vitro*. The system is well characterized, consisting of a viral coat protein that binds with high affinity to an RNA hairpin within the phage genome. *In vivo* the interaction of the coat protein with the RNA hairpin plays two roles during phage infection: the coat protein acts as a translational repressor of replicase synthesis (Eggens and Nathans (1969) *J. Mol. Biol.* 39:293), and the complex serves as a nucleation site for encapsidation (Ling et al. (1970) *Virology* 40:920; Beckett et al. (1988) *J. Mol. Biol.* 204:939). Many variations of the wild-type hairpin sequence also bind to the coat protein with high affinity (Tuerk & Gold (1990) *Science* 249:505; Gott et al. (1991) *Biochemistry* 30:6290; Schneider et al. (1992) *J. Mol. Biol.* 228:862).

The selection of nucleic acid ligands according to

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the SELEX method may be accomplished in a variety of ways, such as on the basis of physical characteristics. Selection on the basis of physical characteristics may include physical structure, electrophoretic mobility, solubility, and partitioning behavior. U. S. patent application Serial No. 07/960,093, filed October 14, 1992, entitled Method for Selecting Nucleic Acids on the Basis of Structure, herein specifically incorporated by reference, describes the selection of nucleic acid sequences on the basis of specific electrophoretic behavior. The SELEX technology may also be used in conjunction with other selection techniques, such as HPLC, column chromatography, chromatographic methods in general, solubility in a particular solvent, or partitioning between two phases.

BRIEF SUMMARY OF THE INVENTION

In one embodiment, the present invention includes a method for selecting and identifying nucleic acid ligands from a candidate mixture of randomized nucleic acid sequences on the basis of the ability of the randomized nucleic acid sequences to bind and/or photocrosslink to a target molecule. This embodiment is termed Covalent SELEX generally, and PhotoSELEX specifically when irradiation is required to form covalent linkage between the nucleic acid ligand and the target.

In one variation of this embodiment, the method comprises preparing a candidate mixture of nucleic acid sequences which contain photoreactive groups; contacting the candidate mixture with a target molecule wherein nucleic acid sequences having increased affinity to the target molecule bind the target molecule, forming nucleic acid-target molecule complexes; irradiating the nucleic acid-target molecule mixture, wherein some nucleic acids incorporated in nucleic acid-target molecule complexes

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crosslink to the target molecule via the photoreactive functional groups; taking advantage of the covalent bond to partition the crosslinked nucleic acid-target molecule complexes from free nucleic acids in the candidate mixture; and identifying the nucleic acid sequences that were photocrosslinked to the target molecule. The process can further include the iterative step of amplifying the nucleic acids that photocrosslinked to the target molecule to yield a mixture of nucleic acids enriched in sequences that are able to photocrosslink to the target molecule.

In another variation of this embodiment of the present invention, nucleic acid ligands to a target molecule selected through SELEX are further selected for their ability to crosslink to the target. Nucleic acid ligands to a target molecule not containing photoreactive groups are initially identified through the SELEX method. Photoreactive groups are then incorporated into these selected nucleic acid ligands, and the ligands contacted with the target molecule. The nucleic acid-target molecule complexes are irradiated and those able to photocrosslink to the target molecule identified.

In another variation of this embodiment of the present invention, photoreactive groups are incorporated into all possible positions in the nucleic acid sequences of the candidate mixture. For example, 5-iodouracil and 5-iodocytosine may be substituted at all uracil and cytosine positions. The first selection round is performed with irradiation of the nucleic acid-target molecule complexes such that selection occurs for those nucleic acid sequences able to photocrosslink to the target molecule. Then SELEX is performed with the nucleic acid sequences able to photocrosslink to the target molecule to select crosslinking sequences best able to bind the target molecule.

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In another variation of this embodiment of the present invention, nucleic acid sequences containing photoreactive groups are selected through SELEX for a number of rounds in the absence of irradiation, resulting in a candidate mixture with a partially enhanced affinity for the target molecule. PhotoSELEX is then conducted with irradiation to select ligands able to photocrosslink to the target molecule.

In another variation of this embodiment of the present invention, SELEX is carried out to completion with nucleic acid sequences not containing photoreactive groups, and nucleic acid ligands to the target molecule selected. Based on the sequences of the selected ligands, a family of related nucleic acid sequences is generated which contain a single photoreactive group at each nucleotide position. PhotoSELEX is performed to select a nucleic acid ligand capable of photocrosslinking to the target molecule.

In a further variation of this embodiment of the present invention, a nucleic acid ligand capable of modifying the bioactivity of a target molecule through binding and/or crosslinking to a target molecule is selected through SELEX, photoSELEX, or a combination of these methods.

In a further variation of this embodiment of the present invention, a nucleic acid ligand to a unique target molecule associated with a specific disease process is identified. In yet another variation of this embodiment of the present invention, a nucleic acid ligand to a target molecule associated with a disease state is used to treat the disease in vivo.

The present invention further encompasses nucleic acid sequences containing photoreactive groups. The nucleic acid sequences may contain single or multiple photoreactive groups. Further, the photoreactive groups

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may be the same or different in a single nucleic acid sequence. The photoreactive groups incorporated into the nucleic acids of the invention include any chemical group capable of forming a crosslink with a target molecule upon irradiation. Although in some cases irradiation may not be necessary for crosslinking to occur.

The nucleic acids of the present invention include single- and double-stranded RNA and single- and double-stranded DNA. The nucleic acids of the present invention may contain modified groups such as 2'-amino (2'-NH₂) or 2'-fluoro (2'-F)-modified nucleotides. The nucleic acids of the present invention may further include backbone modifications.

The present invention further includes the method whereby candidate mixtures containing modified nucleic acids are prepared and utilized in the SELEX process, and nucleic acid ligands are identified that bind or crosslink to the target species. In one example of this embodiment, the candidate mixture is comprised of nucleic acids wherein all uracil residues are replaced by 5-halogenated uracil residues, and nucleic acid ligands are identified that form covalent attachments to the selected target.

An additional embodiment of the present invention, termed solution SELEX, presents several improved methods for partitioning between ligands having high and low affinity nucleic acid-target complexes is achieved in solution and without, or prior to, use of a partitioning matrix. Generally, a central theme of the method of solution SELEX is that the nucleic acid candidate mixture is treated in solution and results in preferential amplification during PCR of the highest affinity nucleic acid ligands or catalytic RNAs. The solution SELEX method achieves partitioning between high and low affinity nucleic acid-target complexes through a number

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of methods, including (1) Primer extension inhibition which results in differentiable cDNA products such that the highest affinity ligands may be selectively amplified during PCR. Primer extension inhibition is achieved with the use of nucleic acid polymerases, including DNA or RNA polymerases, reverse transcriptase, and Q β -replicase. (2) Exonuclease hydrolysis inhibition which also results in only the highest affinity ligands amplifying during PCR. This is achieved with the use of any 3' \rightarrow 5' double-stranded exonuclease. (3) Linear to circle formation to generate differentiable cDNA molecules resulting in amplification of only the highest affinity ligands during PCR.

In one embodiment of the solution SELEX method, synthesis of cDNAs corresponding to low affinity oligonucleotides are preferentially blocked and thus rendered non-amplifiable by PCR. In another embodiment, low affinity oligonucleotides are preferentially removed by affinity column chromatography prior to PCR amplification. Alternatively, high affinity oligonucleotides may be preferentially removed by affinity column chromatography. In yet another embodiment of the SELEXES method, cDNAs corresponding to high affinity oligonucleotides are preferentially rendered resistant to nuclease enzyme digestion. In a further embodiment, cDNAs corresponding to low affinity oligonucleotides are rendered preferentially enzymatically or chemically degradable.

Solution SELEX is an improvement over prior art partitioning schemes. With the method of the present invention, partitioning is achieved without inadvertently also selecting ligands that only have affinity for the partitioning matrix, the speed and accuracy of partitioning is increased, and the procedure may be readily automated.

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The present disclosure provides non-limiting examples which are illustrative and exemplary of the invention. Other partitioning schemes and methods of selecting nucleic acid ligands through binding and photocrosslinking to a target molecule will be become apparent to one skilled in the art from the present disclosure.

BRIEF DESCRIPTION OF THE FIGURES

FIGURE 1 shows structures of photoreactive chromophores which have been incorporated into nucleic acids.

FIGURE 2 shows a Jablonski energy level diagram for the 5-bromouracil chromophore and the reactivity of the various excited states.

FIGURE 3 shows the model reactions for photocrosslinking of the 5-bromouracil chromophore to amino acid residues such as tyrosine, tryptophan, histidine, and cystine.

FIGURE 4 compares UV absorption by thymidine, 5-bromouracil, 5-iodouracil, and L-tryptophan in TMK pH 8.5 buffer (100 mM tris(hydroxymethyl)aminomethane hydrochloride, 10 mM magnesium acetate, and 80 mM potassium chloride). The emission wavelengths of the XeCl and HeCd lasers are also indicated. Of particular importance is absorption by 5-iodouracil at 325 nm without absorption by tryptophan or thymidine. The molar extinction coefficient for 5-iodouracil at 325 nm is 163 L/mol·cm.

FIGURE 5 shows structures of photoreactive chromophores which can be incorporated into randomized

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nucleic acid sequences.

FIGURE 6 shows the structures of hairpin RNA sequences RNA-1 (SEQ ID NO:1), RNA-2 (SEQ ID NO:2), and RNA-3 (SEQ ID NO:3) containing 5-bromouracil, 5-iodouracil, and uracil, respectively. These are variants of the wild-type hairpin of the R17 bacteriophage genome which bind tightly to the R17 coat protein.

FIGURE 7 shows binding curves for RNA-1 (SEQ ID NO:1), RNA-2 (SEQ ID NO:2), and RNA-3 (SEQ ID NO:3) to R17 coat protein. The binding constants calculated from the binding curves are also given.

FIGURE 8 shows the percent of RNA-1 (SEQ ID NO:1) and RNA-2 (SEQ ID NO:2) photocrosslinked to R17 coat protein with monochromatic irradiation at 308 nm from a XeCl excimer laser as a function of time. Photocrosslinking of RNA-1 (SEQ ID NO:1) maximized at 40% because of competitive photodamage to the coat protein during the irradiation period. Less photodamage to coat protein occurred with RNA-2 (SEQ ID NO:2) because of the shorter irradiation time.

FIGURE 9 shows the percent of RNA-2 (SEQ ID NO:2) photocrosslinked to R17 coat protein with monochromatic irradiation at 325 nm from a HeCd laser as a function of time. The data are also presented in the original electrophoretic gel format. The symbol IU XL marks RNA crosslinked to protein. A near-quantitative yield of photocrosslinking was obtained.

FIGURE 10 shows the percent of RNA-1 (SEQ ID NO:1) and RNA-2 (SEQ ID NO:2) photocrosslinked to R17 coat protein with broad-band irradiation in the region of 312

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nm from a Transilluminator as a function of time. Less than quantitative yields of photocrosslinking were obtained because of photodamage to the protein and possibly to the RNA sequences.

5

FIGURE 11 shows formation of the same product, Structure 6, from irradiation at 308 nm of 5-iodouracil and 5-bromouracil in the presence of excess N-acetyltyrosine N-ethylamide (Structure 5).

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FIGURE 12 pictures photocrosslinking of RNA-7 (SEQ ID NO:4) to R17 coat protein with 308 nm light followed by enzymatic digestion of most of the coat protein.

15

FIGURE 13 shows formation of a complementary DNA from the RNA template after enzymatic digestion of the coat protein of Figure 12 (SEQ ID NO:4).

20

FIGURE 14 shows the polyacrylamide gel of Example 8 showing production of a cDNA from an RNA template bearing modified nucleotides as shown in Figures 12 and 13. The modified nucleotides were 5-iodouracil and uracil substituted at the 5-position with a small peptide. Based upon model studies shown in Figure 11, the peptide was most likely attached to the uracil via the phenolic ring of a tyrosine residue.

25

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FIGURE 15 shows the photocrosslinking of [α -³²P] GTP labelled pool RNA to HIV-1 Rev protein using tRNA competition.

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FIGURE 16 (SEQ ID NOS:5-55) shows the sequence of the previously identified RNA ligand to HIV-1 Rev protein that is referred to herein as 6a (SEQ ID NO:5). Also shown are 52 sequences from round 13 selected for

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photocrosslinking to HIV-1 Rev protein.

FIGURE 17 (SEQ ID NOS:56-57) shows the consensus for class 1 ligands and class 2 ligands. Class 1: Consensus secondary structure for class 1 and class 2 molecules. N_1-N_1' indicate 1-2 complementary base pairs; N_2-N_2' indicates 1-4 complementary base pairs, D-H' is an A-U, U-A, or G-C base pair; K-M' is a G-C or U-A base pair (16). Class 2: Bold sequences represent the highly conserved 10 nucleotides that characterize class 2 molecules; base-pairing is with the 5' fixed end of the molecule.

FIGURE 18 (SEQ ID NO:58) shows the sequence and predicted secondary structure of trunc2 (A). Also shown (B) is a gel demonstrating the specificity of trunc2 photocrosslinking to ARM proteins.

FIGURE 19 shows the sequence and predicted secondary structure of trunc24 (SEQ ID NO:59) (A). Also shown (B) is a gel demonstrating the specificity of laser independent crosslinking to ARM proteins.

FIGURE 20 shows the trunc24 (SEQ ID NO:59) photo-independent crosslinking with HIV-1 Rev in the presence of human nuclear extracts.

FIGURE 21 illustrates the cyclical relationship between SELEX steps. A single-stranded nucleic acid repertoire of candidate oligonucleotides is generated by established procedures on a nucleic acid synthesizer, and is amplified by PCR to generate a population of double-stranded DNA molecules. Candidate DNA or RNA molecules are generated through asymmetric PCR or transcription, respectively, purified, and allowed to complex with a

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target molecule. This is followed by partitioning of bound and unbound nucleic acids, synthesis of cDNA, and PCR amplification to generate double-stranded DNA.

5 FIGURE 22 illustrates one embodiment of the solution SELEX method in which primer extension inhibition is used to create differentiable cDNA pools - an amplifiable high affinity oligonucleotide cDNA pool and a non-amplifiable low affinity oligonucleotide cDNA pool. In this
10 embodiment, the first cDNA extension is performed in the presence of chain terminating nucleotides such as ddG. After removal of the target molecule and dideoxynucleotides, the second cDNA extension is
15 conducted in the presence of four dNTPs. Full-length cDNA is preferentially synthesized from the high affinity oligonucleotides and therefore, the high affinity cDNA pool is amplified in the subsequent PCR step.

20 FIGURE 23 illustrates the cyclic solution SELEX process for the embodiment shown in Figure 22.

25 FIGURE 24 illustrates one embodiment of the cyclic solution SELEX process wherein partitioning between oligonucleotides having high and low affinity to a target molecule is achieved by restriction enzyme digestion. In
30 this embodiment, the first cDNA extension is conducted with four dNTPs and results in cDNAs corresponding to the low affinity oligonucleotides. The target is then removed and a second cDNA extension is conducted in the presence of modified nucleotides resistant to enzymatic
35 cleavage. The cDNA pools are then incubated with restriction enzyme and only the cDNA pool corresponding to high affinity oligonucleotides is amplifiable in the subsequent PCR step.

FIGURE 25 illustrates one embodiment of the cyclic solution SELEX process wherein partitioning between oligonucleotides having high and low affinity to a target molecule is achieved by affinity chromatography. The first cDNA extension is performed in the presence of a modified nucleotide such as a biotinylated nucleotide, which allows the cDNA pool corresponding to the low-affinity oligonucleotide to be retained on an affinity column.

10

FIGURE 26 illustrates one embodiment of the solution SELEX process wherein partitioning between oligonucleotides having high and low affinity to a target molecule is achieved by exonuclease inhibition and results in formation of a double-stranded nucleic acid population with high affinity for the target molecule.

FIGURE 27 illustrates one embodiment of the solution
SELEX process wherein catalytic nucleic acids are
selected and isolated.

DETAILED DESCRIPTION OF THE INVENTION

The present invention includes a variation of the SELEX method for selecting nucleic acid ligands. This application hereby specifically incorporates by reference the full text including the definitions provided in the earlier SELEX patent applications, specifically those provided in United States Patent Application Serial No. 07/536,428, filed June 11, 1990, now abandoned, and 07/714,131, filed June 10, 1991. The method of one embodiment of the present invention, termed covalent SELEX or photoSELEX, identifies and selects nucleic acid ligands capable of binding and/or photocrosslinking to target molecules.

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This application also presents a method for improved partitioning of nucleic acid ligands identified through the SELEX method.

5 In its most basic form, the SELEX process may be defined by the following series of steps:

10 1) A candidate mixture of nucleic acids of differing sequence is prepared. The candidate mixture generally includes regions of fixed sequences (i.e., each of the members of the candidate mixture contains the same sequences in the same location) and regions of randomized sequences. The fixed sequence regions are selected either: a) to assist in the amplification steps described below; b) to mimic a sequence known to bind to the target; or c) to enhance the potential of a given
15 structural arrangement of the nucleic acids in the candidate mixture. The randomized sequences can be totally randomized (i.e., the probability of finding a base at any position being one in four) or only partially randomized (e.g., the probability of finding a base at
20 any location can be selected at any level between 0 and 100 percent).

2) The candidate mixture is contacted with the selected target under conditions favorable for binding between the target and members of the candidate mixture.
25 Under these circumstances, the interaction between the target and the nucleic acids of the candidate mixture can be considered as forming nucleic acid-target pairs between the target and the nucleic acids having the strongest affinity for the target.

30 3) The nucleic acids with the highest affinity for the target are partitioned from those nucleic acids with lesser affinity to the target. Because only an extremely small number of sequences (and possibly only one molecule of nucleic acid) corresponding to the highest affinity
35 nucleic acids exist in the candidate mixture, it is

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5 4) Those nucleic acids selected during partitioning as having the relatively higher affinity to the target are then amplified to create a new candidate mixture that is enriched in nucleic acids having a relatively higher affinity for the target.

The SELEX Patent Applications describe and elaborate on this process in great detail. Included are targets that can be used in the process; methods for the preparation of the initial candidate mixture; methods for partitioning nucleic acids within a candidate mixture; and methods for amplifying partitioned nucleic acids to generate enriched candidate mixtures. The SELEX Patent Applications also describe ligand solutions obtained to a number of target species, including both protein targets wherein the protein is and is not a nucleic acid binding protein.

Partitioning means any process whereby ligands bound to target molecules can be separated from nucleic acids not bound to target molecules. More broadly stated, partitioning allows for the separation of all the nucleic acids in a candidate mixture into at least two pools based on their relative affinity to the target molecule.

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Partitioning can be accomplished by various methods known in the art. Nucleic acid-protein pairs can be bound to nitrocellulose filters while unbound nucleic acids are not. Columns which specifically retain nucleic acid-target complexes can be used for partitioning. For example, oligonucleotides able to associate with a target molecule bound on a column allow use of column chromatography for separating and isolating the highest affinity nucleic acid ligands. Liquid-liquid partitioning can be used as well as filtration gel retardation, and density gradient centrifugation.

I. PhotoSELEX.

The present invention encompasses nucleic acid ligands which bind, photocrosslink and/or photoinactivate target molecules. Binding as referred to herein generally refers to the formation of a covalent bond between the ligand and the target, although such binding is not necessarily irreversible. Certain nucleic acid ligands of the present invention contain photoreactive groups which are capable of photocrosslinking to the target molecule upon irradiation with light. Additional nucleic acid ligands of the present invention are capable of bond formation with the target in the absence of irradiation.

In one embodiment, the present invention encompasses nucleic acid ligands which are single- or double-stranded RNA or DNA oligonucleotides. The nucleic acid ligands of the present invention may contain photoreactive groups capable of crosslinking to the selected target molecule when irradiated with light. Further, the present invention encompasses nucleic acid ligands containing any modification thereof. Reference to a photoreactive group herein may simply refer to a relatively simple modification to a natural nucleic acid residue that confers increased reactivity or photoreactivity to the

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nucleic acid residue. Such modifications include, but are not limited to, modifications at cytosine exocyclic amines, substitution with halogenated groups, e.g., 5'-bromo- or 5'-iodo-uracyl, modification at the 2'-
5 position, e.g., 2'-amino (2'-NH₂) and 2'-fluoro (2'-F), backbone modifications, methylations, unusual base-pairing combinations and the like. For example, the nucleic acid ligands of the present invention may include modified nucleotides such as 2'-NH₂-iodouracil, 2'-NH₂-
10 iodocytosine, 2'-NH₂-iodoadenine, 2'-NH₂-bromouracil, 2'-NH₂-bromocytosine, and 2'-NH₂-bromoadenine.

In one embodiment of the photoSELEX method of the present invention, a randomized set of nucleic acid sequences containing photoreactive groups, termed the
15 candidate mixture, is mixed with a quantity of the target molecule and allowed to establish an equilibrium binding with the target molecule. The nucleic acid-target molecule mixture is then irradiated with light until photocrosslinking is complete as indicated by
20 polyacrylamide gel electrophoresis. Only some of those nucleic acids binding tightly to the target molecules will efficiently crosslink with the target.

The candidate mixture of the present invention is comprised of nucleic acid sequences with randomized
25 regions including chemically reactive or a photoreactive group or groups. Preferably the reactive groups are modified nucleic acids. The nucleic acids of the candidate mixture preferably include a randomized sequence portion as well as conserved sequences necessary
30 for efficient amplification. The variable sequence portion may contain fully or partially random sequence; it may also contain subportions of conserved sequence incorporated within the randomized sequence regions.

Preferably, each oligonucleotide member of the
35 candidate mixture contains at least one chemically

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reactive or photoreactive group. Further, each oligonucleotide member of the candidate mixture may be partially or fully substituted at each position by modified nucleotides containing reactive groups. The candidate mixture may also be comprised of oligonucleotides containing more than one type of reactive group.

The target molecules bound and/or photocrosslinked by the nucleic acid ligands of the present invention are commonly proteins and are selected based upon their role in disease and/or toxicity. Examples are enzymes for which an inhibitor is desired or proteins for which detection is desired. However, the target molecule may be any compound of interest for which a ligand is desired. A target molecule can be a protein, peptide, carbohydrate, polysaccharide, glycoprotein, hormone, receptor, antigen, antibody, virus, substrate, metabolite, transition state analog, cofactor, inhibitor, drug, dye, nutrient, growth factor, etc., without limitation.

A photoreactive group for the purpose of this application is preferably a modified nucleotide that contains a photochromophore, and that is capable of photocrosslinking with a target species. Although referred to herein as photoreactive groups, in some cases as described below, irradiation is not necessary for covalent binding to occur between the nucleic acid ligand and the target. Preferentially, the photoreactive group will absorb light in a spectrum of the wavelength that is not absorbed by the target or the non-modified portions of the oligonucleotide. This invention encompasses, but is not limited to, oligonucleotides containing a photoreactive group selected from the following: 5-bromouracil (BrU), 5-iodouracil (IU), 5-bromovinyluracil, 5-iodovinyluracil, 5-azidouracil, 4-thiouracil, 5-

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bromocytosine, 5-iodocytosine, 5-bromovinylcytosine, 5-iodovinylcytosine, 5-azidocytosine, 8-azidoadenine, 8-bromoadenine, 8-iodoadenine, 8-azidoguanine, 8-bromoguanine, 8-iodoguanine, 8-azidohypoxanthine, 8-bromohypoxanthine, 8-iodohypoxanthine, 8-azidoxanthine, 8-bromoxanthine, 8-iodoxanthine, 5-bromodeoxyuridine, 8-bromo-2'-deoxyadenine, 5-iodo-2'-deoxyuracil, 5-iodo-2'-deoxycytosine, 5-[(4-azidophenacyl)thio]cytosine, 5-[(4-azidophenacyl)thio]uracil, 7-deaza-7-iodoadenine, 7-deaza-7-iodoguanine, 7-deaza-7-bromoadenine, and 7-deaza-7-bromoguanine (Fig. 5). In one embodiment, the photoreactive groups are 5-bromouracil (BrU) and 5-iodouracil (IU).

The photoreactive groups of the present invention are capable of forming bonds with the target species upon irradiation of an associated nucleic acid-target pair. The associated pair is referred to herein as a nucleoprotein complex, and in some cases irradiation is not required for bond formation to occur. The photocrosslink that typically occurs will be the formation of a covalent bond between the associated nucleic acid and the target. However, a tight ionic interaction between the nucleic acid and target may also occur upon irradiation.

In one embodiment, photocrosslinking occurs due to electromagnetic irradiation. Electromagnetic irradiation includes ultraviolet light, visible light, X-ray and gamma ray. 5-Halo substituted deoxyuracils and deoxycytosines are known to sensitize cells to ionizing radiation (Szybalski (1974) Cancer Chemother. Rep. 58:539).

Crosslinking experiments have shown that a precise juxtaposition of either IU or BrU and a tyrosine, tryptophan, or histidine is required for a high yield crosslinking to occur. The present invention takes

The randomized set of RNA oligonucleotides containing BrU or IU are mixed with a quantity of a target protein. The photoreactive chromophore is incorporated randomly into RNA as BrU or IU in place of uracil using standard methodology. The RNA-target protein mixture is irradiated with near ultraviolet light in the range of 300 to 325 nm until photocrosslinking is complete. Only those photoreactive groups adjacent to a reactive amino acid residue in a nucleoprotein complex form a covalent bond to the protein. Excited BrU or IU, returns to the ground state unless it is near a reactive functional group such as an oxidizable amino acid residue. Amino acid residues which have been established as being reactive with the lowest triplet state of 5-bromouracil include tyrosine, tryptophan, histidine, and cystine (see Fig. 3). Others of potential reactivity based upon mechanistic studies are phenylalanine, methionine, cysteine, lysine, and arginine.

Nucleoprotein complexes which do not form crosslinks may be easily disrupted by adjusting the reaction medium such as by denaturing with heat and/or salt. Nucleic acids covalently bound to the protein may be separated

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from free nucleic acids on a nitrocellulose filter or by other partitioning methods known to those skilled in the art. Alternate methods for separating nucleic acids covalently bound to targets from free nucleic acids include gel electrophoresis followed by electroelution, precipitation, differential solubility, and chromatography. To one skilled in the art, the method of choice will depend at least in part on the target molecule of interest. The crosslinked nucleic acids are released from the nitrocellulose filter by digestion of the protein material with enzymes such as Proteinase K. At this point 5-halouracil groups which have photocrosslinked to the target protein are bound to a single amino acid or to a short peptide. The read-through ability of reverse transcriptase is not effected by incorporation of a substituent at the 5-position of uracil because reverse transcriptase (RT) does not differentiate the 5-position of uracil from that of thymine. Derivatization of the 5-position has been used to incorporate groups as large as biotin into RNA molecules. In one embodiment of the present invention, the target is removed from the selected photocrosslinked nucleic acid by photo or chemical dissociation.

Complementary nucleic acid copies of the selected RNA sequences are prepared with an appropriate primer. The cDNA is amplified with a DNA polymerase and a second primer. 5-Halo-2'-deoxyuracil is not employed in the DNA synthesis and amplification steps. The amplified DNAs are then transcribed into RNA sequences using 5-halouridine triphosphate in place of uridine triphosphate in the same or different ratio of 5-halouridine to uridine in the candidate mixture.

For the subsequent round of photoSELEX, the partially selected RNA sequences are again allowed to complex with a quantity of the target protein.

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Subsequently, the nucleoprotein complexes are irradiated in the region of 300-325 nm. RNA sequences which have crosslinked to protein are again separated from RNA sequences which have not crosslinked. cDNAs are prepared and amplified and a third set of RNA sequences containing 5-halouracil are prepared. The cycle is continued until it converges to one or several RNA ligands which bind with high affinity and photocrosslink to the target protein. Shortening of the irradiation time in later cycles can further enhance the selection. The cDNAs of the selected RNA ligands are amplified, gel purified, and sequenced. Alternatively, the RNA sequences can be sequenced directly. The selected RNA sequences are then transcribed from the appropriate synthesized DNA template, again using 5-halouridine triphosphate in place of uridine triphosphate (Example 11).

In another embodiment of the present invention, photoSELEX is performed on oligonucleotide sequences preselected for their ability to bind the target molecule (Example 12). SELEX is initially performed with oligonucleotides which do not contain photoreactive groups. The RNA ligand is transformed into a photoreactive ligand by substitution of photoreactive nucleic acid nucleotides at specific sites in the ligand. The photochemically active permutations of the initial ligand may be developed through a number of approaches, such as specific substitution or partial random incorporation of the photoreactive nucleotides. Specific substitution involves the synthesis of a variety of oligonucleotides with the position of the photoreactive nucleotide changed manually. The location of the substitution is directed based upon the available data on binding of the ligand to the target molecule. For example, substitutions are made to the initial ligand in areas of the molecule that are known to interact with the

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target molecule. For subsequent selection rounds, the photoSELEX method is used to select for the ability to crosslink to the target molecule upon irradiation.

5 In another embodiment of the present invention, nucleic acid ligands are selected by photoSELEX followed by SELEX. PhotoSELEX is performed initially with oligonucleotide sequences containing photoreactive groups. Sequences selected for their ability to crosslink to the target molecule are then selected for
10 ability to bind the target molecule through the SELEX method (Example 13).

In another embodiment of the present invention, a limited selection of oligonucleotides through SELEX is followed by selection through photoSELEX (Example 14).
15 The initial SELEX selection rounds are conducted with oligonucleotides containing photoreactive groups. After a number of SELEX rounds, photoSELEX is conducted to select oligonucleotides capable of binding the target molecule.

20 In yet another embodiment of the present invention, nucleic acid ligands identified through SELEX are subjected to limited randomization, followed by selection through photoSELEX (Example 15). SELEX is first carried out to completion with nucleic acid sequences not
25 containing photoreactive groups. The sequence of the nucleic acid ligand is used to generate a family of oligonucleotides through limited randomization. PhotoSELEX is subsequently performed to select a nucleic acid ligand capable of photocrosslinking to the target
30 molecule.

In another embodiment of the present invention, photoSELEX is used to identify a nucleic acid ligand capable of modifying the biological activity of a target molecule (Example 16).

35 In a further embodiment of the present invention,

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the photoSELEX methodology is applied diagnostically to identify unique proteins associated with specific disease states (Example 17). In yet another embodiment of the present invention, nucleic acid ligands capable of crosslinking a target molecule associated with a specific disease condition are used *in vivo* to crosslink to the target molecule as a method of treating the disease condition (Examples 18 and 19).

In one embodiment of the present invention, RNA ligands identified by photoSELEX are used to detect the presence of the target protein by binding to the protein and then photocrosslinking to the protein. Detection may be achieved by incorporating ^{32}P or ^{33}P -labels and detecting material which is retained by a nitrocellulose filter by scintillation counting or detecting material which migrates correctly on an electrophoretic gel with photographic film. Alternatively, photoSELEX creates a fluorescent chromophore which is detected by fluorescence emission spectroscopy. Fluorescence emission for the products of reaction of 5-bromouracil to model peptides (as shown in Fig. 3) has been reported by Dietz and Koch (1987) *supra*. In another embodiment of the invention, a fluorescent label is covalently bound to the RNA and detected by fluorescence emission spectroscopy. In another embodiment of the invention, RNA ligands selected through photoSELEX are used to inhibit the target protein through the same process. In yet another embodiment, the photoselected ligand is bound to a support and used to covalently trap a target.

In a one embodiment of the invention, 5-iodouracil is incorporated into the RNA sequences of the candidate mixture, and light in the range of 320-325 nm is used for irradiation. This combination assures regionselective photocrosslinking of the 5-halouracil chromophore to the target protein without other non-specific photoreactions.

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In particular, tryptophan residues of protein and thymine and uracil bases of nucleic acids are known to be photoreactive. As shown in Figure 4, 5-iodouracil absorbs at 325 nm but tryptophan and the standard nucleic acid bases do not. The molar extinction coefficient for 5-iodouracil at 325 nm is 163 L/mol·cm. Monochromatic light in the region of 320-325 nm is preferably supplied by a frequency doubled tunable dye laser emitting at 320 nm or by a helium cadmium laser emitting at 325 nm.

10 In one embodiment of the invention a xenon chloride (XeCl) excimer laser emitting at 308 nm is employed for the photocrosslinking of 5-iodouracil-bearing RNA sequences to a target protein. With this laser, a high yield of photocrosslinking of nucleoprotein complexes is achieved within a few minutes of irradiation time.

15 In another embodiment of the invention, photocrosslinking of 5-iodouracil-bearing RNA sequences to a target protein is achieved with wavelength filtered 313 nm high pressure mercury lamp emission or with low pressure mercury lamp emission at 254 nm absorbed by a phosphor and re-emitted in the region of 300-325 nm. The latter emission is also carefully wavelength filtered to remove 254 nm light not absorbed by the phosphor and light in the region of 290-305 nm which could damage the protein.

25 In a further embodiment of the invention, photocrosslinking of BrU- or IU-bearing RNA sequences to a target protein is achieved with light in the region of 350-400 nm which populates directly the triplet state from the ground state. Monochromatic light from the third harmonic of a Neodymium YAG laser at 355 nm or the first harmonic from a xenon fluoride (XeF) excimer laser at 351 nm may be particularly useful in this regard.

30 In yet another embodiment of the invention the photoreactive nucleotides are incorporated into single
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stranded DNAs and amplified directly with or without the photoreactive nucleotide triphosphate.

5 A. Covalent SELEX and Nucleic Acid Ligands That Bind to HIV-1 Rev Protein With and Without Irradiation.

10 The target protein chosen to illustrate photo-SELEX is Rev from HIV-1. Rev's activity in vivo is derived from its association with the Rev-responsive element (RRE), a highly structured region in the HIV-1 viral RNA. Previous RNA SELEX experiments of Rev have allowed the isolation of very high affinity RNA ligands. The highest affinity ligand, known as "6a," (SEQ ID NO:5) has a K_d of approximately 1nM and is shown in Figure 16. The secondary structure of 6a, and its interaction with Rev, have been well characterized.

15 The construction of the nucleic acid library for photo-SELEX was based upon the Rev 6a sequence (SEQ ID NO:5). During the synthesis of the deoxyoligonucleotide templates for SELEX, the random region of the template was substituted by a "biased randomization" region, in which the ratio of the four input bases was biased in favor of the corresponding base in the 6a sequence. (Actual ratios were 62.5:12.5:12.5:12.5.) For example, if the 6a base for a particular position is G, then the base input mixture for this synthesis step is 62.5% G, and 12.5% of the other three bases.

20 The photoreactive uracil analogue 5-iodouracil (iU), which has been used to generate high-yield, region-specific crosslinks between singly-substituted iU nucleic acids and protein targets (Willis et al. (1993) Science 262:1255) was used for this example. The iU chromophore is reactive under long-wavelength ultraviolet radiation, and may photocouple to the aromatic amino acids of protein targets by the same mechanism as 5-bromouracil

25 (Dietz et al. (1987) J. Am. Chem. Soc. 109:1793). As

30

35

A "biased randomization" DNA oligonucleotide library, based upon the high affinity Rev ligand sequence 6a (SEQ ID NO:5), contains approximately 10^{14} unique sequences. This template was used for *in vitro* T7 transcription with 5-iUTP to generate fully-substituted iU RNA for selection. The photo-SELEX procedure alternated between affinity selection for Rev using nitrocellulose partitioning and monochromatic UV irradiation of the nucleoprotein complexes with denaturing polyacrylamide gel partitioning of the crosslinked complexes away from non-crosslinked RNA sequences. The final procedure utilized a simultaneous selection for affinity and crosslinking using competitor tRNA. Each round constitutes a selection followed by the conversion of recovered RNA to cDNA, polymerase chain reaction (PCR) amplification of the DNA, and *in vitro* transcription to generate a new pool of iU-RNA. To amplify RNA's recovered as covalent nucleoprotein complexes, the appropriate gel slice was isolated and proteinase K treated.

After 13 rounds of selection, the PCR products were
30 cloned and 52 isolates sequenced (Fig. 16, SEQ ID NOS:5-
55). Class 1 molecules, which comprise 77% of the total
sequences, contain a very highly conserved motif,
5'KDAACAN ...N'UGUUH'M'3' (SEQ ID NO:56) (Fig. 17).
Computer folding algorithms predict that this conserved
35 motif is base-paired and lies in a stem-loop structure.

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Subclasses a-d (Fig. 16, SEQ ID NOS:5-43) illustrate different strategies utilized in the "biased randomization" pool to obtain the consensus motif. Class 2 molecules show a highly conserved 10-base sequence (Fig. 17, SEQ ID NO. 57), which is predicted to fold with the 5' fixed region of the RNA and forms a structure distinct from either the class 1 or the 6a (SEQ ID NO:5) motif. All class 1 sequences exhibit biphasic binding to Rev, with high affinity dissociation constants (K_d s) ranging from 1-10nM. Class 2 sequences show monophasic binding to Rev with K_d s approximately of 30-50 nM. Analysis of round 13 sequences reveal that the frequency of the consensus motifs for class 1 and class 2 populations was very small in the starting pool, and some individual sequences arose only through the mutational pressures of the photo-SELEX procedure.

Cross-linking behavior differs between the two classes. Under high Rev concentrations (500 nM), and a 4 min. of 325 nm irradiation, class 2 molecules produce greater crosslink yield and efficiency than class 1 molecules (data not shown); presumably, this behavior allows the class 2 molecules, with relatively low affinity for Rev, to compete under the photo-SELEX procedures. For class 1 molecules, longer irradiation times will produce higher molecular weight crosslinked species. Although not bound by theory, it is proposed that the RNAs, which contain both an evolved binding domain for Rev, and the fixed regions needed for amplification in SELEX, are able to bind more than one Rev molecule per RNA. Since each RNA contains on average 21 iU bases (RNA length - 86 bases), it is thought that there is a certain promiscuity of the photoreaction that allows crosslinking of a single RNA to more than one Rev molecule at high protein concentrations. Class 2 molecules produce fewer high molecular weight species

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upon photocrosslinking; they are, on average, iU poor and may contain structures which do not allow binding/crosslinking to additional Rev molecules.

Analysis of individual round 13 RNAs revealed that a subpopulation could crosslink to Rev without laser irradiation. Thus, the single set of experiments demonstrated that both covalent SELEX without irradiation and photoSELEX with irradiation can be found in the same system. 4 of 15 round 13 sequences analyzed crosslink without laser irradiation (Fig. 16). From these few sequences, it was not readily possible to identify a sequence motif that confers laser independent crosslinking, although all molecules considered to date belong to the 1a subclass.

To further investigate laser-dependent and laser-independent crosslinking (LD-XL and LI-XL, respectively) and avoid the secondary photoproducts formed with full-length class 1 molecules, several small RNAs containing only the evolved sequences were constructed. Trunc2 and trunc24 (Figs. 18 and 19) (SEQ ID NOS:58 AND 59) are based upon clones #3 and #24, respectively, and show monophasic binding to Rev with Kds of 0.5 nM (trunc2) and 20 nM (trunc24). Trunc2 (Fig. 18) exhibits Ld-XL behavior, and trunc24 (Fig. 19) is capable of both LI-XL and LD-XL.

To explore the conformation and chemical requirements for LD-XL and LI-XL, crosslinking reactions were performed with trunc2 (SEQ ID NO:58) and trunc24 (SEQ ID NO:59) and several Arginine Rich Motif (ARM) proteins. The class of RNA-binding proteins includes the target protein, HIV-1 Rev, and also HIV-1 Tat and the highly similar HIV-2 Rev. LD-XL reactions with trunc2 (Fig. 18, SEQ ID NO:58) show that trunc2 is capable of crosslinking specifically to both HIV-1 and HIV-2 Rev proteins, but not HIV-1 Tat. The two slightly different

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migrating nucleoprotein complexes probably represent the ability of trunc2 to use one of two iU nucleotides to crosslink the Rev proteins. Although not bound by theory, it is proposed that a tryptophan residue present in the highly similar ARMs of both Rev proteins is the amino acid necessary for the specific photo-crosslinking of our high-affinity RNA ligands.

Trunc24 LI-XL (Fig. 19, SEQ ID NO:59), performed with the same proteins, shows crosslinking only to HIV-1 Rev. Like trunc3, trunc24 can photo-crosslink to HIV-2 Rev (data not shown). It was also observed that this LI-crosslink is reversible under highly denaturing conditions, or with high concentration of nucleic acid competitors. Although not bound by theory, these observations lead to the postulation that LI-XL proceeds by a Michael adduct between the 6 position of an IU and a cystein residue, or possible a 5 position substitution reaction. This postulation is consistent both with the observation that iU undergoes Michael adduct formation more readily than U, and the fact that HIV-1 Rev contains three cysteines, while HIV-2 Rev contains none.

To test for the ability of trunc24 (SEQ ID NO:59) to discriminate HIV-1 Rev in a complex mixture, trunc24 and 10 μ g of human fibroblast nuclear extract together with decreasing amounts of HIV-1 Rev (Fig. 20). At 50 nM Rev and a 1:100 weight ratio of Rev to nuclear extract, it was possible to see a very significant crosslinked product between trunc24 and Rev. Nuclear extracts and trunc24 alone resulted in no crosslinked products.

Example 1 describes the synthesis of hairpin RNA oligonucleotides RNA-1 (SEQ ID NO:1), RNA-2 (SEQ ID NO:2), and RNA-3 (SEQ ID NO:3) using 5-bromouridine triphosphate, 5-iodouridine triphosphate and uridine triphosphate, respectively. Experiments determining the RNA-protein binding curves for RNA-1 (SEQ ID NO:1), RNA-2

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(SEQ ID NO:2), and RNA-3 (SEQ ID NO:3) to the bacteriophage R17 coat protein are described in Example 2. Example 3 describes the photocrosslinking of the RNA oligonucleotides to the R17 coat protein. The amino acid residue of the R17 coat protein photocrosslinked by RNA-1 (SEQ ID NO:1) after illumination via xenon chloride (XeCl) excimer laser at 308 nm is described in Example 4. Example 5 describes the photocrosslinking of iodouracil-substituted RNA-2 (SEQ ID NO:2) to the R17 coat protein by monochromatic emission at 325 nm. Example 6 describes the photocrosslinking of RNA-1 (SEQ ID NO:1) and RNA-2 (SEQ ID NO:2) to the R17 coat protein achieved after broad-band emission illumination with a transilluminator. Example 7 describes the photoreaction of 5-iodouracil with N-acetyltyrosine N-ethyl amide, which appeared to yield a photocrosslink similar to that achieved with 5-bromouracil-substituted nucleic acids to associated proteins. The preparation of a cDNA from a RNA photocrosslinked to the R17 coat protein is described in Example 8. Example 9 describes the photocrosslinking of an IC-substituted RNA ligand to the R17 coat protein.

Example 10 describes the incorporation of halogenated nucleotides into DNA. Examples 11-15 describes photoSELEX protocols which may be used to produce specific photoreactive nucleic acid ligands. Example 11 describes a continuous photoSELEX method. Example 12 describes a method in which nucleic acid ligands initially selected through SELEX are subsequently selected through photoSELEX for the capacity to crosslink to the target molecule. Example 13 describes one embodiment in which nucleic acid ligands identified through photoSELEX are then subjected to selection through SELEX and selected for ability to bind the target molecule. Example 14 describes another embodiment wherein a limited SELEX selection is followed by

5 Example 16 describes a method for selecting a nucleic
acid ligand capable of modifying the biological activity
of a target molecule. Example 17 describes a diagnostic
procedure which uses the SELEX and photoSELEX methods to
identify proteins associated with specific disease
10 processes.

15 introduced into a patient in a number of ways known to
the art. For example, the photoSELEX ligand may be
transiently or constitutively expressed in the
appropriate cells of a patient with the disease.
Alternatively, the photoSELEX ligand may be taken into a
20 patient's cells as a double-stranded DNA which is
transcribed in the cell in the presence of iodinated
cytosine. Iodinated cytosine may be administered to the
patient, followed by irradiation with X-rays. IC
incorporated into the nucleic acid ligand synthesized in
25 the appropriate cells allows the ligand to crosslink and
inactivate the target molecule. Further methods of
introducing the photoSELEX ligand into a patient include
liposome delivery of the halogenated ligand into the
patient's cells.

30 Example 20 describes the production of modified
nucleic acid ligands the crosslink, with or without
irradiation, to HIV-1 Rev protein. Figure 15 shows the
results of crosslinking to the bulk candidate mixture at
various rounds of SELEX. rd1-round 1 pool RNA; rd7-round
35 7 pool RNA; rd10-round 10 pool RNA; rd13-round 13 pool

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RNA; rd13/PK, photocrosslinked round 13 pool RNA
 proteinase K treated (35ul of a 100 ul reaction was
 incubated in 0.5% SDS, 50 ug/ml Proteinase K and 1mM EDTA
 at 65 C. for one hour); rd13/no iU-round 13 pool RNA
 5 transcribed with UTP (no iU). R-free RNA; XL-crosslinked
 nucleoprotein complex.

Figure 16 shows the sequences sequenced after 13
 rounds of SELEX (SEQ ID NOS:5-55). The sequences are
 aligned for maximum homology to the 6a sequence (SEQ ID
 10 NO:5). Underlines represent potential base pairing as
 indicted by computer RNA folding algorithms. Dashed
 underlines represent the 6a ligand "bubble" motif.
 Sequences flanked by underline represent either loop or
 bulge regions. Dashes are placed to maximize alignment
 15 with 6a. * denotes that two isolates were obtained. +
 indicates laser independent crosslinking and - denotes
 the lack of laser independent crosslinking to HIV-1 Rev.
 Figure 17 (SEQ ID NOS:56-57) shows the consensus for
 class 1 and class 2 ligands. Figures 18 and 19 show the
 20 sequence of Trunc2 (SEQ ID NO:58) and Trunc24 (SEQ ID
 NO:59) and the specificity results. 500 nM protein, 20µg
 tRNA, and approximately 1 nM of kinased trunc2 RNA were
 incubated for 10 min. at 37°C and irradiated for 4 min at
 325 nm. t2-trunc2 RNA irradiated without added protein;
 25 t2Rev1/O'-trunc2 RNA, HIV-1 Rev protein and 0 min. of
 irradiation; t2/Rev1/4'-trunc2 RNA, HIV-1 Rev protein,
 adn 4 min. of irradiation; t2/Rev1/4'/PK-trunc2 RNA, HIV-
 1 Rev protein, 4 min. or irradiation, and proteinase K
 treated as in Fig. 1; t2/Rev2/4'-trunc2 RNA, HIV-2 Rev
 30 protein, and 4 min. of irradiation; t2/Tat/4'-trunc2 RNA,
 HIV-1 Tat protein, and 4 min. of irradiation. R-free RNA;
 XL-crosslinked nucleoprotein complex. Figure 20 shows
 the trunc24 photoindependent crosslinking with HIV-1 Rev
 in the presence of human nuclear extract.

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II. Solution SELEX.

This embodiment of the present invention presents several improved methods for partitioning between oligonucleotides having high and low affinity for a target molecule. The method of the present invention has several advantages over prior art methods of partitioning: (1) it allows the isolation of nucleic acid ligands to the target without also isolating nucleic acid ligands to the partitioning matrix; (2) it increases the speed and accuracy by which the oligonucleotide candidate mixture is screened; and (3) the solution SELEX procedure can be accomplished in a single test tube, thereby allowing the partitioning step to be automated.

The materials and techniques required by the method of the present invention are commonly used in molecular biology laboratories. They include the polymerase chain reaction (PCR), RNA or DNA transcription, second strand DNA synthesis, and nuclease digestion. In practice, the techniques are related to one another in a cyclic manner as illustrated in Figure 21.

In the SELEX method, described by Tuerk and Gold (1990) Science 249:1155 and illustrated in Figure 21, a single-stranded nucleic acid candidate mixture is generated by established procedures on a nucleic acid synthesizer, and is incubated with dNTP and Klenow fragment to generate a population of double-stranded DNA templates. The double-stranded DNA or the RNA transcribed from them are purified, and contacted with a target molecule. RNA sequences with enhanced affinity to the target molecule form nucleic acid-target complexes. This is followed by partitioning of bound and unbound nucleic acids, and separation of the target molecule from the bound nucleic acids. cDNA is synthesized from the enhanced affinity nucleic acids and double-stranded DNA generated by PCR amplification. The cycle is repeated

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until the complexity of the candidate mixture has decreased and its affinity as well as specificity to the target has been maximized.

5 A novel feature of the solution SELEX method is the means by which the bound and free members of the nucleic acid candidate mixture are partitioned. In one embodiment of the method of the present invention, generation of two physically distinct cDNA pools is accomplished by use of primer extension inhibition. One
10 cDNA extension step is added to the basic SELEX protocol between steps 2 and 3 above, which allows the generation of two physically distinct cDNA pools -- one having high affinity for the target and one having low affinity for the target -- which are easily distinguished and
15 separated from each other. Primer extension inhibition analysis is a common technique for examining site-bound proteins complexed to nucleic acids (Hartz et al. (1988) Methods Enzymol. 164:419), and relies on the ability of high affinity complexes to inhibit cDNA synthesis.
20 Examples of protein-nucleic acid interactions studied by primer extension inhibition include ribosome binding to the mRNA ribosome-binding site (Hartz et al. (1988) Meth. Enzym. 164:419) as well as binding of the unique *E. coli* translation factor, SELB protein, to the mRNA
25 selenocysteine insertion sequence (Baron et al. (1993) Proc. Natl. Acad. Sci. USA 90:4181).

In one embodiment of the solution SELEX scheme, the first cDNA extension is performed in the presence of chain terminating nucleotide triphosphates. Under these
30 conditions, oligonucleotides with low affinity for the target which form fast dissociating complexes with the target are converted into truncated cDNAs with a 3'-end terminated with a nonextendible nucleotide. The truncated cDNA chain is unable to anneal to the PCR
35 primers, and therefore, is non-amplifiable. In contrast,

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tight complexes formed between high affinity oligonucleotides and the target molecule, characterized by slow dissociation rates, inhibit cDNA extension. The chain terminators are not incorporated into the nascent cDNA chain synthesized from the high affinity oligonucleotide because cDNA synthesis is blocked by the tightly bound target molecule. Full length cDNA from the high affinity complexes are obtained during a second round of cDNA extension in which the target and chain terminators have been removed from the system. Thus, weak affinity complexes are converted into truncated cDNA lacking the primer annealing site while tight complexes are converted into full length cDNA and are amplified by PCR (Fig. 22). The stringency of this method is easily modified by varying the molar ratio of chain terminators and dNTPs or the concentration of the polymerase, as primer extension inhibition is sensitive to polymerase concentration (Ringquist et al. (1993) Biochemistry in press). As used in the present disclosure, the term "stringency" refers to the amount of free RNA that will be converted into PCR product.

Therefore, one crucial feature of the invention is its ability to partition strong and weak affinity complexes into amplifiable and non-amplifiable nucleic acid pools without requiring a partitioning matrix. It is the unique properties of these cDNA pools that allow selective amplification of the high affinity ligand.

The target molecule can be a protein (either nucleic acid or non-nucleic acid binding protein), nucleic acid, a small molecule or a metal ion. The solution SELEX method allows resolution of enantiomers as well as the isolation of new catalytic nucleic acids.

Primer extension inhibition may be achieved with the use of any of a number of nucleic acid polymerases, including DNA or RNA polymerases, reverse transcriptase,

and Q β -replicase.

The candidate mixture of nucleic acids includes any nucleic acid or nucleic acid derivative, from which a complementary strand can be synthesized.

5 Prior art partitioning included use of
nitrocellulose or an affinity column. One disadvantage
of the prior art partitioning was the phenomenon of
matrix binders in which nucleic acids that specifically
10 bind the partitioning matrix are selected along with
those that specifically bind the target. Thus, one
advantage of the method of the present invention is that
it overcomes unwanted selective pressure originating with
use of a partitioning matrix by only using such matrixes
after nucleic acids with high affinity for the target
15 have been partitioned in solution and amplified.
Moreover, the ability to partition strong and weak
affinity complexes during cDNA synthesis, based on the
ability of only the strongest complexes to inhibit
extension by a polymerase, results in the selection of
20 only the highest affinity nucleic acid ligands. It is
estimated that complexes with dissociation constants in
the nanomolar or less range will efficiently block cDNA
synthesis. The method of the present invention is
expected to preferentially screen nucleic acid candidate
25 mixtures for members that bind the target at this limit.

The use of primer extension inhibition allows partitioning of the oligonucleotide candidate mixture into two pools - those oligonucleotides with high target affinity (amplifiable) and those with low target affinity (non-amplifiable). As described above, chain terminators may be used to poison the first extension product, rendering the low affinity cDNAs non-amplifiable.

In another embodiment of the method of the present invention, restriction enzymes are used to selectively
35 digest the cDNA generated from the low affinity nucleic

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acids. A number of restriction enzymes have been identified that cleave single-stranded DNA. These enzymes cleave at specific sequences but with varying efficiencies. Partitioning of weak and strong affinity nucleic acids is accomplished by primer extension in the presence of the four dNTPs, followed by removal of the target and a second extension with modified nucleotides that are resistant to enzymatic cleavage. The cDNA pools can then be incubated with the appropriate restriction enzyme and the cDNA synthesized during the first extension cleaved to remove the primer annealing site and yield a non-amplifiable pool. Increased efficiency of cleavage is obtained using a hairpin at the restriction site (RS) to create a localized double-stranded region (Fig. 24).

In another embodiment of method of the present invention, cDNA sequences corresponding to low affinity nucleic acids are rendered selectively degradable by incorporation of modified nucleotide into the first cDNA extension product such that the resulting cDNA is preferentially degraded enzymatically or chemically.

In another embodiment of the method of the present invention, the first extension product can be removed from the system by an affinity matrix. Alternatively, the matrix could be used to bind the second extension product, e.g., the cDNAs corresponding to high affinity nucleic acids. This strategy relies on the incorporation of modified nucleotides during cDNA synthesis. For instance, the first cDNA extension could be performed in the presence of modified nucleotides (e.g., biotinylated, iodinated, thiolabelled, or any other modified nucleotide) that allow retention on an affinity matrix (Fig. 25). In an alternate embodiment of the method of the present invention, a special sequence can also be incorporated for annealing to an affinity matrix. Thus,

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first synthesis cDNAs can be retarded on commercially obtainable matrices and separated from second synthesis cDNA, synthesized in the absence of the modified nucleotides and target.

5 In another embodiment of the invention, exonuclease hydrolysis inhibition is used to generate a pool of high affinity double-stranded nucleic acid ligands.

10 In yet another embodiment of the invention, the solution SELEX method is used to isolate catalytic nucleic acids.

In another embodiment of the invention, solution SELEX is used to preferentially amplify single-stranded nucleic acids.

15 In a further embodiment of the invention, the solution SELEX method is automated.

20 Removal of the target to allow cDNA synthesis from the high affinity nucleic acids can also be accomplished in a variety of ways. For instance, the target can be removed by organic extraction or denatured by temperature, as well as by changes in the electrolyte content of the solvent. In addition, the molecular repertoire of the candidate mixture that can be used with the invention include any from which a second complementary strand can be synthesized. Single-stranded DNA as well as RNA can be used, as can a variety of other modified nucleotides and their derivatives.

25 The following non-limiting examples illustrate the method of the present invention. Example 21 describes the solution SELEX process wherein partitioning between high and low affinity nucleic acids is achieved by primer extension inhibition. Example 22 illustrates the solution SELEX process wherein partitioning is achieved by restriction enzyme digestion of low affinity RNA. Example 23 describes the solution SELEX process wherein
35 low affinity nucleic acids are separated from high

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affinity nucleic acids by affinity chromatography. Example 24 describes the isolation of high affinity double-stranded nucleic acid ligands with the use of exonuclease inhibition. Example 25 describes the isolation of catalytic nucleic acids. Example 26 describes an automated solution SELEX method.

The examples provided are non-limiting illustrations of methods of utilizing the present invention. Other methods of using the invention will become apparent to those skilled in the art from the teachings of the present disclosure.

Example 1. Synthesis of RNA Sequences RNA-1, RNA-2, RNA-3, and RNA-7 and R17 Coat Protein.

RNA-1 (SEQ ID NO:1), RNA-2 (SEQ ID NO:2), and RNA-3 (SEQ ID NO:3) shown in Figure 6 and RNA-7 (SEQ ID NO:4) shown in Figure 12 were prepared by *in vitro* transcription from synthetic DNA templates or plasmids using methodology described by Milligan and co-workers (Milligan et al. (1987) Nucleic Acids Res. 15:8783). Transcription reactions contained 40 mM tris(hydroxymethyl)aminomethane hydrochloride (Tris-HCl, pH 8.1 at 37°C), 1 mM spermidine, 5 mM dithiothreitol (DTT), 50 µg/ml of bovine serum albumin (BSA), 0.1% (v/v) Triton X-100, 80 mg/ml of polyethylene glycol (m_r 8000), and 0.1 mg/ml of T7 RNA polymerase. Larger quantities of RNA were prepared with 3-5 mM of each of the nucleotide triphosphates (NTPs), 25 mM magnesium chloride, and 1 µM DNA template or 0.1 µg/ml of plasmid. Body-labeled RNAs were prepared in 100 µM reactions with 1 mM each of the three NTPs, 0.25 mM of the equivalent radiolabelled NTP ($[\alpha\text{-}^{32}\text{P}]$ NTP, 5 µCi), 15 mM MgCl_2 , and 0.1 mg/ml of T7 RNA polymerase. Nucleotides, including 5-iodouridine triphosphate and 5-bromouridine triphosphate, were obtained from Sigma Chemical Co., St. Louis, Missouri.

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RNA fragments were purified by 20% denaturing polyacrylamide gel electrophoresis (PAGE). The desired fragment was eluted from the polyacrylamide and ethanol-precipitated in the presence of 0.3 M sodium acetate.

5 R17 bacteriophage was propagated in *Escherichia coli* strain S26, and the coat protein was purified using the procedure described by Carey and coworkers (Carey et al. (1983) Biochemistry 22:4723).

10 Example 2. Binding Constants for RNA-1 and RNA-2 to R17 Coat Protein.

RNA-protein binding curves for hairpin variants RNA-1 (SEQ ID NO:1), RNA-2 (SEQ ID NO:2) and RNA-3 (SEQ ID NO:3) to the bacteriophage R17 coat protein are shown in
15 Figure 7. The association constants between coat protein and the RNA hairpin variants were determined with a nitrocellulose filter retention assay described by Carey and co-workers (Carey et al. (1983) supra). A constant, low-concentration of ^{32}P -labeled RNA was mixed with a
20 series of coat protein concentrations between 0.06 nM and 1 μM in 10 mM magnesium acetate, 80 mM KCL, 80 $\mu\text{g/ml}$ BSA, and 100 mM Tris-HCl (pH 8.5 at 4°C) (TMK buffer). These were the same solution conditions used in the crosslinking experiments. After incubation at 4°C for
25 45-60 min, the mixture was filtered through a nitrocellulose filter and the amount of complex retained on the filter determined by liquid scintillation counting. For each experiment the data points were fit to a non-cooperative binding curve and the K_d value shown
30 in Figure 7 was calculated.

Example 3. Photocrosslinking of RNA-1 and RNA-2 to R17 Coat Protein at 308 nm.

^{32}P -Labeled RNA sequences RNA-1 (SEQ ID NO:1) and
35 RNA-2 (SEQ ID NO:2) (5 nM) and R17 coat protein (120 nM)

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were each incubated on ice in 100 mM Tris-HCl (pH 8.5 at 4°C), 80 mM KCl, 10 mM magnesium acetate, 80 µg/ml of BSA for 15-25 min before irradiations. These are conditions under which the RNA is fully bound to the coat protein.

5 The RNAs were heated in water to 85°C for 3 min and quick cooled on ice before use to ensure that the RNAs were in a hairpin conformation (Groebe and Uhlenbeck (1988) Nucleic Acids Res. 16:11725). A Lambda Physik EMG-101 excimer laser charged with 60 mbar of xenon, 80 mbar of
10 5% HCl in helium and 2360 mbar of helium was used for 308 nm irradiations. The output of the XeCl laser was directed unfocused toward a 4 mm wide by 1 cm path length quartz cuvette containing the RNA-protein complex. The laser was operated in the range of 60 mJ/pulse at 10 Hz;
15 however, only about 25% of the laser beam was incident upon the reaction cell. Photocrosslinking yields of RNA-1 (SEQ ID NO:1) and RNA-2 (SEQ ID NO:2) to the R17 bacteriophage coat protein as a function of irradiation time are shown in Figure 8. Crosslinked RNA was
20 separated from uncrosslinked RNA by PAGE, and the yields were determined by autoradiography. Crosslinking of 5-bromouracil-containing variant RNA-1 (SEQ ID NO:1) maximized at about 40% because of competitive photodamage to the coat protein which inhibits binding to the RNA
25 (Gott et al. (1991) *supra*). Less photodamage to coat protein occurred with RNA 2 because of the shorter irradiation time.

Crosslinking as a function of photons absorbed indicated that the quantum yield for crosslinking of BrU-
30 RNA 1 is 0.014 and for crosslinking of IU-RNA-2 (SEQ ID NO:2), 0.006 with irradiation at 308 nm. In spite of the lower quantum yield, a higher crosslinking yield was obtained with IU-RNA 2 because of the seven times higher absorption probability of the IU chromophore at 308 nm.
35 BrU and IU absorb at 308 nm with molar extinction

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coefficients of 385 and 2640 L/mol·cm, respectively. Hence, a high level of photocrosslinking of the IU-RNA was achieved prior to protein damage.

5 Example 4. Identification of the Amino Acid Residue
 Involved in the Crosslink of RNA-1 to R17
 Coat Protein

Large scale crosslinking of RNA-1 (SEQ ID NO:1) to
10 R17 coat protein. A 10 ml solution containing 300 nM 5'-
end-labeled RNA and 500 nM coat protein was incubated on
ice in the presence of 100 mM Tris-HCl (pH 8.5 at 4°C),
10 mM Mg(OAc)₂, 80 mM KCL, 80 mg bovine serum albumin
(BSA), and 5 mM dithiothreitol (DTT) for 10-90 min. A
15 Lambda Physik EMG-101 excimer laser was used for
monochromatic irradiation at 308 nm. The beam output was
measured at 69 ± 5 mJ/pulse at 10 Hz. Approximately 50%
of the beam was focused through a 7 mm-diameter circular
beam mask into a 1 cm path length quartz cuvette in a
thermostated cell holder. The laser power was measured
20 with a Scientech 360-001 disk calorimeter power meter.
The temperature was regulated at 4 ± 2°C with a Laude RC3
circulating bath.

 The 10 ml reaction mixtures were prepared just prior
to the irradiations which were performed in 2 ml
25 fractions. After 5 min of irradiation the protein
concentration was brought to 1 μM. The reaction mixture
was then incubated for 3 min to allow exchange of
photodamaged protein for fresh protein in the
nucleoprotein complex and irradiated for an additional 5
30 min. This step was repeated nine times to give 90 ml of
irradiated sample. The crosslinking, analyzed by 20%
denaturing PAGE, and quantitated on a Molecular Dynamics
Phosphoimager, revealed 22% crosslinking.

 The 90 ml sample contained 5.9 nmol of crosslinked
35 RNA, 21 nmol of free RNA, 97 nmol of free coat protein,

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and 7.2 mg of BSA. The total volume was reduced to 20 ml and split equally between two 50 ml polypropylene screw cap centrifuge tubes (Nalgene) and ethanol precipitated overnight at -20°C. The RNA and proteins were spun down to a pellet at 13,000 rpm in a fixed angle J-20 rotor with an Beckman J2-21 centrifuge. Each pellet was resuspended in 1 ml of 0.5 M urea, 50 mM Tris-HCl pH 8.3, and 0.2% SDS for 48 h at 4°C with shaking. The fractions were combined, and the SDS was then removed by precipitation so as not to decrease the activity of trypsin. This was achieved using 40 mM KCl, and the precipitate was removed by spinning through a 0.22 µm cellulose acetate spin filter. The trypsin conditions were optimized using 500 µl of the solution.

Proteolytic Digestion. The remaining 1.5 ml of crosslinked RNA solution containing free RNA and protein was brought to 6 ml to contain 1 M urea, 20 mM CaCl₂, and 6 mM DTT, and then 1.61 mg (1:5 w/w.) trypsin-TPCK (251 units/mg) was added. The reaction proceeded at 36°C for 2 h at which time 1.61 mg more trypsin was added. At 4 h a 100 µL aliquot was removed and the reaction stopped by quick freezing. The reaction was analyzed by 20% polyacrylamide 19:1 crosslinked, 7 M urea, 90 mM Tris-borate/2 mM EDTA (TBE) gel electrophoresis (20% urea denaturing PAGE).

Purification of the digested crosslinked RNA. The trypsin reaction mixture was brought to 10 ml to reduce the molar concentration of salt, and run through a 240 µl DEAE ion exchange centrifuge column. The column was washed with 100 mM NaCl and spun dry in a bench top centrifuge to remove free peptide. The column bound material containing the RNA and crosslinked tryptic fragment was eluted from the column with 1 ml of 600 mM NaCl and the column spun dry. An additional 200 µl of 600 mM NaCl was spun through the column. The two

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fractions were pooled, ethanol precipitated and pelleted at 10,000 rpm for 35 min at 4°C. The pellet was resuspended in 25 µl of 7 M urea-TBE buffer, 10 mM DTT, 0.1% bromophenol blue, 0.1% xylene cyanol, and heated to 85°C for 4 min and purified by 20% denaturing PAGE. The gel ran for 3.5 h at 600 V. A 5 min phosphorimage exposure was taken of the gel. The digested protein-RNA crosslink was then electrolytically blotted from the gel onto a PVDF protein sequencing membrane (0.2 micron) from Bio-RAD. The membrane was air dried, coomassie stained for 1 min, destained for 2 min in 50% MeOH: 50% H₂O, and rinsed twice with deionized H₂O. An autoradiogram was made of the membrane to visualize the digested protein RNA crosslink which was excised from the membrane and submitted for Edman degradation. The immobilized peptide was sequenced by automated Edman degradation, performed on an Applied Biosystems 470A sequencer using manufacturer's methods and protocols (Clive Slaughter, Howard Hughes Medical Institute, University of Texas, Southwestern). The Edman analysis indicated that the position of the crosslink was tyrosine-85 based upon the known amino acid sequence (Weber (1983) Biochemistry 6:3144).

Example 5. Photocrosslinking of RNA-2 to R17 Coat Protein at 325 nm.

In an experiment analogous to that described in Example 3, IU-substituted RNA-2 (SEQ ID NO:2) was photocrosslinked to R17 coat protein with monochromatic emission at 325 nm from an Omnicrome HeCd laser (model 3074-40M325). The power output of the HeCd laser was 37 mW and the total beam of diameter 3 mm was incident upon the sample. To increase excitation per unit time the beam was reflected back through the sample with a dielectric-coated concave mirror. Crosslinked RNA was

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separated from uncrosslinked RNA by PAGE, and the yields were determined with a PhosphoImager. The percent of the RNA crosslinked to the protein as a function of irradiation time is shown in Figure 9. High-yield crosslinking occurred without photodamage to the R17 coat protein. In a separate experiment analogous irradiation of coat protein alone at 325 nm with yet a higher dose resulted in protein which showed the same binding constant to R17 coat protein. Irradiation at 325 nm of BrU-containing RNA-1-R17 coat protein complex did not result in crosslinking because the BrU chromophore is transparent at 325 nm.

Example 6. Photocrosslinking of RNA-1 and RNA-2 to R17 Coat Protein with a Transilluminator.

In an experiment analogous to that described in Example 3, RNA-1 (SEQ ID NO:1) and RNA-2 (SEQ ID NO:2) were photocrosslinked to the R17 coat protein with broadband emission in the range of 312 nm from a Fisher Biotech Transilluminator (model FBTIV-816) filtered with polystyrene. Crosslinked RNA was separated from uncrosslinked RNA by PAGE, and the yields were determined by autoradiography. Percent RNAs crosslinked to protein as a function of irradiation time is shown in Figure 10.

Example 7. Photoreaction of 5-Iodouracil with N-Acetyltyrosine N-Ethyl Amide.

N-acetyltyrosine N-ethylamide was prepared as described by Dietz and Koch (1987) *supra*. Irradiation of a pH 7, aqueous solution of iodouracil and 10 mol equivalent excess of N-acetyltyrosine N-ethyl amide at 308 nm with a XeCl excimer laser gave a photoadduct identical to the photoadduct (structure 6) from irradiation of bromouracil and N-acetyltyrosine N-ethylamide (Dietz and Koch (1987) *supra*) as shown in

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Figure 11. Product comparison was performed by C-18 reverse phase HPLC and by ^1H NMR spectroscopy. Although little is known about the mechanism of photocrosslinking of IU-substituted nucleic acids to associated proteins, this result suggests that it is similar to that of photocrosslinking of BrU-substituted nucleic acids to associated proteins.

Example 8. Preparation of a cDNA from an RNA Photocrosslinked to a Protein.

RNA-7 (SEQ ID NO:4) (Fig. 11) was prepared using methodology as reported in Example 1 using a plasmid instead of a DNA template. The photocrosslinking was performed as described in Example 3. A 4 ml reaction mixture consisting of 6.75 nM RNA and 120 nM R17 coat protein was irradiated, 2 ml at a time, at 308 nm with unfocused emission from a XeCl excimer laser. The laser produced 50 mJ/pulse and was operated at 10 Hz. The reaction proceeded to near quantitative crosslinking, 85-90%, in 5 min of irradiation. After crosslinking, 1 ml of the total reaction mixture was removed; EDTA (80 mM), SDS (0.1%), and CaCl_2 (0.1 mM) were added; the free (unbound) RNA present was purified away; and the protein digested with Proteinase K at 60°C for 30 min. The RNA bound to residual protein was ethanol precipitated to remove salts and spun to a pellet. The pellet was washed three times with 70% ethanol to remove any residual salts. A reverse transcription reaction was employed to make a complementary DNA copy of the RNA template. A 13-base promoter was annealed to the RNA and the reverse transcription reaction was performed under the standard conditions of the manufacturer, Gibco (Gaithersburg, MD), and was stopped after 1 hr. The cDNA was body labelled with ^{32}P -labelled deoxycytidine triphosphate. The RNA template was then removed by hydrolyzing with 0.2 M

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sodium hydroxide at 100°C for 5 min. The formation of the cDNA was followed by PAGE. A hydrolysis ladder and markers were added to the gel to determine the length of the cDNA. The cDNA co-migrated with the 44 nucleotide RNA template. If there had been a stop in the cDNA as a result of crosslinking modification, a shortened product of 31 nucleotides would have been observed. A small amount of a stop product was observed in the 22-25 nucleotide region of the gel, but this may have resulted from the hairpin secondary structure which begins at position 25 of the cDNA on the RNA template. No stop in the 31 nucleotide region of the gel appeared; this established that the reverse transcriptase had read through the position of the crosslink. A diagram of the gel appears in Figure 14.

Example 9. Iodocytosine Photocrosslinking.

5-iodocytosine (IC) was incorporated in a hairpin RNA (RNA 8) that contained cytosine at the -5 position and bound the R17 coat protein with high affinity. The IC-bearing RNA is designated RNA 9. RNA 9 (5 nM) and R17 coat protein (120 nM) were incubated on ice in 100 mM Tris-HCl (pH 8.5 at 4°C)/80 mM KCl/10 mM magnesium acetate/ 80 µg/ml BSA for 15-25 min prior to irradiation. The RNA in water was heated to 85°C for 3 min and quick cooled on ice before use to ensure that it would be in a conformation that bound the coat protein (Groebe and Uhlenbeck (1988) *supra*). The complex was irradiated for 5 min at 4°C, and the experiment was compared to control irradiations of RNA 2 and RNA 8 coat protein complexes. Irradiation of RNA 8-coat protein complex resulted in no crosslinked product. Irradiation of RNA 9-coat protein complex resulted in the formation of a crosslink that formed in high yield (70-80%) similar to the yield of the control irradiation of RNA 2-coat protein complex (80-

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90%) Crosslinking of RNA 9 is presumed to occur through a similar mechanism as RNAs containing IU at position -5 of the loop hairpin (Figs. 6 and 12). This assumption is based on the specificity of the crosslink since RNA 8 did not photocrosslink.

Example 10. Incorporation of Halogenated Nucleotides into DNA Ligands.

Photoreactive nucleotides may be incorporated into a DNA ligand capable of crosslinking to a target molecule upon irradiation by the methods discussed above. 5-Bromodeoxyuracil (BrdU), 8-bromo-2'-deoxyadenine, and 5-iodo-2'-deoxyuracil are examples of such photoreactive nucleotides.

Example 11. PhotoSELEX.

In one embodiment of the present invention, the photoSELEX method is applied to completion in the selection of a nucleic acid ligand which binds and photocrosslinks to a target molecule.

A randomized set of nucleic acid oligonucleotides is synthesized which contain photoreactive groups. The oligonucleotides of the candidate mixture may be partially or fully saturated at each available position with a photoreactive group. The candidate mixture is contacted with the target molecule and irradiated at the appropriate wavelength of light. Oligonucleotides crosslinked to the target molecule are isolated from the remaining oligonucleotides and the target molecule removed. cDNA copies of the isolated RNA sequences are made and amplified. These amplified cDNA sequences are transcribed into RNA sequences in the presence of photoreactive groups, and the photoSELEX process repeated as necessary.

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Example 12. Selection of Enhanced Photocrosslinking
Ligands: SELEX Followed by PhotoSELEX.

In one embodiment of the method of the present invention, selection of nucleic acid ligands through
5 SELEX is followed by selection through photoSELEX for ligands able to crosslink the target molecule. This protocol leads to ligands with high binding affinity for the target molecule that are also able to photocrosslink to the target.

10 Photoreactive nucleotides are incorporated into RNA by T7 polymerase transcription with the reactive nucleotide triphosphate in place of a specified triphosphate. For example, 5-bromouridine triphosphate is substituted for uridine triphosphate or 8-bromo-
15 adenosine triphosphate is substituted for adenosine triphosphate. A randomized set of RNA sequences containing photoreactive nucleotides are generated and the SELEX methodology applied. The initial SELEX rounds are used to eliminate intrinsically poor binders and
20 enhance the pool of molecules that converge to form a pool of RNAs that contain the photoreactive group(s) and which bind to the target molecule. Aliquots from the initial SELEX rounds are irradiated and the enhancement of photocrosslinking followed via PAGE as the rounds
25 proceed. As a slower migrating band representing crosslinked products starts to become evident, the pool of RNAs are introduced into rounds of photoSELEX. RNAs that have a photoreactive group adjacent to a reactive amino acid residue in the nucleoprotein complexes form a
30 crosslink and are selected and RNAs that do not have reactive nucleotides in proximity to reactive target residues are eliminated.

This protocol selectively applies photoSELEX selection to previously identified ligands to a target
35 molecule.

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Example 13. PhotoSELEX Followed by SELEX.

In another embodiment of the method of the present invention, an RNA ligand able to photocrosslink a target molecule is preselected through the photoSELEX methodology. Subsequently, SELEX is performed to select a crosslinking oligonucleotide for ability to bind the target molecule.

Example 14. Limited SELEX Followed by PhotoSELEX.

In this embodiment of the present invention, nucleic acid ligands are selected through the SELEX process for a limited number of selection rounds. SELEX is not applied to completion as in Example 12. Rather, the candidate mixture is partially selected for oligonucleotides having relatively enhanced affinity for the target molecule. The random oligonucleotides of the candidate mixture contain photoreactive groups and the initial SELEX selection is conducted in the absence of irradiation. PhotoSELEX is then performed to select oligonucleotides able to crosslink to the target molecule.

This protocol allows the selection of crosslinking ligands from a pool of oligonucleotides with a somewhat enhanced capacity to bind the target molecule and may be useful in circumstances where selection to completion through SELEX does not yield crosslinking ligands.

Example 15. Limited Directed PhotoSELEX.

In one embodiment of the method of the present invention, in which nucleic acid ligands identified through SELEX are subjected to limited randomization, followed by selection through photoSELEX.

The construction of the DNA template used to transcribe the partially randomized RNA is based on the sequence of the initially selected ligand and contains at each position primarily the nucleotide that is

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complementary to that position of the initial selected RNA sequence. However, each position is also partially randomized by using small amounts of each of the other three nucleotides in the sequencer, which varies the original sequence at that position. A limited RNA pool is then transcribed from this set of DNA molecules with a photoreactive triphosphate replacing a specific triphosphate in the reaction mix (i.e., BrU for U). The partially randomized set of RNA molecules which contains the photoreactive nucleotides is mixed with a quantity of the target protein. Bound RNAs that have a photoreactive group adjacent to a reactive amino acid residue in the nucleoprotein complex form covalent crosslinks upon irradiation. RNAs that bind and crosslink are selected through several rounds of photoSELEX and separated away from RNAs that bind but do not crosslink.

Example 16. Methods for Modifying a Target Molecule.

In another embodiment of the method of the present invention, photoSELEX is applied to develop a ligand capable of modifying a target molecule. Under these circumstances, incorporation of a photoreactive group onto or into a ligand selected by photoSELEX or SELEX may modify the target in several ways such that the biological activity of the target molecule is modified. For example, the target molecule may be inactivated by photocrosslinked ligand. Mechanisms of inactivation include electron or hydrogen abstraction from the target molecule or radical addition to the target molecule that elicit a chemical modification. These different mechanisms may be achieved by changing the mode of irradiation.

A ligand selected through photoSELEX used as a diagnostic for a target molecule with ultraviolet (UV) light may also inactivate the same target in vivo if the

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source of irradiation is changed to X-rays or gamma rays. The resultant vinyl radical may work similarly to a hydroxyl radical, that is, by abstraction of hydrogen atoms from the binding domain of the target molecule.

5 X-ray irradiation of the R17 coat protein bound to radio-labelled IU- or BrU-substituted RNA hairpin sequences may result in the formation of a crosslink. The BrU or IU chromophore may also be excited to a higher energy state by X-ray irradiation resulting in the
10 formation of a vinyl radical (Mee (1987) in: *Radiation Chemistry: Principles and Applications* (Farhataziz and Rodgers, eds.), VCH Publishers, New York, pp. 477-499). The radical abstracts a hydrogen from the binding domain of the R17 coat protein, thereby reducing or inhibiting
15 its ability to bind the RNA ligand. Inactivation is tested by X-ray irradiation of the R17 coat protein in the presence and absence of substituted RNAs. The formation of crosslinked complexes is analyzed by PAGE. The effect of X-ray irradiation of RNA resulting in
20 modification of binding by modification of the protein domain is followed by nitrocellulose binding assay.

Example 17. Diagnostic Use of PhotoSELEX To Identify Unique Proteins Associated with Specific Disease Processes.

25 A goal of diagnostic procedures is to correlate the appearance of unique proteins with specific disease processes. Some of these correlations are obvious, e.g., after bacterial or viral infections, one can detect
30 antigens which are antigen specific or antibodies to such antigens not found in the blood of uninfected subjects. Less obvious correlations include the appearance in serum of α -foeto protein which is directly correlated with the presence of the most common form of testicular cancer.

35 The photoSELEX method may be applied to the

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discovery of heretofore unknown correlations between biological proteins and important human diseases. In one embodiment of the present invention, serum is taken from a patient with a disease, RNA ligands to all the proteins in the serum are produced and adsorbed to normal sera. RNA ligands to serum proteins may be identified through the SELEX method, with subsequent incorporation of photoreactive groups, or may be identified through photo-SELEX, initially selected from a candidate mixture of oligonucleotides containing one or more photoreactive groups. RNA ligands left unbound are those which specifically bind only unique proteins in the serum from patients with that disease. For example, RNA ligands are initially identified to a limited number of serum proteins (e.g., 11). The RNA ligands identified contain a modified NTP having a reversible or photoreactive functional group capable of crosslinking reversibly or non-reversibly with the target protein. Optionally, the presence of a cross-linked ligand to every protein may be verified. The RNA ligands are then removed and amplified. RNA is then transcribed for a second SELEX round. RNA is now bound to a large excess of 10 of the original 11 proteins, leaving an RNA ligand specific for the unique (11th) protein. This RNA is then amplified. This is a subtractive technique.

In one embodiment of the diagnostic method of the present invention, the method described above is used to identify a ligand to an abnormal protein, for example, an α -foeto protein. Sera from patients with important diseases is obtained and RNA ligands to all proteins present identified. The RNA ligands are adsorbed to normal sera, leaving an unbound ligand. The unbound ligand is both a potential diagnostic agent and a tool for identifying serum proteins specifically associated with a disease.

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Example 18. Method of Treating Disease by In Vivo Use of Photocrosslinking Nucleic Acid Ligand.

5 A nucleic acid ligand to a target molecule associated with a disease state is selected through the photoSELEX process (Example 11). The photoSELEX selected nucleic acid ligand may be introduced into a patient in a number of ways known to the art. For example, the non-halogenated photoSELEX ligand is cloned into stem cells which are transferred into a patient. The ligand may be
10 transiently or constitutively expressed in the patient's cells. IC administered to the patient is incorporated into the oligonucleotide product of the cloned sequence. Upon irradiation, the ligand is able to crosslink to the target molecule. Irradiation may include visible, 325
15 nm, 308 nm, X-ray, ultraviolet, and infrared light.

Alternatively, the photoSELEX ligand may be taken into a patient's cells as a double-stranded DNA which is transcribed in the cell in the presence of iodinated cytosine. Further methods of introducing the photoSELEX
20 ligand into a patient include liposome delivery of the halogenated ligand into the patient's cells.

Example 19. PhotoSELEX Ligands for Use in In Vitro Diagnostic, In Vivo Imaging and
25 Therapeutic Delivery.

PhotoSELEX may be used to identify molecules specifically associated with a disease condition and/or abnormal cells such as tumor cells. PhotoSELEX-identified oligonucleotides may be produced that react
30 covalently with such marker molecules.

In one embodiment of the present invention, the target for photoSELEX is the abnormal serum or tumor cell (e.g., the target mixture). A library candidate mixture of oligonucleotides is generated containing photoreactive
35 groups. Using one of the above-described photoSELEX

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protocols, oligonucleotides able to photocrosslink to the unique proteins in the abnormal serum or on the tumor cells are identified. Oligonucleotides able to crosslink to a marker protein on a tumor cell are useful as in vitro diagnostics or when coupled to enhancing agents for in vitro imaging. Further, oligonucleotides able to crosslink to a marker protein on a tumor cell may be used therapeutically, for example, as a method for immune activation, as a method of inactivation, or as a method of delivering specific target-active pharmaceutical compounds.

Example 20. PhotoSELEX and HIV-1 Rev.

At each position of the template deoxy-oligonucleotide synthesis, the nucleotide reagent ratio was 62.5:12.5:12.5:12.5. The nucleotide added in greater amount at each position corresponds to the nucleotide found in the 6a sequence (SEQ ID NO:5) at the same position.

Cloning and Sequencing procedure: RNA's isolated from each round were reverse transcribed to produce cDNA and PCR amplified producing a 111 bp fragment with unique BamHI and HindIII restriction sites at the ends. The phenol/CHCl₃ treated fragment and a pUC18 vector were digested together overnight with BamHI and HindIII at 37°C., phenol/ChCl₃ treated and precipitated. The digested vector and PCR product was ligated at room temperature for 4 hours and with T4 DNA ligase and transformed to competent DH5 α -F' cells which were then grown on ampicillin-containing LB plates. Individual colonies were grown overnight in LB- ampicillin media and plasmid was prepared using Wizard (Promega) plasmid preparation kit. Sequencing was performed utilizing a Sequenase (USB) kit.

Conditions for nitrocellulose filter binding

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selections: All rounds utilized approximately 20 nM RNA. Round 1 and 2: 6 nM Rev. Round 3: 3 nM Rev. Round 8: 1nM Rev. Round 9-10: 3 nM Rev. Binding reaction volumes ranged from 5 mls to 1 ml.

5 Conditions for crosslinking selections:

Approximately 50-100 nM of folded pool RNA was added to 0.2 (Rounds 4-6) or 0.5 (Round 7) μ M Rev, 1 μ M BSA in 1x BB (50mM TrisAc pH 7.7, 200mM KOAc, 10 mM DTT) on ice and incubated 5 minutes at 37°C. The samples were then
10 irradiated at 37°C, for 3 minutes at 308 nm by a XeCl excimer laser (round 4), 30 minutes at 325 nm by a HeCd laser (round 5), 10 minutes at 325 nm, (round 6), or 1 minute at 325 nm, (round 7). Approximately one-half of the sample was heated in 50% formamide, 40 ug tRNA at
15 90°C. for 4 minutes and separated by electrophoresis in an 8 percent polyacrylamide-/8 M urea gel.

The following procedure was utilized to elute crosslinked RNAs from acrylamide gels with approximately 80% recovery: The nucleoprotein containing gel slice was
20 crushed to a homogenous slurry in 1X PK buffer (100mM Tris-Cl pH 7.7, 50mM NaCl and 10mM EDTA). Proteinase K was added to 1 mg/ml concentration and incubated at 42°C for 30 minutes. 15 minute incubations at 42°C with increasing urea concentrations of approximately 0.7 M,
25 1.9 M, and 3.3M were performed. The resulting solution was passed through DMCS treated glass wool and 0.2 um cellulose acetate filter. The filtered solution was extracted twice with phenol/ CHCl_3 and then precipitated with a 1:1 volume mixture of EtOH:isopropanol.

30 The crosslinked band from each round was placed in scintillation fluid and counted in a Beckman LS-133 Liquid Scintillation System. The percent crosslinked =
cpms of crosslinked product from RNA + Rev after 4 minutes irradiation at 325 nm minus cpms in crosslinked
35 region for RNA only irradiated divided by total cpms.

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The fold increase in crosslinking is % R13 crosslinked divided by % D37 crosslinked.

Simultaneous selection for affinity and crosslinking using competitor tRNA was performed as follows. 10uM yeast tRNA was added to 0.5μM Rev, 1 μM BSA in 1x BB (50mM TrisAc (pH=7.5), 200mM KOAc, 10 mM DTT) and incubated 10 minutes on ice. 200,000 cpms (approximately 50-100 nM final concentration RNA) was added and incubated an additional 15 to 60 minutes on ice followed by 5 minutes at 37°C. The samples were then irradiated 4 minutes at 325 nm by a HeCd laser at 37°C. Approximately one-third of the sample was heated in 50% formamide, 40 ug tRNA at 90 C. for 4 minutes and separated by electrophoresis in an 8 percent polyacrylamide-8M urea gel.

The LI crosslinking RNA ligands form additional crosslinked product with a 4 minute 325 nm laser irradiation.

The template oligos used to produce the truncated RNA's are: PTS-1; 5'-TAATACGACTCACTATA-3', (SEQ ID NO:60) DNA-2; 5'-GAGTGGAAACACACGTGGTGTTC-CATACACCCTATAGTGAGTCGTATTA-3' (SEQ ID NO: 61), and DNA-24; 5'-AGGGTTAACAGGTGTGCCTGTTAATCCCCTATAGT-GAGTCGTATTA-3' (SEQ ID NO:62). PTS-1 was annealed with DNA-2 or DNA-24 to produce a template for T7 transcription.

To calculate the number of changes for individual molecules compared to 6a (SEQ ID NO:5), each was aligned to 6a for maximum similarity. Gaps are calculated as one change and truncated molecules were counted as unchanged. To calculate the average probability of finding molecules within each class; the average number of specific (s) and non-specific (ns) changes and unchanged (u) were calculated and used in the equation:

$(P) = (.125)^s (.375)^{ns} (.625)^u$. Class Ia (P)= 9×10^{-15} ; Ib (P)= 3×10^{-15} ; Ic (P)= 7×10^{-13} ; Id (P)= 3×10^{-15} ; Class

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II(P) = 2×10^{-14} . Since the starting population consists of 10^{14} molecules, sequences with (P) $< 10^{-14}$ will not be represented. (s) are those changes required to produce the uppercase, consensus nucleotides and (ns) are additional changes.

5 Trunc24 (SEQ ID NO:59) photo-independent crosslinking with HIV-1 Rev in the presence of human nuclear extracts was determined as follows: Trunc24 RNA, nuclear extracts, and Rev protein were combined and
10 incubated on ice for 10 min. Samples were mixed 1:1 with 8 M urea loading buffer and placed on a 7 M urea, 8% polyacrylamide gel for analysis, XL indicates the nucleoprotein complex, RNA indicates free trunc24 RNA.

15 Example 21. Primer Extension Inhibition Solution
SELEX.

Primer extension inhibition relies on the ability of a tightly bound target molecule to inhibit cDNA synthesis of high affinity oligonucleotides and results in
20 formation of an amplifiable cDNA pool corresponding to high affinity oligonucleotides and a non-amplifiable cDNA pool corresponding to low affinity oligonucleotides. Thus, the PCR step of solution SELEX acts as a partitioning screen between two cDNA pools. General
25 protocols for nucleic acid synthesis, primer extension inhibition and PCR are herein provided. Further, N-acryloylamino phenyl mercuric gel electrophoretic conditions for separation of selected nucleic acid ligands is described. The methods of cloning and
30 sequencing nucleic acid ligands is as described by Tuerk and Gold (1990) *supra*.

RNA Synthesis. The RNA candidate mixture was generated by incubating RNA polymerase and DNA templates. The reaction conditions are 8% polyethylene glycol 8000,
35 5 mM dithiothreitol, 40 mM Tris-HCl (pH 8.0), 12 mM

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MgCl₂, 1 mM spermidine, 0.002% Triton X-100, 2 mM nucleotide triphosphates, and 1 unit/ μ l RNA polymerase. Reactions are incubated at 37°C for 2 hours.

5 The transcription protocol may be used to generate RNAs with modified nucleotides. The transcription reaction may either be primed with a nucleotide triphosphate derivative (to generate a modified 5' end), modified nucleotides may be randomly incorporated into the nascent RNA chain, or oligonucleotides or their
10 derivatives ligated onto the 5' or 3' ends of the RNA product.

Primer Extension Inhibition. Primer extension inhibition is performed as described by Hartz et al. (1988) *supra*. Briefly, an oligonucleotide primer is
15 annealed to the 3' end of the oligonucleotides of the candidate mixture by incubating them with a 2-fold molar excess of primer at 65°C for 3 min in distilled water. The annealing reaction is cooled on ice, followed by the addition of 1/10 volume of 10X concentrated extension
20 buffer (e.g., 10 mM Tris-HCl (pH 7.4), 60 mM NH₄Cl, 10 mM Mg-acetate, 6 mM β -mercaptoethanol, and 0.4 mM nucleotide triphosphates). Primer extension is initiated by addition of polymerase and incubation at any of a variety of temperatures ranging between 0-80°C, and for times
25 ranging from a few seconds to several hours. In one embodiment of the method of the present invention, primer extension is first conducted in the presence of chain terminating nucleotide triphosphates such that low-affinity nucleic acids preferentially incorporate these
30 chain terminators. A second primer extension is then conducted after removing the target from high affinity nucleic acids and removing the chain terminating nucleotides triphosphates.

Polymerase Chain Reaction. The polymerase chain
35 reaction (PCR) is accomplished by incubating an

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oligonucleotide template, either single- or double-stranded, with 1 unit/ μ l thermal stable polymerase in buffer (50 mM KCl, 10 mM Tris-HCl (pH 8.6), 2.5 mM $MgCl_2$, 1.7 mg/ml BSA, 1 mM deoxynucleotide triphosphates, and 1 μ M primers). Standard thermal cycles are 95° C for 30 sec, 55°C for 30 sec, and 72°C for 1 min, repeated as necessary. One modification of the PCR protocol generates single-strand DNA by incubating either single- or double-stranded template with a single, elongated primer oligonucleotides and results in an elongated product. PCR preferentially amplifies the oligonucleotides rendered amplifiable in the primer extension steps described above.

(N-Acryloylamino)phenyl mercuric gel electrophoresis. Polyacrylamide gel electrophoresis using N-acryloylamine phenyl mercury (APM) was performed as described by Igloi (1988) Biochemistry 27:3842. APM was synthesized by mixing 8 ml of acetonitrile to 0.35 g of (p-aminophenyl)mercuric acetate at 0°C, followed by 2 ml of 1.2 M $NaHCO_3$. A total of 0.2 ml of acryloyl chloride was then added with vigorous stirring and the reaction incubated overnight at 4°C. The solid phase was collected by centrifugation and washed with water, dissolved by warming to 50°C in 8.5 ml of dioxane, followed by filtration to remove undissolved contaminants. APM crystals were formed upon standing at room temperature and the solid was washed again with water and dried under vacuum. APM was stored at 4°C. APM-polyacrylamide gels were prepared by addition of a appropriate aliquot of a 1 mg/ml solution of APM in formamide to a solution containing a given amount of acrylamide, bis(acrylamide), an urea in 0.1 M Tris-borate/EDTA (pH 8.3). Polymerization was initiated by addition of 0.5 ml of 1% ammonium persulfate and 7 μ l of TEMED per 10 ml of gel solution.

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Example 22. Enzymatic or Chemical Degradation Solution SELEX.

Enzymes or chemicals may be used to selectively degrade the pool of cDNA corresponding to low-affinity oligonucleotides. In one embodiment of the present invention, restriction enzymes are used to selectively degrade the cDNA pool corresponding to low-affinity oligonucleotides. A number of restriction enzymes have been identified that cleave single-stranded DNA. These enzymes cleave at specific sequences but with varying efficiencies.

Restriction enzyme digestion may be performed with a variety of sequence specific restriction endonucleases. Endonucleases that cleave single-stranded DNA include DdeI, HaeIII, HgaI, HinfI, HincPI, MnlI, PstI, and RsaI. These enzymes are used under standard conditions known to those skilled in the field of molecular biology. Double-stranded nucleic acids may also be cleaved using the proper combination of nucleic acid restriction sequences and site specific restriction nucleases.

The basic solution SELEX procedure is followed as described in the SELEX Patent Applications. The first cDNA extension is performed in the presence of four dNTPS, followed by removal of the target. The second cDNA extension is performed with modified nucleotides that are resistant to enzymatic cleavage by restriction endonucleases. The mixture of cDNA extension products is incubated with the appropriate restriction enzyme. The product of the first cDNA extension from free nucleic acid is cleaved to remove the primer annealing site, rendering this cDNA pool non-amplifiable by PCR. The efficiency of cleavage by restriction endonucleases may be improved using a hairpin at the restriction site (RS) to create a localized double-stranded region, as shown in Figure 24.

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Alternatively, the first cDNA extension product is rendered selectively degradable by other classes of enzymes by incorporation of modified nucleotides. For example, cDNA corresponding to low affinity ligands may be synthesized with nucleotides sensitive to uracil DNA glycosylase, while cDNA corresponding to high affinity ligands may incorporate resistant nucleotides.

Chemical degradation of cDNA corresponding to low affinity ligands can be accomplished by incorporation of 7-methylguanosine, 5-bromouracil, or 5-iodouracil as described using piperidine or photodegradation (Sasse-Dwight and Gralla (1991) Methods Enzymol. 208:146; Aigen and Gumpert (1991) Methods Enzymol. 208:433; Hockensmith et al. (1991) Methods Enzymol. 208:211).

Example 23. Solution SELEX Followed by Affinity Chromatography.

Selective removal of either the first or second cDNA extension products may be achieved through affinity chromatography. Removal of the first cDNA extension product preferentially removes the cDNA pool corresponding to free or low-affinity nucleic acids. Removal of the second cDNA extension product preferentially retains cDNA corresponding to the high-affinity ligand. This strategy relies on the incorporation of modified nucleotides during cDNA synthesis.

Selective Removal of First Extension Product.

Following the basic solution SELEX protocol, the first cDNA extension is performed in the presence of modified nucleotides (e.g., biotinylated, iodinated, thiolabelled, or any other modified nucleotide) that allow retention of the first cDNA pool on an affinity matrix (Fig. 25). The target is then removed and the second cDNA extension performed in the presence of non-modified nucleotides.

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The cDNAs that have incorporated the modified nucleotides may be removed by affinity chromatography using a column containing the corresponding affinity ligand. The cDNA pool corresponding to nucleic acids with high affinity for the target remain and are then amplified by PCR.

Selective Removal of the Second Extension Product.

Following the basic protocol, the first cDNA extension is performed in the presence of four dNTPs, and the second cDNA extension is performed in the presence of modified nucleotides (e.g., biotinylated, iodinated, thiolabelled, or any other modified nucleotide) that allow retention of the second cDNA pool on an affinity matrix as described above.

Incorporation of Specific Sequences for Annealing to An Affinity Matrix. In an alternate embodiment of the method of the present invention, a special sequence can also be selectively incorporated for annealing to an affinity matrix. Thus, either first or second synthesis cDNAs can be retarded and purified on commercially obtainable matrices as desired.

Example 24. Exonuclease Inhibition Solution SELEX.

Exonuclease inhibition may be used to isolate double-stranded ligands. Double-stranded nucleic acid ligands tightly bound to the target molecule will inhibit exonuclease hydrolysis at the 3' edge of the binding site. This results in a population of nucleic acid molecules resistant to hydrolysis that also contain a long single-stranded 5' overhang and a central base paired region (see Fig. 26). This nucleic acid molecule is a substrate for any polymerase, and incubation with polymerase will generate the double-stranded starting material. This molecule is amplified by PCR. Members of the nucleic acid candidate mixture that are not tightly bound to the target molecule are digested during the

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initial exonuclease step.

3'→5' hydrolysis of double-stranded nucleic acid is accomplished by incubation with any double-stranded specific 3'→5' exonuclease. Exonuclease III specifically hydrolyzes double-stranded DNA 3'→5' and is active in a variety of buffers, including 50 mM Tris-HCl (pH 8.0), 5 mM MgCl₂, 10 mM β-mercaptoethanol at 37°C.

Example 25. Solution SELEX Method for Isolating Catalytic Nucleic Acids.

Solution SELEX may be used to isolate catalytic nucleic acid sequences. This embodiment of the invention takes advantage of a linear to circular transformation to sort non-catalytic nucleic acids from catalytic nucleic acids.

As shown in Figure 27, the PCR step may be exploited to screen the nucleic acid candidate mixture for catalytic members. Catalytic nucleic acids that either self-circularize, or alter their 5' or 3' ends to allow circularization with ligase, will amplify during PCR. The figure illustrates circle formation by catalytic members of the candidate mixture; the non-catalytic oligonucleotide members of the candidate mixture will remain linear. After circularization, the candidate mixture is incubated with a primer that anneals to the extreme 5' end. In this embodiment of the invention, only the circular oligonucleotide members will generate cDNA and be amplified during the PCR step.

This strategy isolates nucleic acids that either directly catalyze self-circularization or that modify their own ends so that the amplifiable form may be generated by incubation with ligase. As shown in Figure 27, the unusual interaction of the cDNA primer with the 5' end of the oligonucleotides of the candidate mixture permits amplification of only the circular molecules. In

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a further embodiment of the method of the present invention, this strategy is modified to allow isolation of catalytic nucleic acids that catalyze novel reactions.

5 Example 26. Automation of Solution SELEX.

 The automated solution SELEX protocol represents a modification of the technology used in the automated DNA synthesizer. The nucleic acid candidate mixture is attached to a solid support by either the biotin/avidin
10 interaction or a variety of covalent chromatographic techniques (e.g., the condensation of modified nucleotides onto maleimide or citraconic anhydride supports). The bound nucleic acid candidate mixture provides a good substrate for targeting binding, and the
15 column allows use of a single reaction vessel for the SELEX procedure. Primer extension inhibition is used to physically sort low and high affinity ligands. Low affinity nucleic acids may be degraded by incorporation of modified nucleotides during the first cDNA extension
20 step that renders the cDNA degradable as described in Example 22, while high affinity ligands are copied into non-degradable cDNA and amplified by PCR. For additional rounds of solution SELEX, the PCR generated candidate mixture is purified or is transcribed into RNA and
25 reattached to a second solid support, in the same or a new reaction vessel as desired. The process is repeated as necessary.

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(1) GENERAL INFORMATION:

- (A) TELEPHONE: (303) 793-3333
(B) TELEFAX: (303) 793-3433

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- (2) INFORMATION FOR SEQ ID NO:1:
(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 19 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

 (ix) FEATURE:
 (D) OTHER INFORMATION: U at position 13 is 5-bromouracil

 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:
GGGAGCGAGC AAUAGCCGC 19
- (2) INFORMATION FOR SEQ ID NO:2:
(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 19 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

 (ix) FEATURE:
 (D) OTHER INFORMATION: U at position 13 is 5-iodouracil

 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:
GGGAGCGAGC AAUAGCCGC 19
- (2) INFORMATION FOR SEQ ID NO:3:
(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 19 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

 (ix) FEATURE:
 (D) OTHER INFORMATION: U at position 13 has hydrogen molecule attached

 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:
GGGAGCGAGC AAUAGCCGC 19
- (2) INFORMATION FOR SEQ ID NO:4:
(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 44 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

 (ix) FEATURE:
 (D) OTHER INFORMATION: all U are 5-iodouracil

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- (2) INFORMATION FOR SEQ ID NO:10:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 37 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:
CUGUGGAUUA ACAGGCACAC CUGUUAACCG UGUACCU 37
- (2) INFORMATION FOR SEQ ID NO:11:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 37 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:
CUGUGGAUUA ACAGGCACAC CUGUUAACCG UGUACCC 37
- (2) INFORMATION FOR SEQ ID NO:12:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 36 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:
AGACGAUUA CAUCCACGGA UGUUAACGCG CUAGAA 36
- (2) INFORMATION FOR SEQ ID NO:13:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 37 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:
AAGACGAUUA ACAACACGU UGUUAACGC AACACCU 37
- (2) INFORMATION FOR SEQ ID NO:14:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 36 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:
GAUUGGAUUA ACAGGCACCC CUGUUAACCU ACCACU 36
- (2) INFORMATION FOR SEQ ID NO:15:
(i) SEQUENCE CHARACTERISTICS:

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- (A) LENGTH: 37 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:
AGGAGGAUUA ACAACAAAGG UUGUUAACCC CGUACCA

37

- (2) INFORMATION FOR SEQ ID NO:16:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 34 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:
UGAAGGAUUA ACAACUAAUG UUGUUAACCA UGUA

34

- (2) INFORMATION FOR SEQ ID NO:17:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 37 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:
UGAGGAUUA ACAGGCACAC CUGCUAACCG UGUACCC

37

- (2) INFORMATION FOR SEQ ID NO:18:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 37 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:
AUGUGGCUUA ACAAGUACGC UUGUUAACCC AAAACG

37

- (2) INFORMATION FOR SEQ ID NO:19:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 35 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:
AGGACGAUGA ACAAACACGU UUGUUCACGC CAUGC

35

- (2) INFORMATION FOR SEQ ID NO:20:
- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 38 base pairs
 - (B) TYPE: nucleic acid

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(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

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(2) INFORMATION FOR SEQ ID NO:25:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 37 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

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- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:
CAGUGUAUGA AACACCACGU GUGUUUCCAC UGUACCU 37
- (2) INFORMATION FOR SEQ ID NO:26:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 35 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:
CAGUGUAUGA AACAAACACGU UUGUUUCCAC UGCCU 35
- (2) INFORMATION FOR SEQ ID NO:27:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 35 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:
GAGUGUAUGA AACAAACACGU UUGUUUCCAC UCCCU 35
- (2) INFORMATION FOR SEQ ID NO:28:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 35 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:
GAGUGUAUGA AACAAACACGU UUGUUUCCAC UGUCU 35
- (2) INFORMATION FOR SEQ ID NO:29:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 35 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:
GAUUGUAUGA AACAAACGUGU UUGUUUCCAC UCCCU 35
- (2) INFORMATION FOR SEQ ID NO:30:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 35 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:
GAAUGUAUGA AACAAACACGU UUGUUUCCAC UGCCU 35

00827"872260

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- (2) INFORMATION FOR SEQ ID NO:31:
(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 37 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:
GAUUGGACUU AACAGACACC CCUGUUAACC UACCACU 37
- (2) INFORMATION FOR SEQ ID NO:32:
(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 34 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:
UGCGACAGUU AGAAACACGA UUGUUUACUG UAUG 34
- (2) INFORMATION FOR SEQ ID NO:33:
(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 36 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:
UACAGGCUUA AGAAACACGU UUGUUAACCA ACCCCU 36
- (2) INFORMATION FOR SEQ ID NO:34:
(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 36 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:
UCGAGCAGUG UGAAACACGA UUGUGUUUCC UGCUCA 36
- (2) INFORMATION FOR SEQ ID NO:35:
(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 36 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:
UGAUGCCUAG AGAAACACAU UAGUGUUUCC CUCUGU 36
- (2) INFORMATION FOR SEQ ID NO:36:
(i) SEQUENCE CHARACTERISTICS:

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- (2) INFORMATION FOR SEQ ID NO:41:
 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 37 base pairs
 (B) TYPE: nucleic acid

- (C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:
AGGAACCUAG AAACACACAG UGUUCCCTUC UGCCCAC

37

(2) INFORMATION FOR SEQ ID NO:42:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 37 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:
GCCUGCAUGG AUUAACACGU AUGUGUUAAC CGACUCC

37

(2) INFORMATION FOR SEQ ID NO:43:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 37 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:
UGAAACACUG AGAAACACGU GUUCCCCUUG GUGUGAU

37

(2) INFORMATION FOR SEQ ID NO:44:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 36 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:
AGGAACCUCA AGCCGCCCU AGAACACUCG GCACCU

36

(2) INFORMATION FOR SEQ ID NO:45:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 37 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:45:
AGGAACCUCA AGAAAGCCCC UGAAACACUC GAAGCCU

37

(2) INFORMATION FOR SEQ ID NO:46:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 37 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:46:
AGGAACCUCA AGAAACCCCC UGAAACACUC AUUACCG 37

- (2) INFORMATION FOR SEQ ID NO:47:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 37 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:47:
AGGAACCUCA AGAAAUCCGA ACGACAACCC UACACCU 37

- (2) INFORMATION FOR SEQ ID NO:48:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 36 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:48:
AGGAACCUCA AGAAACCCCG CCACGGACCC CAACCA 36

- (2) INFORMATION FOR SEQ ID NO:49:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 37 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:49:
GGGAACCUCA AUAAUCACGC ACGCAUACUC GGCAUCU 37

- (2) INFORMATION FOR SEQ ID NO:50:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 34 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:50:
GGGAACCUCA AGAGACCCGA CAGGAUACUC GGAC 34

- (2) INFORMATION FOR SEQ ID NO:51:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 37 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:51:
AAGUGGAACC UCAAUCCCGU AAGAAGAUCC UGUACCU 37

008277" 8722260

- (2) INFORMATION FOR SEQ ID NO:52:
 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 37 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:52:
 AUGUGCAUAG AGAUGUACAU AUGAACCUC AGUAGAG 37
- (2) INFORMATION FOR SEQ ID NO:53:
 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 37 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:53:
 UCAUGCAUAG GCAUAGGCAG AUGGAACCUC AGUAGCC 37
- (2) INFORMATION FOR SEQ ID NO:54:
 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 37 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:54:
 AUGUGCAACA AGGCGCAJGG AUAAGGAACC UCGAAGU 37
- (2) INFORMATION FOR SEQ ID NO:55:
 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 37 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:55:
 GAGUACAGCA CGCAACACGU ACGGGGAACC UCAAAGU 37
- (2) INFORMATION FOR SEQ ID NO:56:
 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
 (ix) FEATURE:
 (D) OTHER INFORMATION: N at positions 1 and 20
 indicates 1-2 complementary base pairs
 (ix) FEATURE:

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- (D) OTHER INFORMATION: N at position 3 indicates 1 or 3 nucleotides
- (ix) FEATURE:
(D) OTHER INFORMATION: N at positions 10 and 12 indicates 1-4 complementary base pairs
- (ix) FEATURE:
(D) OTHER INFORMATION: N at position 11 indicates 4 or 5 nucleotides
- (ix) FEATURE:
(D) OTHER INFORMATION: U is iodouracil
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:56:
NGNKDAACAN NNUGUUHMCN 20
- (2) INFORMATION FOR SEQ ID NO:57:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 23 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (ix) FEATURE:
(D) OTHER INFORMATION: U is iodouracil
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:57:
GGAACCUCAA UUGAUGGCCU UCC 23
- (2) INFORMATION FOR SEQ ID NO:58:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 32 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (ix) FEATURE:
(D) OTHER INFORMATION: U is iodouracil
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:58:
GGGUGUAUGA AACACCACGU GUGUUUCCAC UC 32
- (2) INFORMATION FOR SEQ ID NO:59:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 29 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:59:
GGGGAUUAAC AGGCACACCU GUUAACCCU 29

D0323748.1.2300

-89-

- (2) INFORMATION FOR SEQ ID NO:60:
(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 17 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:60:
TAATACGACT CACTATA

17

- (2) INFORMATION FOR SEQ ID NO:61:
(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 49 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:61:
GAGTGGAAC ACACGTGGTG TTTCATACAC CCTATAGTGA GTCGTATTA

49

- (2) INFORMATION FOR SEQ ID NO:62:
(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 46 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:62:
AGGGTTAACA GGTGTGCCTG TTAATCCCCT ATAGTGAGTC GTATTA

46

- (2) INFORMATION FOR SEQ ID NO:63:
(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 13 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:63:
CTAGAGCCCG GGC

13

- (2) INFORMATION FOR SEQ ID NO:64:
(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 44 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:64:
CTAGAGCCCG GGCGAGCTCG AATTCATGGG TAATCCTCAT GTTC

44

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CLAIMS:

1. A method for identifying nucleic acid ligands of a target molecule from a candidate mixture of nucleic acids, said method comprising:
- 5 a) preparing a candidate mixture of nucleic acids, said nucleic acids containing photoreactive groups;
- b) contacting said candidate mixture with said target molecule, wherein nucleic acid sequences having increased affinity to the target molecule relative to the candidate mixture form nucleic acid-target molecule complexes;
- 10 c) irradiating said candidate mixture, wherein said nucleic acid-target molecule complexes photocrosslink;
- 15 d) partitioning the crosslinked nucleic acid-target molecule complexes from free nucleic acids in the candidate mixture; and
- e) identifying the nucleic acid sequences that photocrosslinked to the target molecule.
- 20
2. The method of claim 1 further comprising the step:
- f) repeating steps b) through d); and
- 25 g) amplifying the nucleic acids that photocrosslinked to the target molecule to yield a mixture of nucleic acids enriched in sequences that are capable of photocrosslinking the target molecule.
3. The method of claim 1 wherein said photoreactive groups are selected from the group consisting of 5-bromouracil, 5-iodouracil, 5-bromovinyluracil, 5-iodovinyluracil, 5-azidouracil, 4-thiouracil, 5-bromocytosine, 5-iodocytosine, 5-bromovinylcytosine, 5-iodovinylcytosine, 5-azidocytosine,
- 30
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8-azidoadenine, 8-bromoadenine, 8-iodoadenine, 8-
azidoguanine, 8-bromoguanine, 8-iodoguanine, 8-
azidohypoxanthine, 8-bromohypoxanthine, 8-
iodohypoxanthine, 8-azidoxanthine, 8-bromoxanthine, 8-
5 iodoxanthine, 5-bromodeoxyuracil, 8-bromo-2'-
deoxyadenine, 5-iodo-2'-deoxyuracil, 5-iodo-2'-
deoxycytosine, 5-[(4-azidophenacyl)thio]cytosine, 5-[(4-
azidophenacyl)thio]uracil, 7-deaza-7-iodoadenine, 7-
deaza-7-iodoguanine, 7-deaza-7-bromoadenine, and 7-deaza-
10 7-bromoguanine.

4. The method of claim 1 wherein said target
molecule is a protein and removal of the target protein
from the nucleic acid-target molecule complex is achieved
15 by proteolytic digestion.

5. The method of claim 1 wherein said nucleic
acid ligand is capable of modifying the biological
activity of said target molecule.
20

6. A method for identifying photocrosslinking
nucleic acid ligands of a target molecule from a
candidate mixture of nucleic acids, said method
comprising:

25 a) preparing a candidate mixture of nucleic
acids;

b) contacting said candidate mixture with said
target molecule, wherein nucleic acid sequences having
increased affinity to the target molecule relative to the
candidate mixture form nucleic acid-target molecule
30 complexes;

c) partitioning the increased affinity nucleic
acids from the remainder of the candidate mixture; and

d) amplifying the increased affinity nucleic
35 acids to yield a ligand-enriched mixture of nucleic

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acids, whereby nucleic acid ligands of the target molecule may be identified;

e) incorporating photoreactive groups into said increased affinity nucleic acids;

5 f) repeating step b)

g) irradiating said increased affinity nucleic acids, wherein said nucleic acid-target molecule complexes photocrosslink;

h) repeating step d) and e).

10

7. A method for identifying a disease comprising producing a nucleic acid ligand by the method of claim 1 to a target molecule specifically associated with said disease.

15

8. A method of treating a disease comprising:

a) identifying a nucleic acid ligand to a target molecule associated with a disease through the method of claim 1;

20 b) introducing a nucleic acid ligand in a
 patient;

c) administering photoreactive molecules wherein said expressed nucleic acid ligand contains photoreactive groups; and

25 d) irradiating said patient, wherein said
nucleic acid ligand crosslinks a target molecule.

9. A method for identifying nucleic acid ligands
able to crosslink a target molecule comprising the method
30 of claim 1 followed by steps b) through d) of claim 6.

10. The method of claim 6 wherein step a) is conducted with a candidate mixture of nucleic acids containing photoreactive groups.

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[illegible]

5 12. The nucleic acid ligands of claim 11 further comprising one or more of the photoreactive groups selected from the group set forth in claim 3.

14. The nucleic acid ligands of claim 13 further
comprising one or more of the photoreactive groups
15 selected from the group set forth in claim 3.

20

16. The nucleic acid ligands of claim 15 further comprising one or more of the photoreactive groups selected from the group set forth in claim 3.

30 a) preparing a candidate mixture of nucleic acids;

 b) contacting said candidate mixture with the target molecule, wherein nucleic acids having increased affinity to the target molecule form nucleic acid-target complexes;

35 c) partitioning the increased affinity nucleic

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acids from the remainder of the candidate mixture, said partitioning step resulting in two differentiable nucleic acid pools; and

5 d) amplifying the increased affinity nucleic acids to yield a ligand-enriched mixture of nucleic acids.

18. The method of claim 17 wherein step c) consists of

10 (i) a first cDNA extension with a nucleic acid polymerase such that full length cDNA is not obtained from said increased affinity nucleic acids forming nucleic acid-target complexes; ,

(ii) removal of the target molecule; and

15 (iii) a second cDNA extension with a nucleic acid polymerase, wherein cDNA is synthesized from said increased affinity nucleic acids.

19. The method of claim 18, wherein said nucleic acid polymerase is selected from the group consisting of
20 DNA polymerase, RNA polymerase, reverse transcriptase, and Q β -replicase.

20. The method of claim 18, wherein said first
25 cDNA extension step is performed in the presence of chain terminating nucleotide triphosphates and said second cDNA extension step is performed in the absence of chain terminating nucleotide triphosphates, wherein only the cDNA product from the increased affinity oligonucleotide
30 is amplifiable by PCR.

21. The method of claim 18, wherein said first
cDNA extension step is performed in the presence of four dNTPs, followed by removal of said target, and said
35 second cDNA extension step is performed in the presence

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of modified nucleotides resistant to enzymatic cleavage by restriction enzymes, single- or double-stranded nucleases, or uracil DNA glycosylase, and incubation of the cDNA products with a nuclease enzyme.

5

22. The method of claim 18, wherein said first cDNA extension step is performed in the presence of modified nucleotides that allow retention of the cDNA product on an affinity matrix, and said second cDNA extension step is performed in the presence of the four dNTPs, wherein cDNA synthesized from free or low affinity oligonucleotides is removed by retention on an affinity matrix.

10

15

23. The method of claim 18, wherein said first cDNA extension step is performed in the presence of four dNTPs, and said second cDNA extension step is performed in the presence of modified nucleotides that allow retention of the cDNA product on an affinity matrix, wherein cDNA synthesized from increased affinity oligonucleotides is removed by retention on an affinity matrix.

20

24. A method for identifying nucleic acid ligands from a candidate mixture of nucleic acids, said nucleic acid ligands being a ligand of a given target molecule comprising:

25

a) preparing a candidate mixture of nucleic acids;

30

b) contacting said candidate mixture with the target molecule, wherein nucleic acids having increased affinity to the target molecule form nucleic acid-target complexes;

35

c) partitioning the increased affinity nucleic acids from the remainder of the candidate mixture, said

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-96-

partitioning step comprising

(i) a first cDNA extension with a nucleic acid polymerase in the presence of dNTPs sensitive to chemical cleavage;

5 (ii) removal of the target molecule;

(iii) a second cDNA extension with a nucleic acid polymerase in the presence of modified nucleotides resistant to chemical cleavage;

10 (iv) incubation of said first and second cDNA extension products with a nucleotide degrading chemical; and

d) amplifying the increased affinity nucleic acids to yield a ligand-enriched mixture of nucleic acids.

15

25. A method to isolate single-stranded nucleic acids comprising

a) preparing a candidate mixture of nucleic acids;

20

b) contacting said candidate mixture with the target molecule, wherein nucleic acids having increased affinity to the target molecule form nucleic acid-target complexes;

25

c) partitioning the increased affinity nucleic acids from the remainder of the candidate mixture, said partitioning step including primer extension inhibition wherein two differentiable cDNA pools are generated; and

d) amplifying the increased affinity nucleic acids comprising

30

(i) amplifying the increased affinity nucleic acid ligands with a 5' PCR primer, wherein a ligand-enriched mixture of truncated, double-stranded nucleic acids is produced;

35

(ii) asymmetric amplification of the truncated, double-stranded nucleic acids with a

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-97-

second 5' primer, wherein a ligand-enriched elongated, single-stranded nucleic acid mixture is produced.

5 26. A method for identifying double-stranded nucleic acid ligands from a candidate mixture of nucleic acids, said nucleic acid ligands being a ligand of a given target molecule comprising:

10 a) preparing a candidate mixture of nucleic acids;

 b) contacting said candidate mixture with the target molecule, wherein nucleic acids having increased affinity to the target molecule form nucleic acid-target complexes;

15 c) partitioning the increased affinity nucleic acids from the remainder of the candidate mixture, said partitioning step comprising

 (i) incubating with exonuclease enzyme wherein full length double-stranded nucleic acids not forming nucleic acid-target complexes are degraded and double-stranded nucleic acids forming nucleic acid-target complexes are partially protected from degradation;

20 (ii) removing said exonuclease enzyme and said target;

25 (iii) extending said double-stranded nucleic acids with polymerase, wherein double-stranded nucleic acid ligand-enriched candidate mixture is regenerated.

30

 27. A method for identifying nucleic acids with catalytic activity from a candidate mixture of nucleic acids, comprising:

35 a) preparing a candidate mixture of nucleic acids;

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b) contacting said candidate mixture with the target molecule, wherein nucleic acids having increased affinity to the target molecule form nucleic acid-target complexes;

5 c) partitioning the increased affinity nucleic acids from the remainder of the candidate mixture, said partitioning step comprising

(i) annealing an extension primer to the extreme 5' end of said nucleic acids;

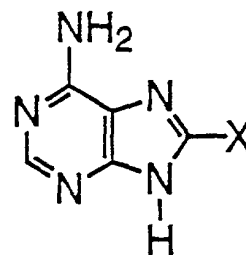
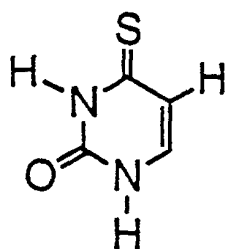
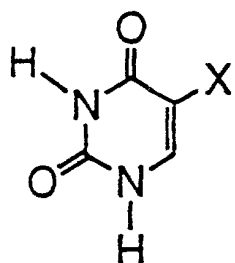
10 (ii) performing a cDNA extension; and

d) amplifying the full length cDNA to yield a second mixture of nucleic acids enriched for catalytic nucleic acids.

15 28. An automated method of identifying nucleic acid ligands comprising the method of claim 17 wherein said nucleic acids are covalently attached to an affinity column and said increased affinity nucleic acids are reattached to the affinity column, wherein all steps are
20 performed in a single reaction vessel.

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X=Br, 5-bromouracil 4-thiouracil X=N₃, 8-azidoadenine
 X=I, 5-iodouracil
 X=N₃, 5-azidouracil X=Br, 8-bromoadenine

FIG. 1

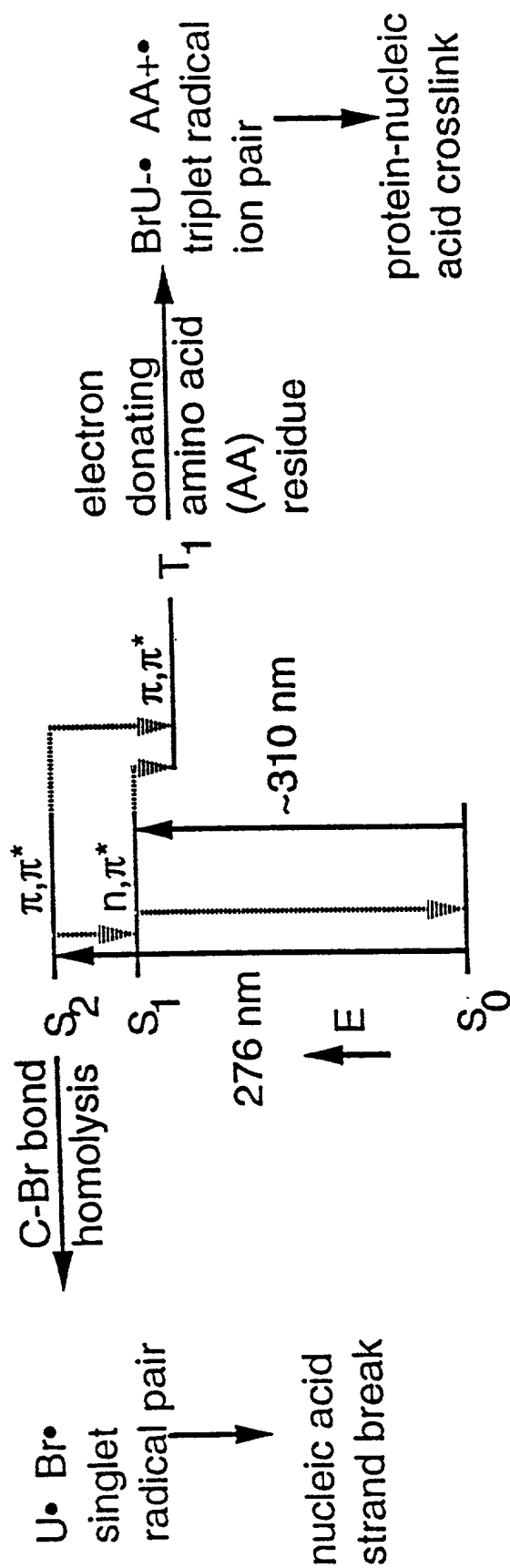


FIG. 2

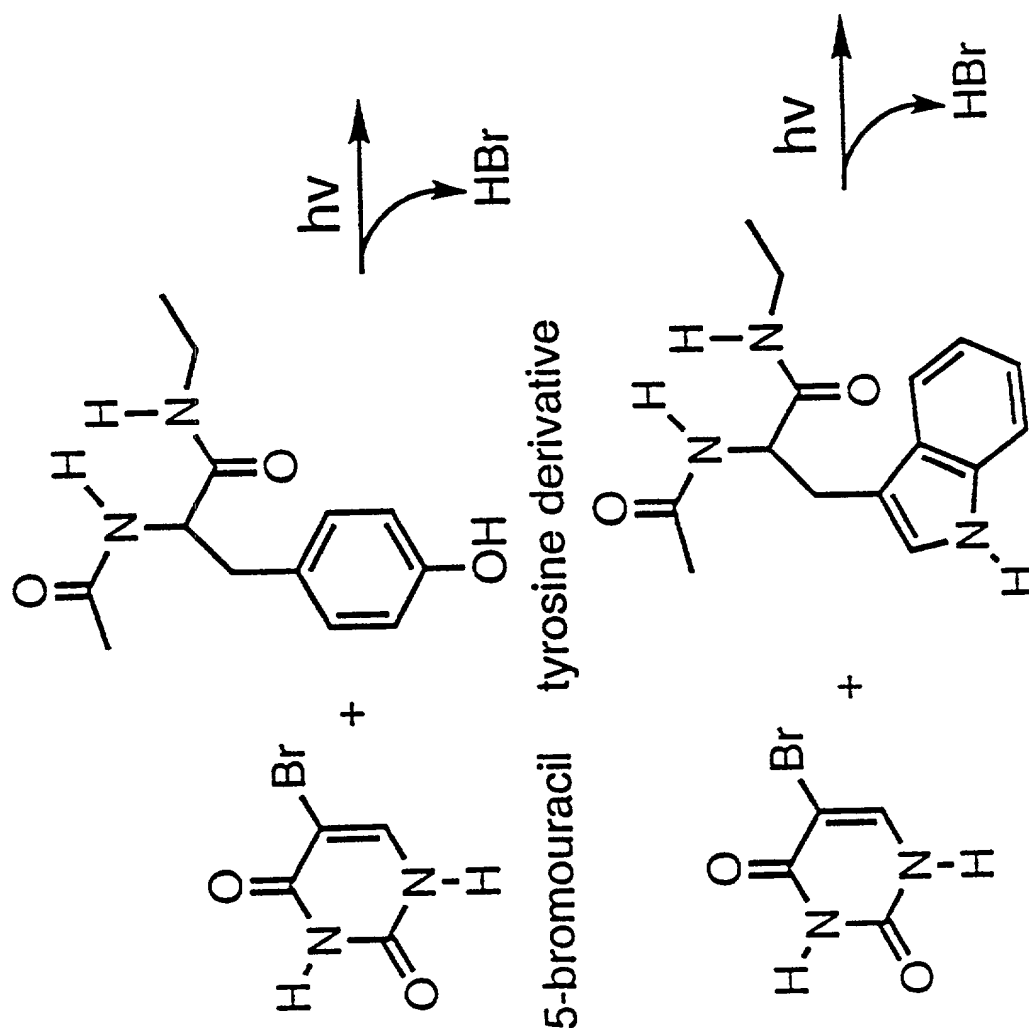
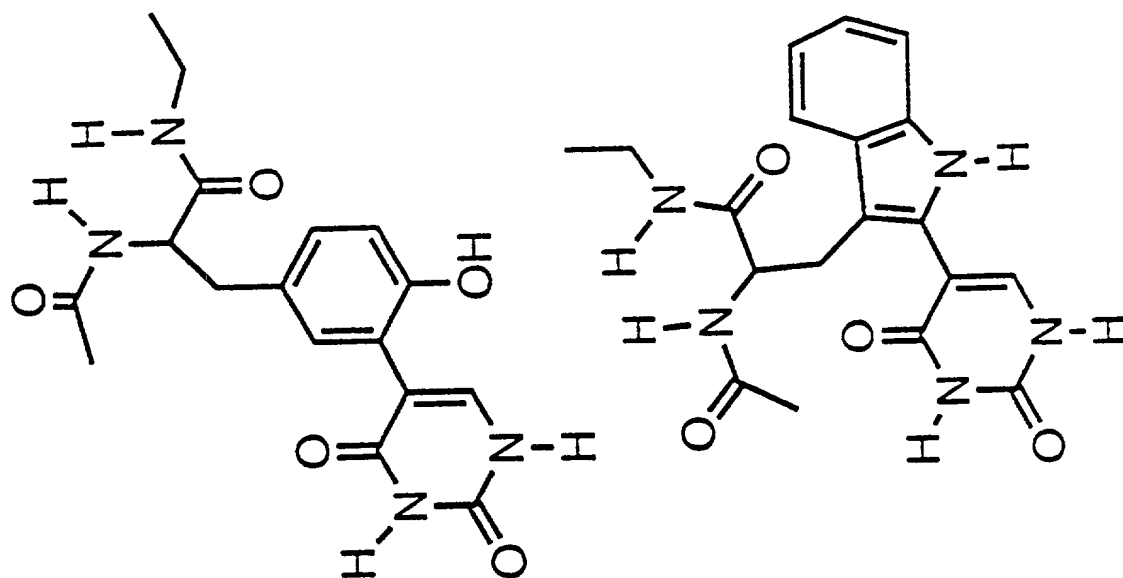


FIG. 3

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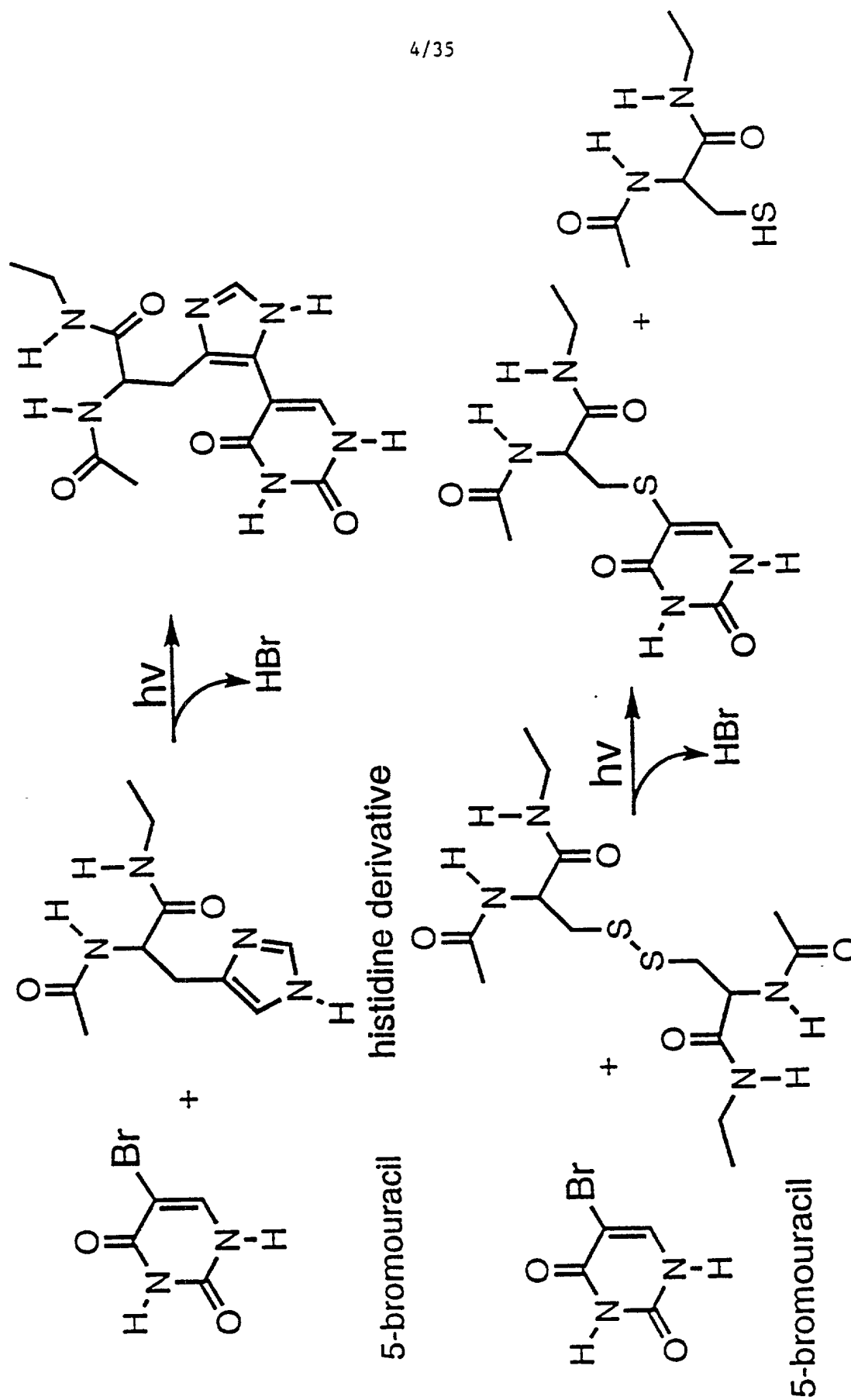


FIG. 3 (CONT'D)

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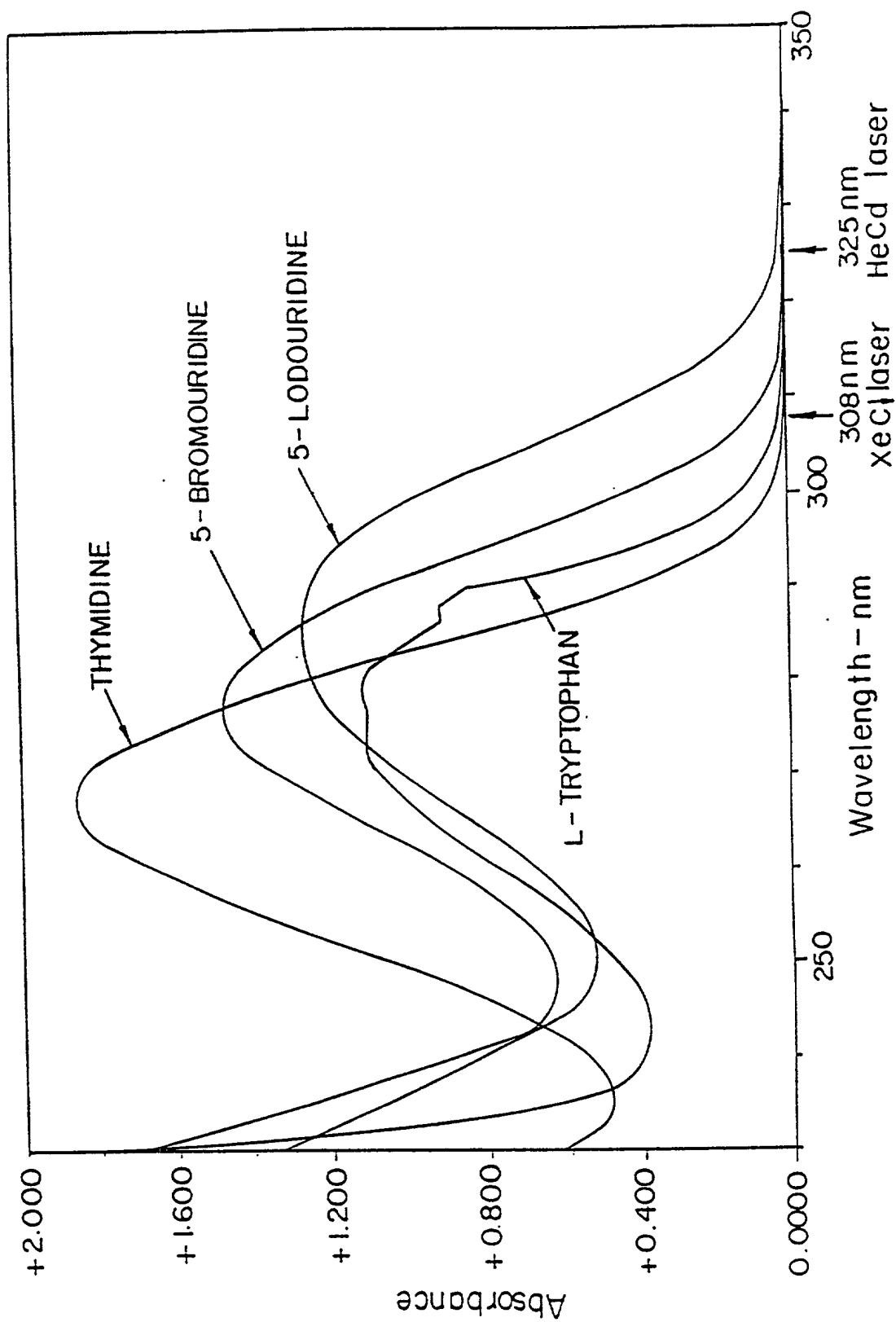


FIG.4

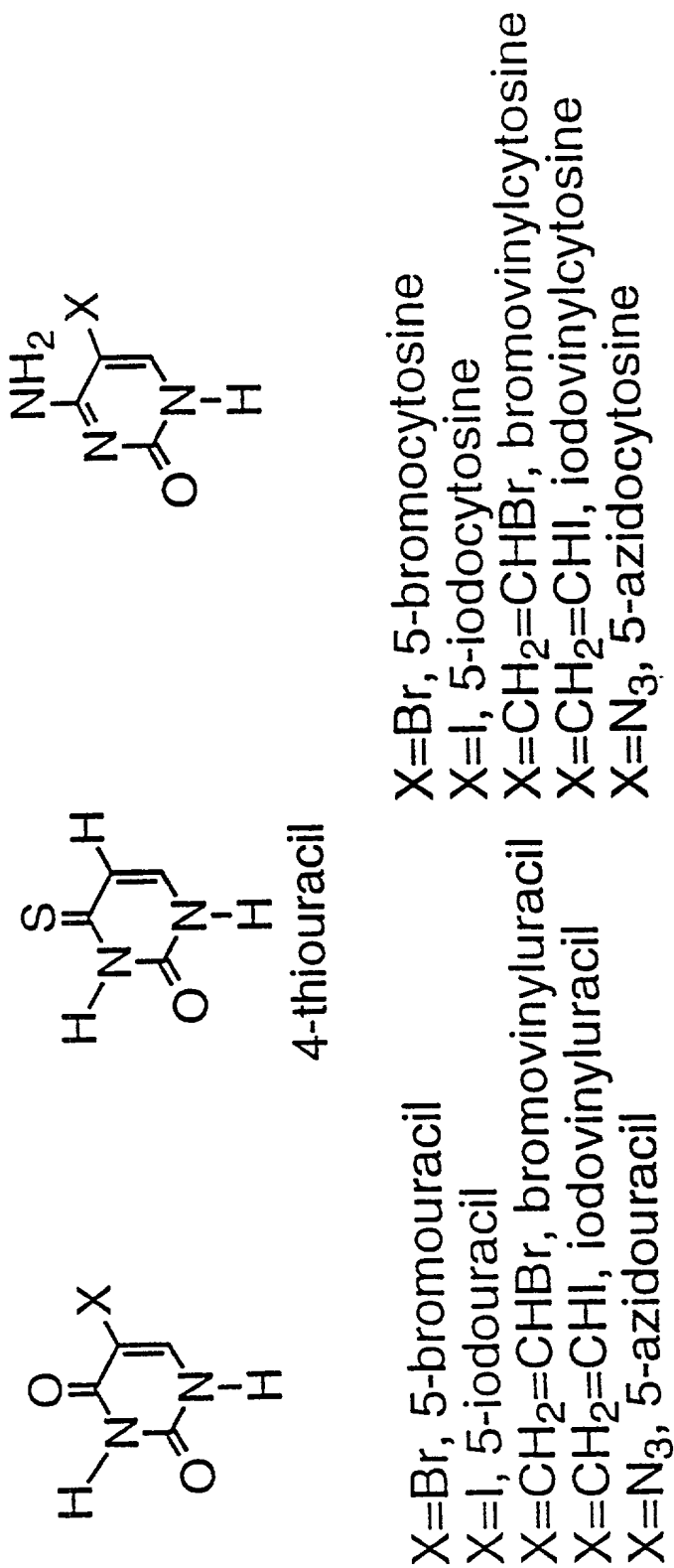


FIG. 5

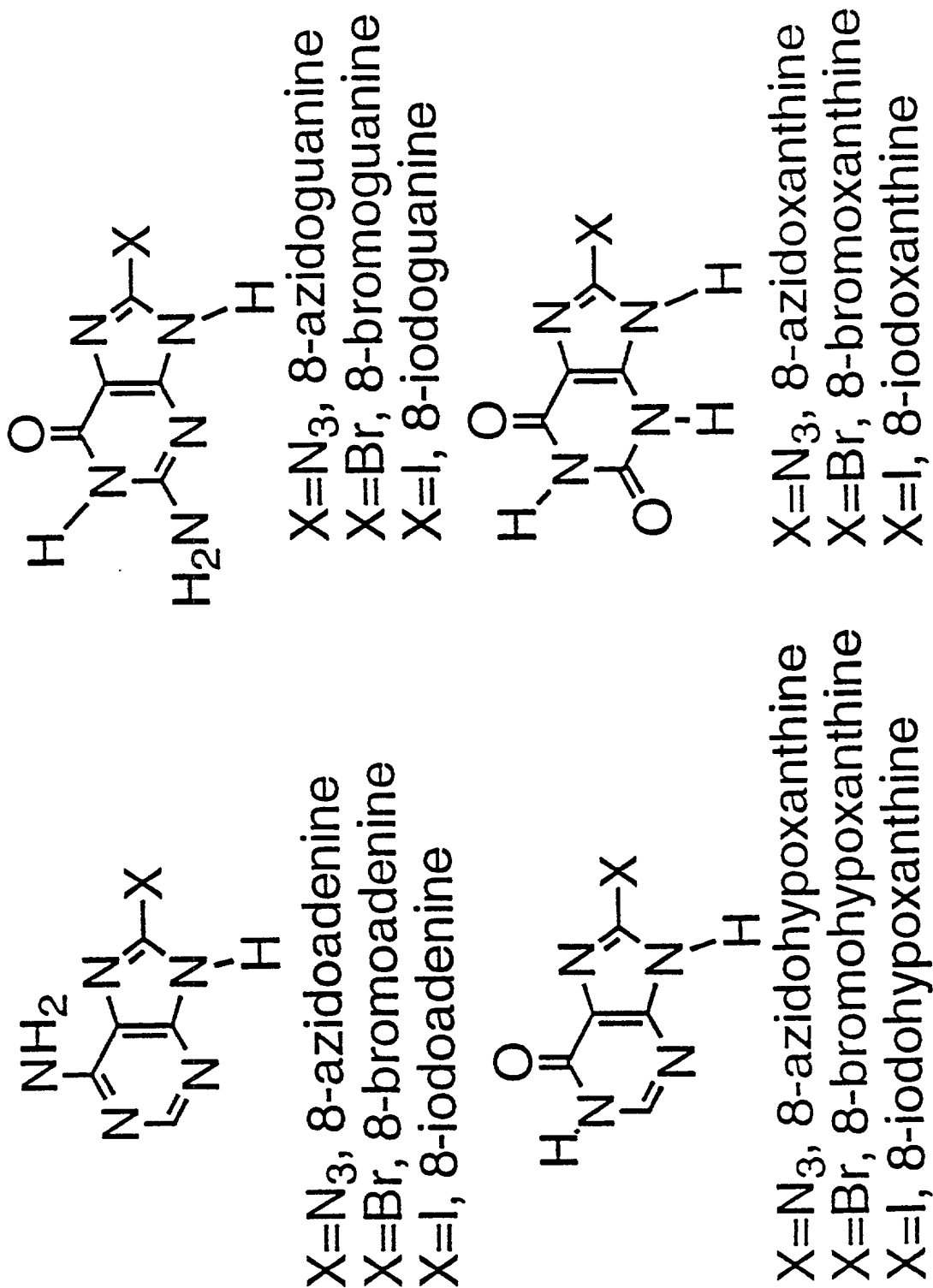


FIG. 5 (CONT'D)

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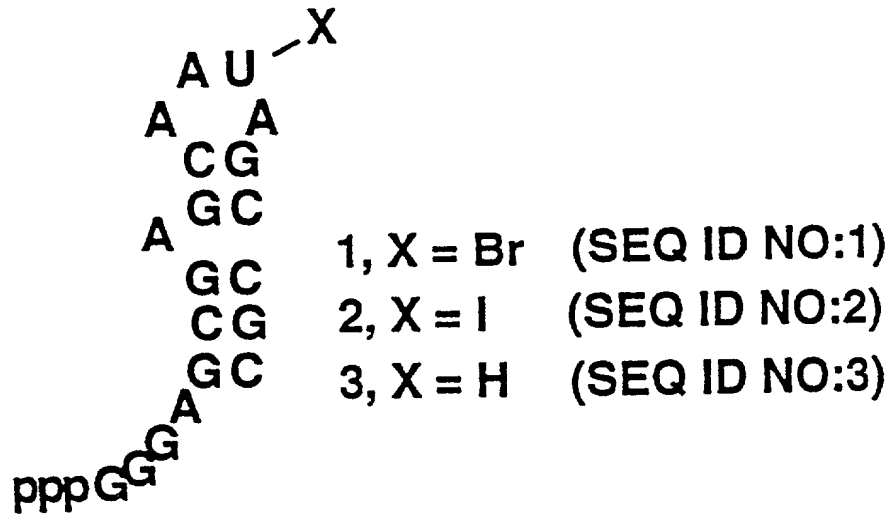


FIG. 6

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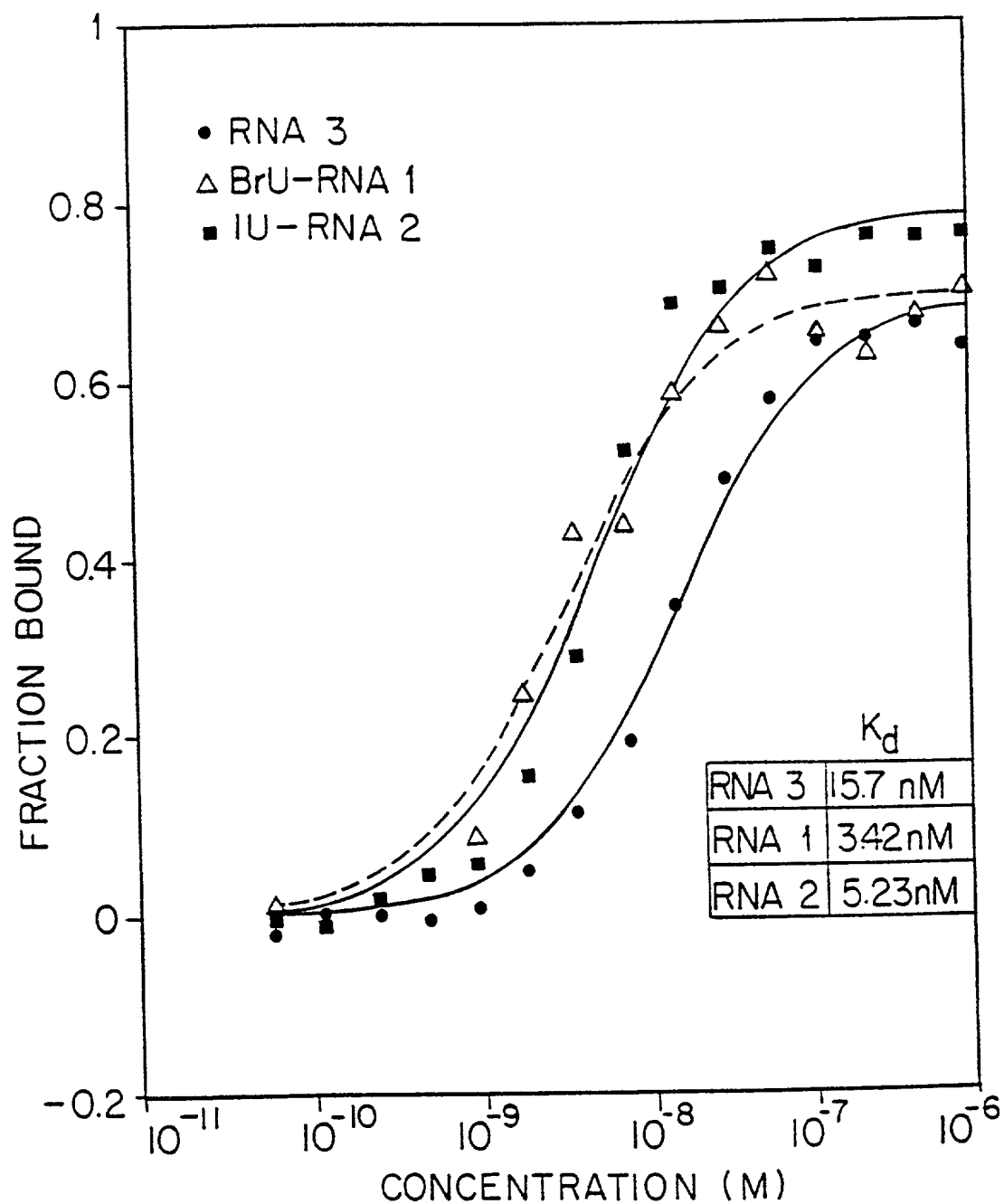


FIG. 7

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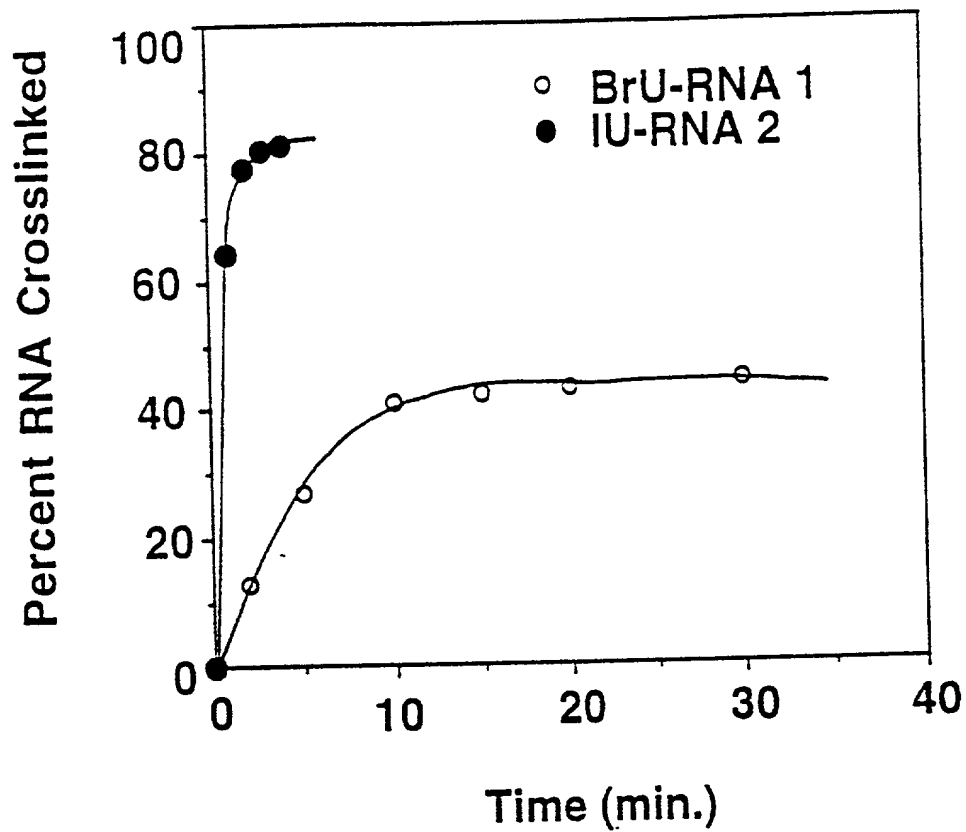


FIG. 8

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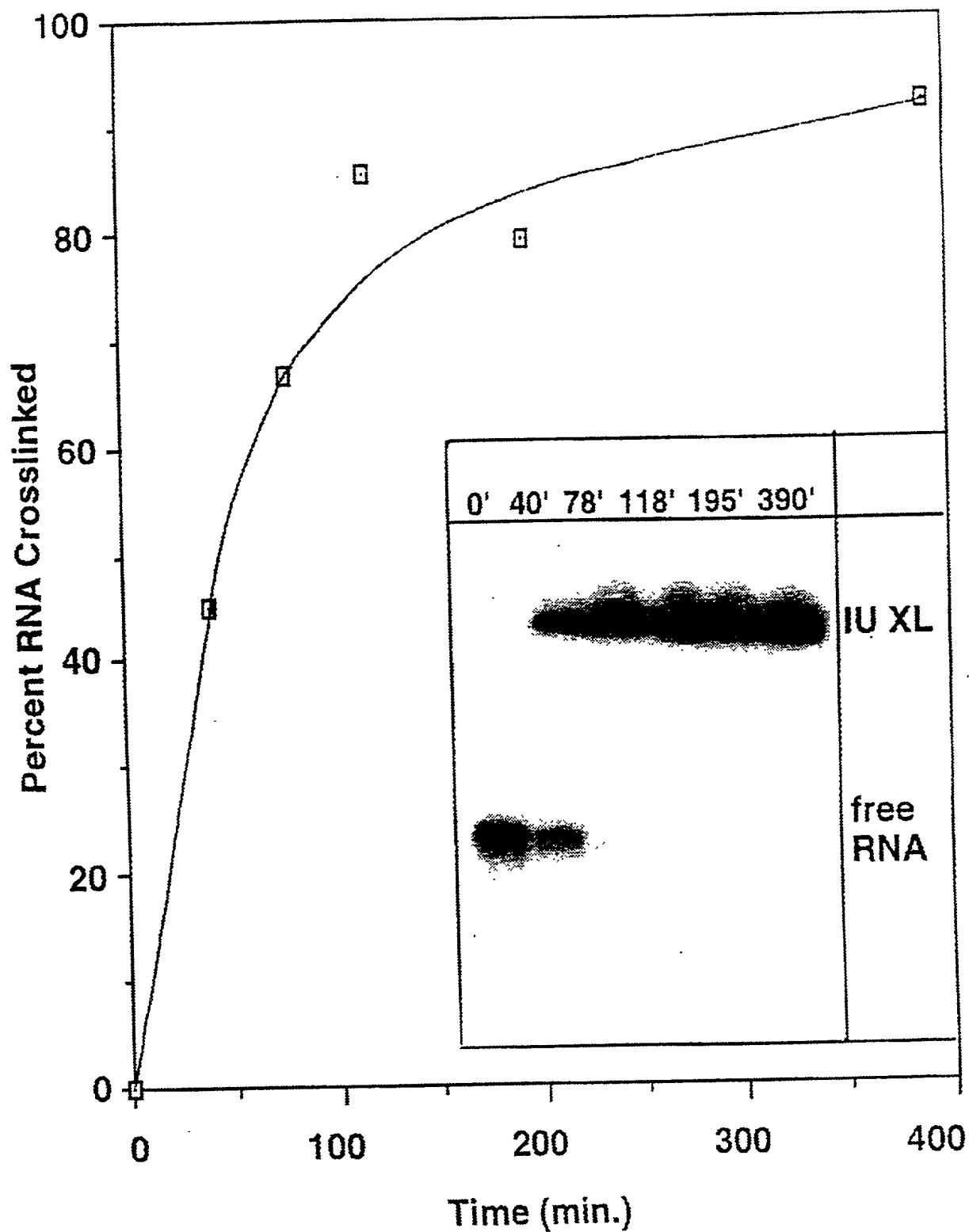


FIG. 9

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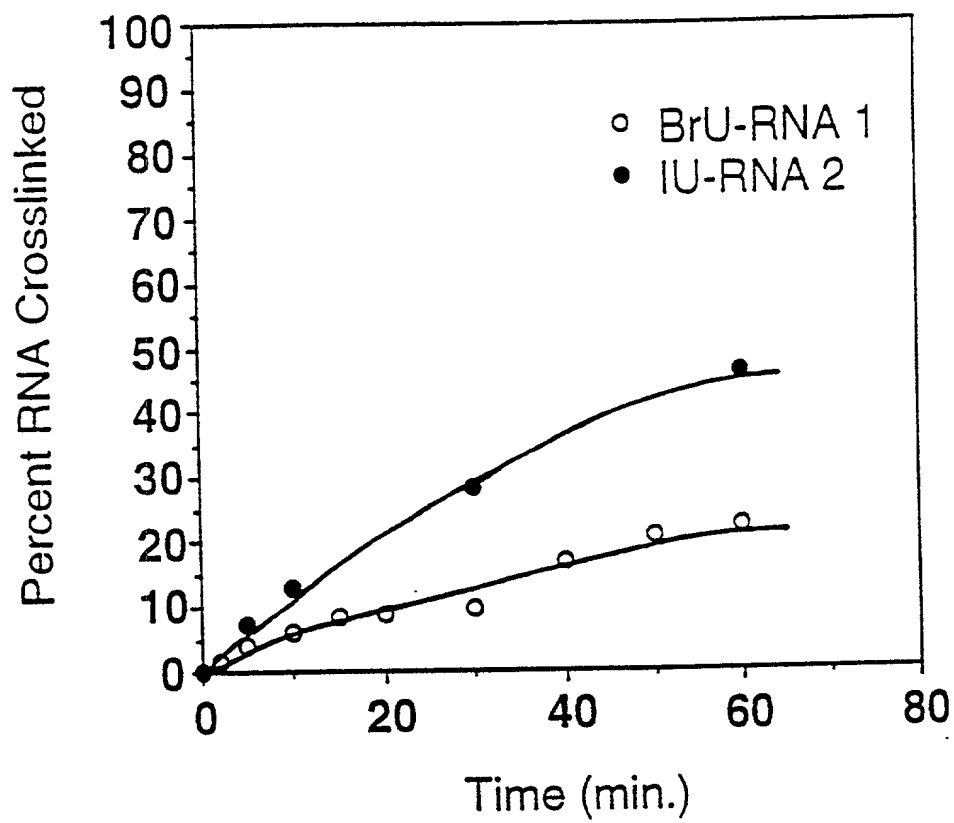


FIG. 10

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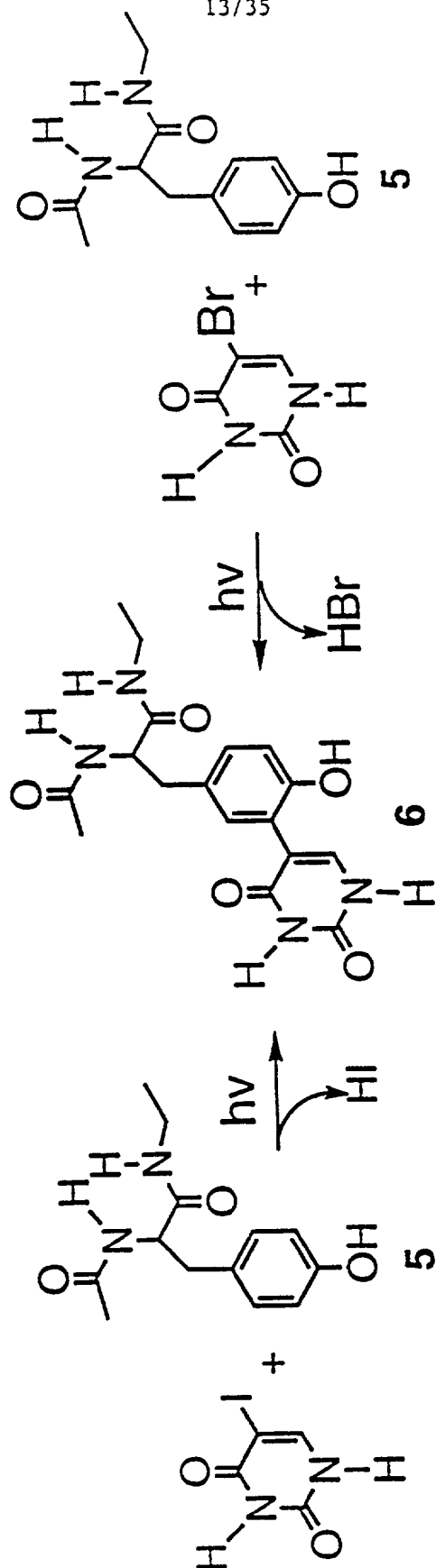


Fig. 11

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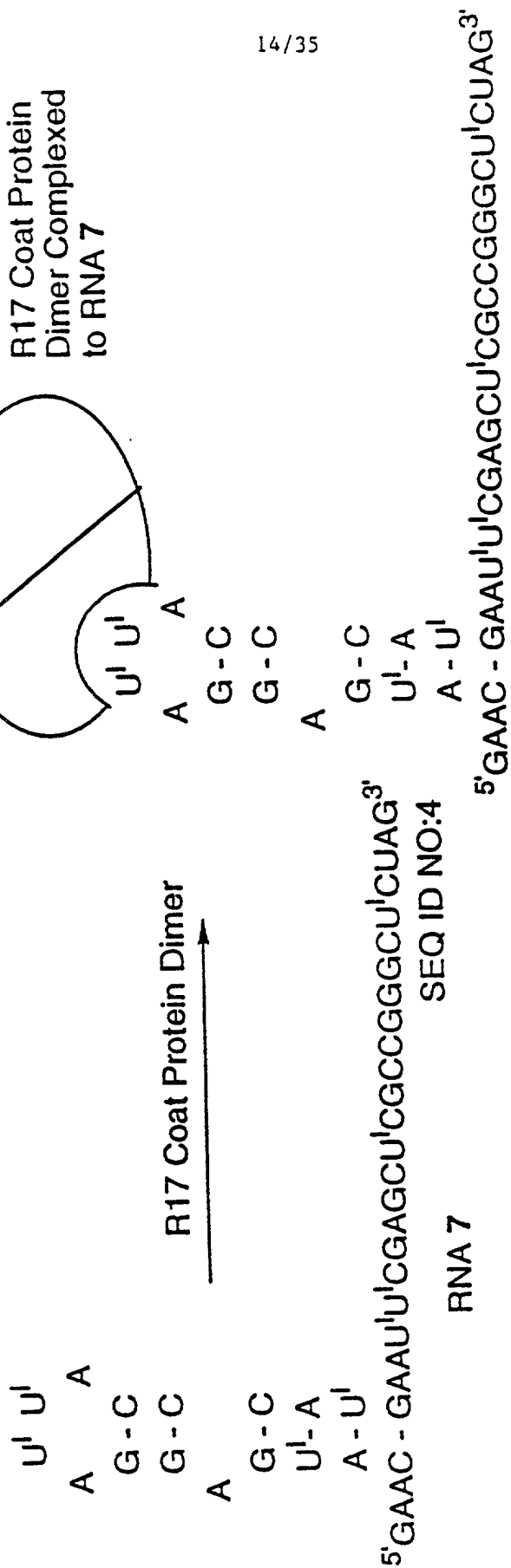


FIG. 12

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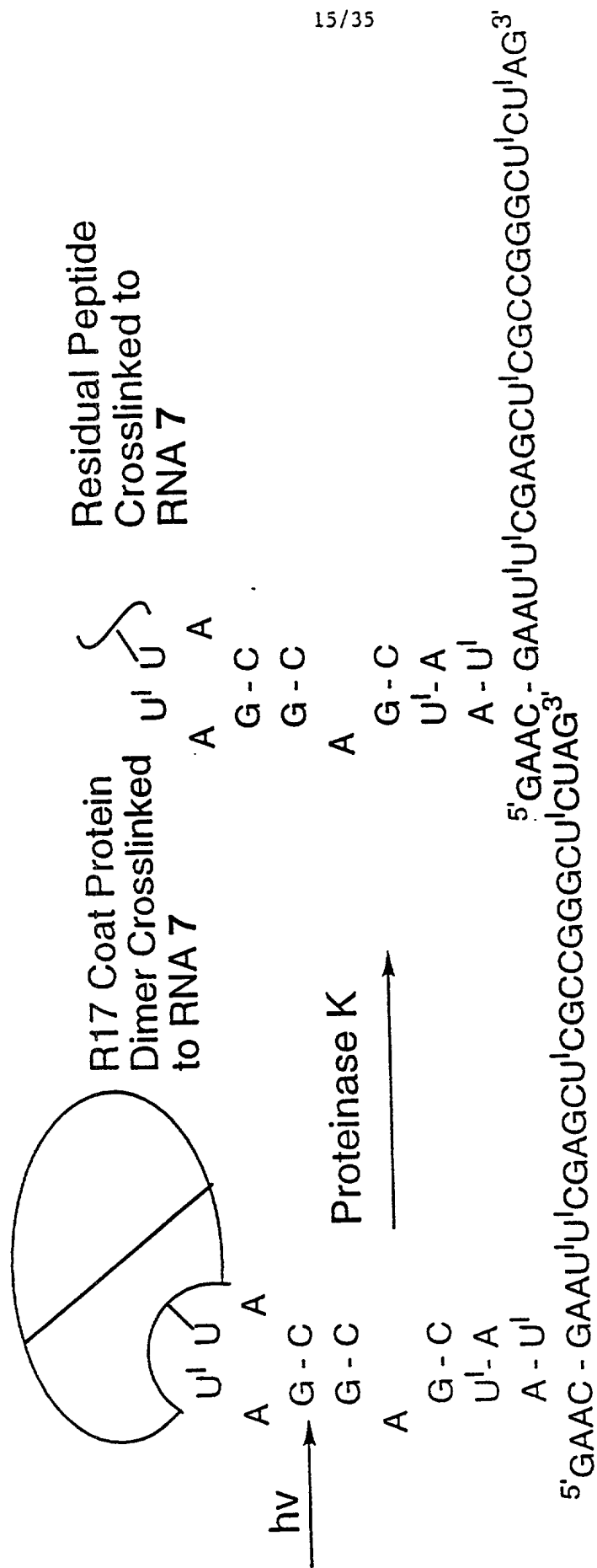


FIG. 12 (CONT'D)

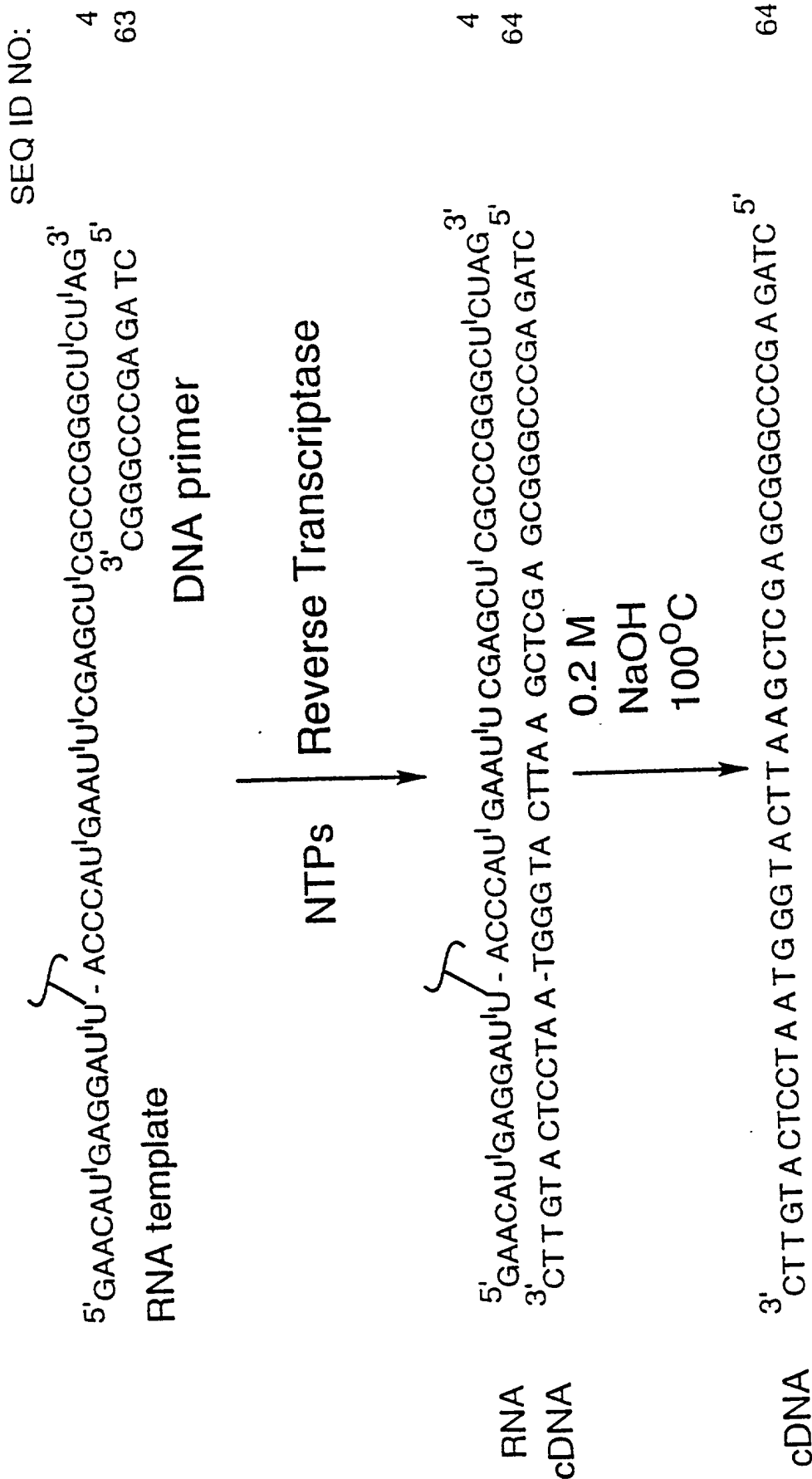


FIG. 13

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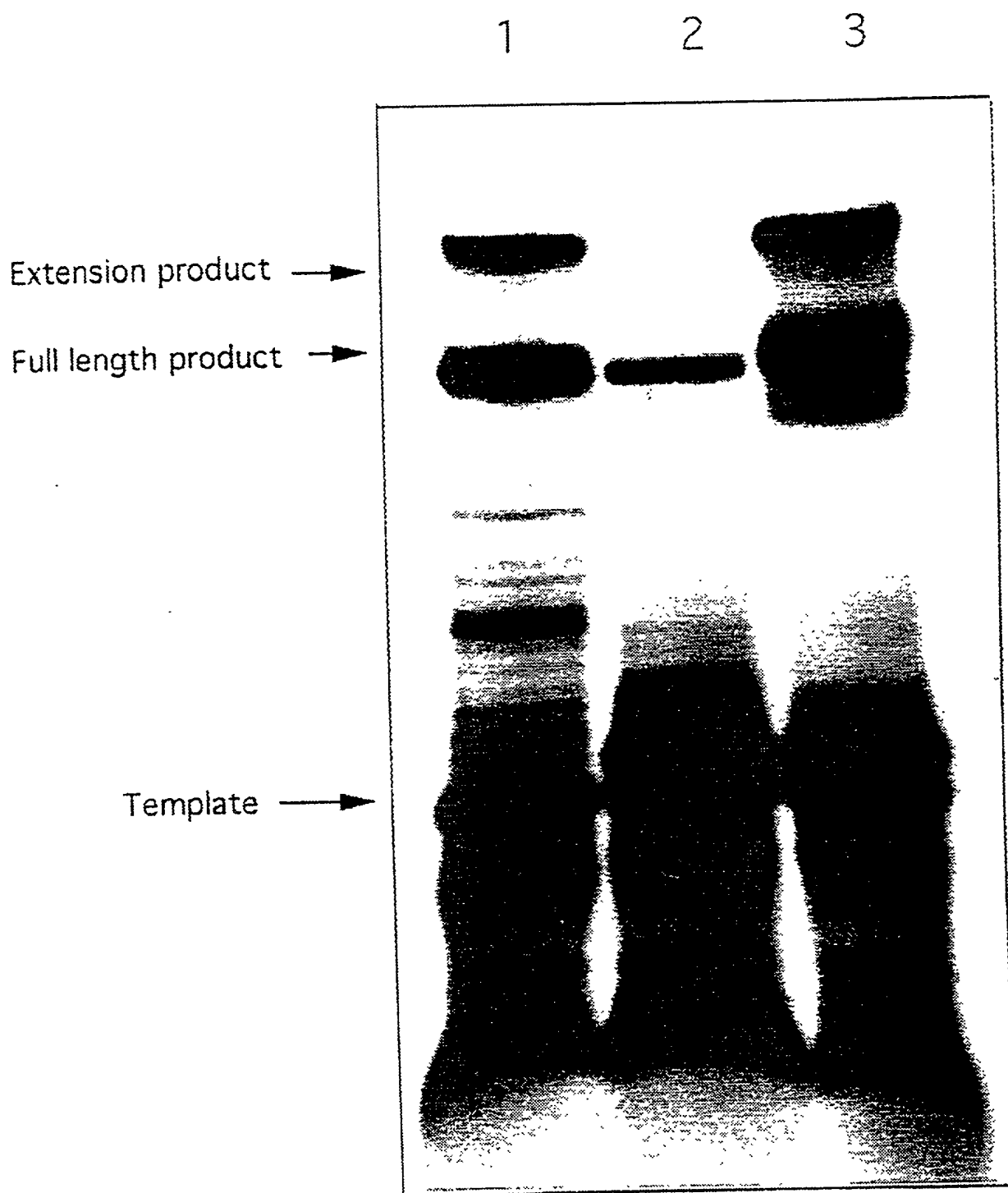


FIG. 14

SUBSTITUTE SHEET (RULE 26)

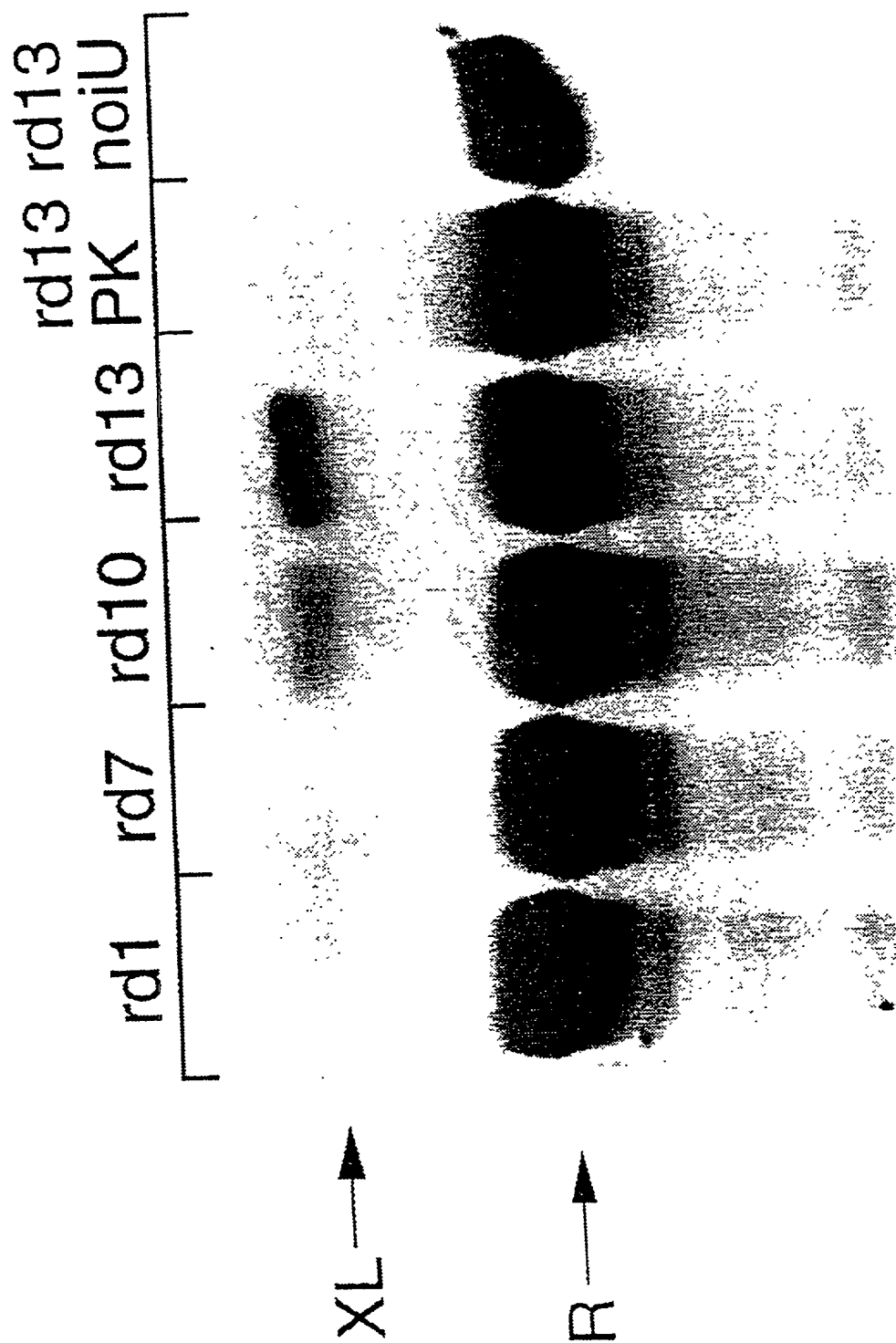
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FIG. 15

SEQ ID NO:

5 gguugcguuagagaaacacgguuuguggacucucuauucu

6a

LI-XL

class 1

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65	<u>uacggcguuACAagcaccuUGUUaaccuagugccu</u>	7
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25	<u>gugcagguuACAaacaac-guUGUUaaccuccucuu</u>	9
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6	<u>aagacgguuACAaaacgguUGUUaaccgcaacaccu</u>	13
66	<u>gauuggguuACAggcaccccUGUUaa-ccuaccacu</u>	14
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FIG. 16

SEQ ID NO:

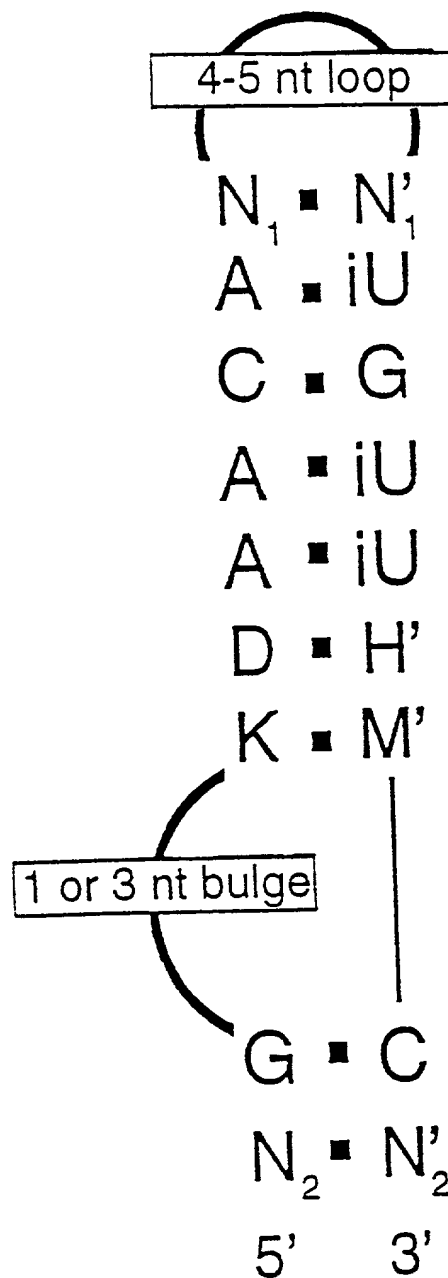
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FIG. 16 (CONT'D)

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class 1 consensus

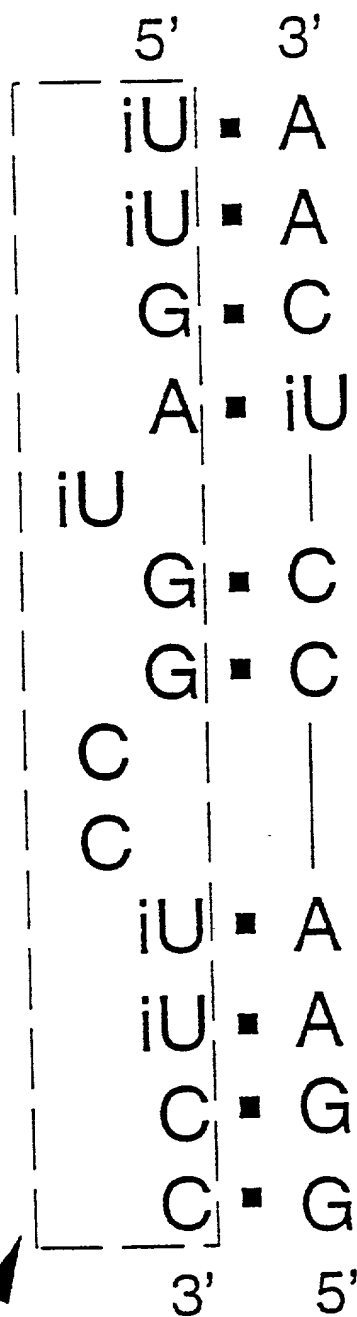


SEQ ID NO:56

FIG. 17

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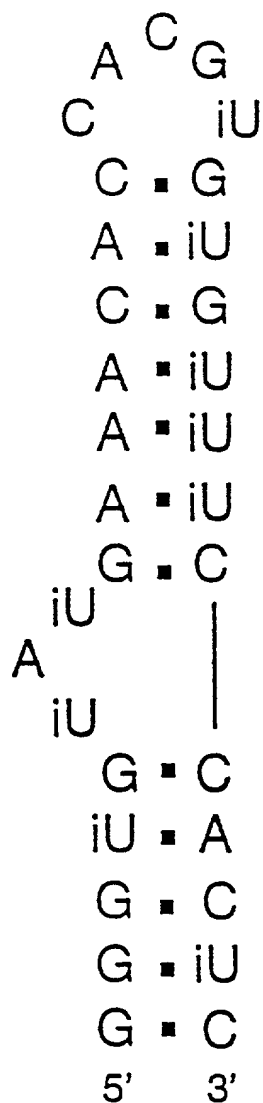
class 2 consensus



5' fixed region

SEQ ID NO:57

FIG. 17 (CONT'D)



trunc2

SEQ ID NO:58

FIG. 18A

t2	t2/	t2/	t2/	t2/	t2/	t2/
	Rev1	Rev1	Rev1	Rev1	Rev 2	Tat
	0'	4'	4'	4'/PK	4'	4'

Figure 1



FIG. 18B



FIG. 19A

[illegible]

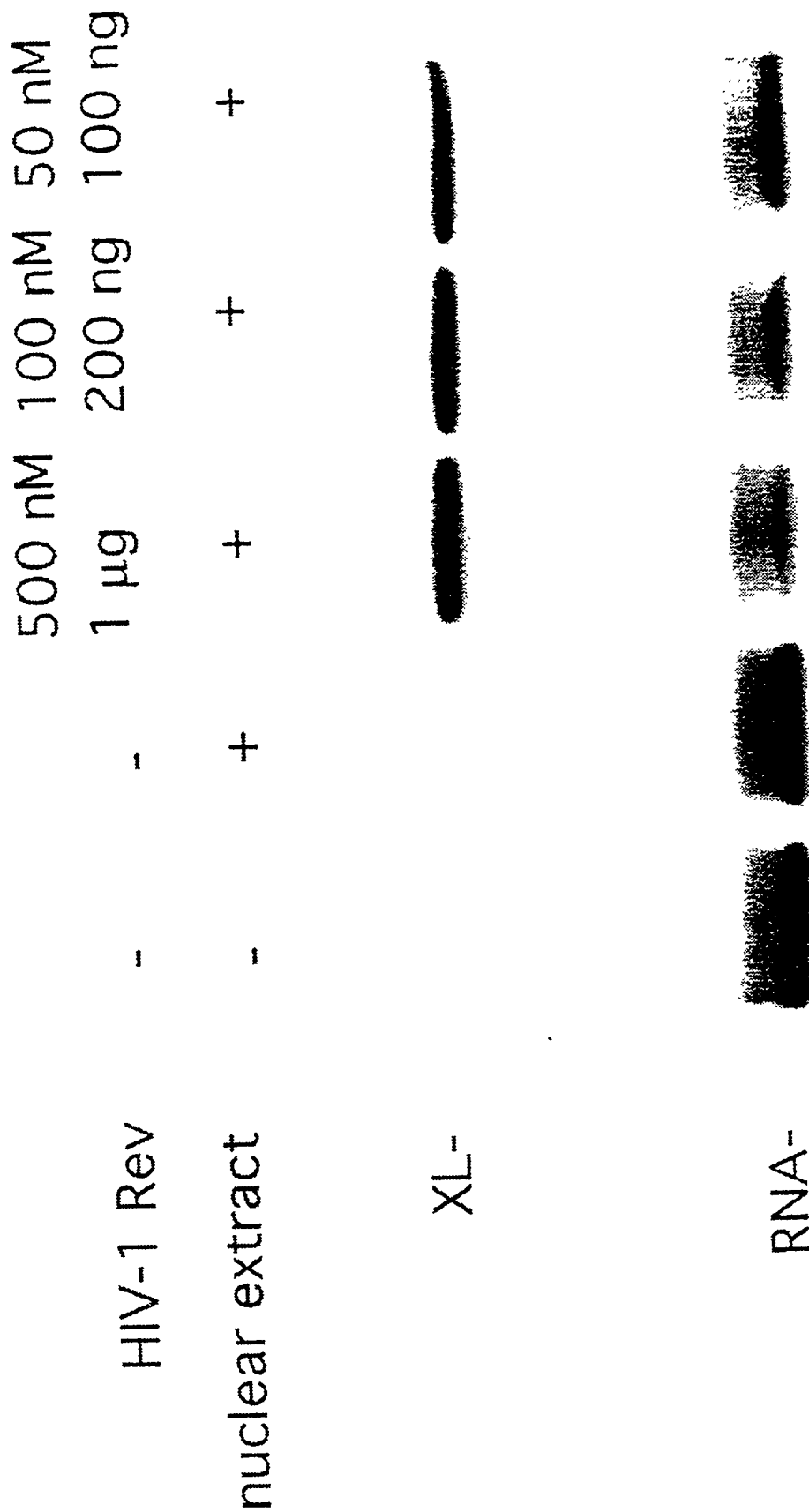


FIG. 20

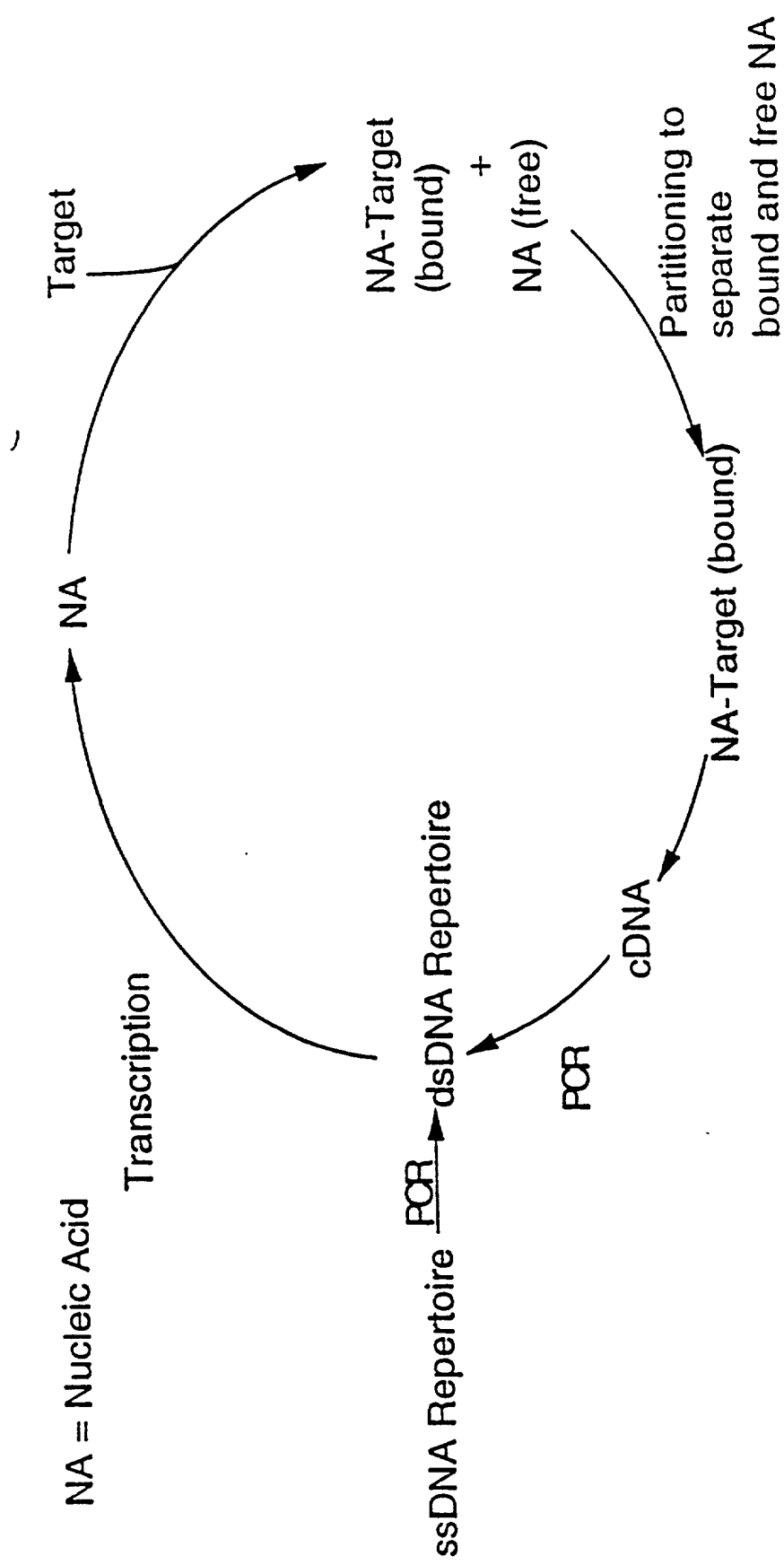


FIG. 21

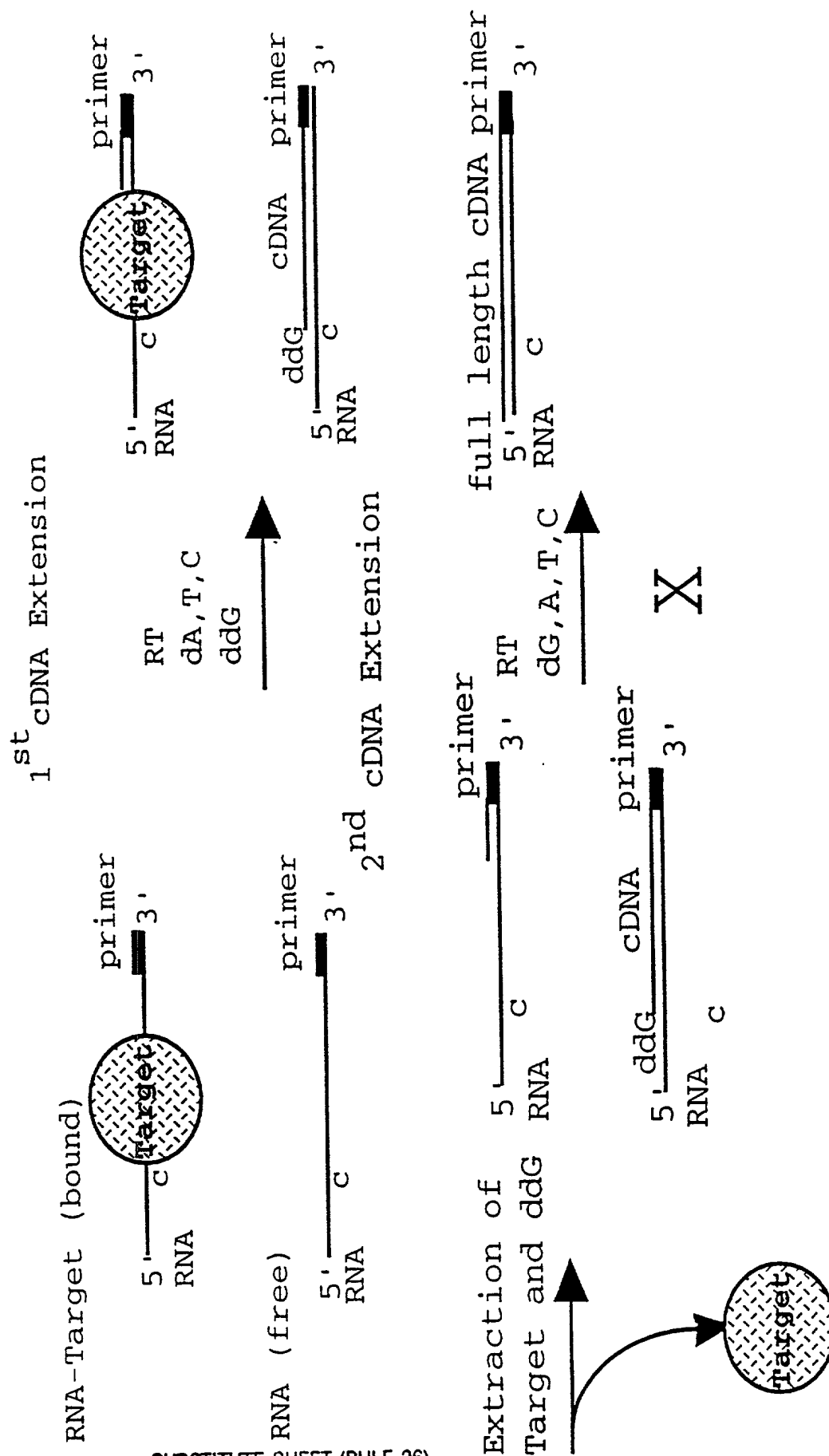


FIG. 22





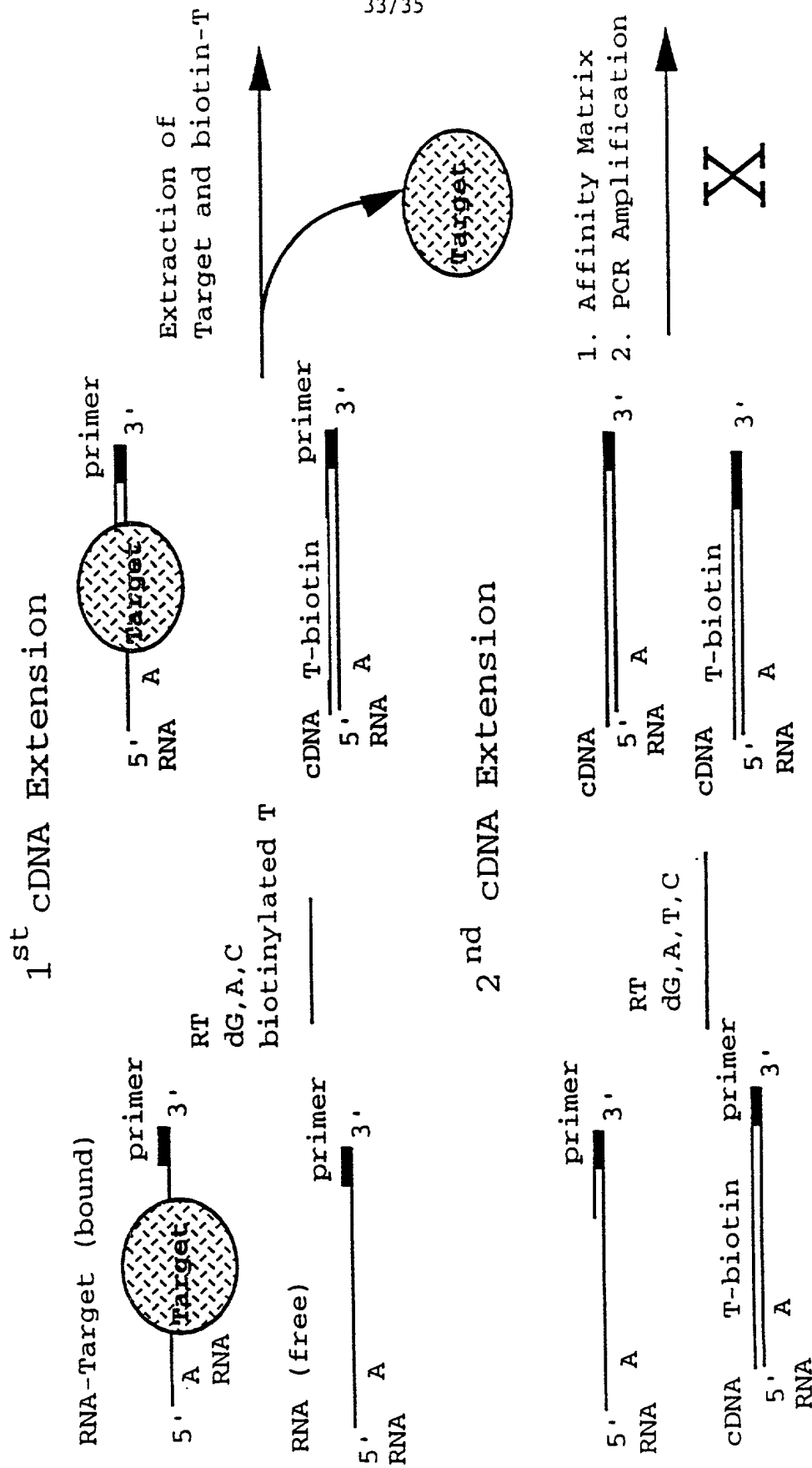


FIG. 25

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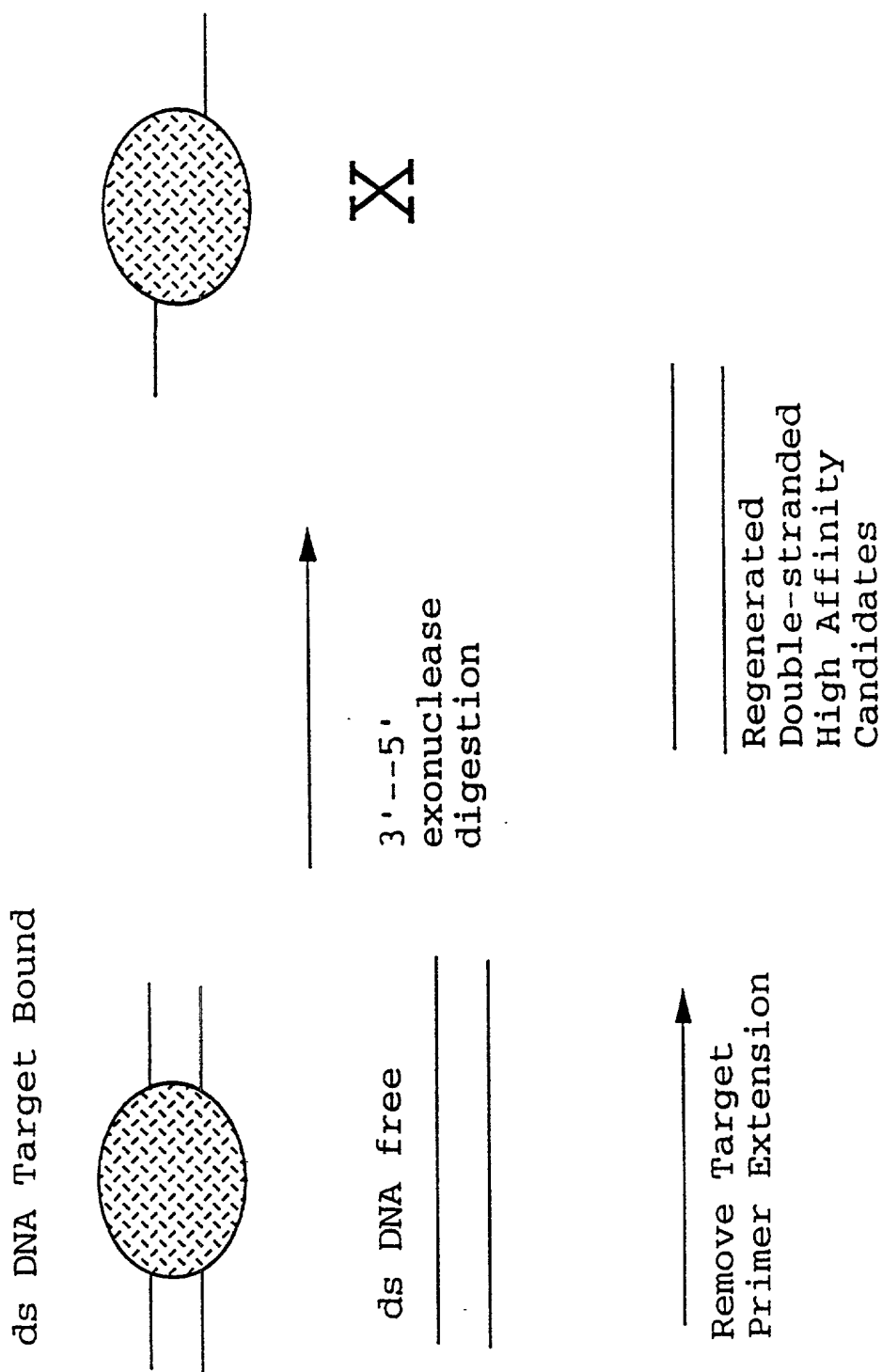
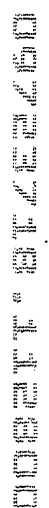
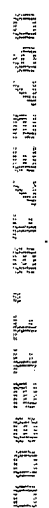
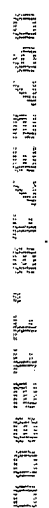
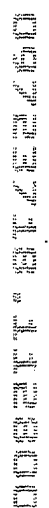


FIG. 26

THE **NEW** **YORK** **PUBLIC** **LIBRARY**

[illegible][illegible][illegible][illegible]

THE **NEW** **YORK** **PUBLIC** **LIBRARY**

[illegible]

THE **NEW** **YORK** **PUBLIC** **LIBRARY**

COMBINED DECLARATION AND POWER OF ATTORNEY(ORIGINAL, DESIGN, NATIONAL STAGE OF PCT, SUPPLEMENTAL, DIVISIONAL,
CONTINUATION OR C-I-P)

As a below named inventor, I hereby declare that:

COPY

TYPE OF DECLARATION

This declaration is of the following type: (check one applicable item below)

- ☐ original
☐ design
☒ supplemental

NOTE: If the declaration is for an International Application being filed as a divisional, continuation or continuation-in-part application, do not check next item; check appropriate one of last three items.

- ☐ national stage of PCT

NOTE: If one of the following 3 items apply, then complete and also attach ADDED PAGES FOR DIVISIONAL, CONTINUATION OR C-I-P.

- ☐ divisional
☐ continuation
☒ continuation-in-part (C-I-P)

INVENTORSHIP IDENTIFICATION

WARNING: If the inventors are each not the inventors of all the claims, an explanation of the facts, including the ownership of all the claims at the time the last claimed invention was made, should be submitted.

My residence, post office address and citizenship are as stated below next to my name. I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

TITLE OF INVENTION

Systematic Evolution of Ligands By Exponential Enrichment: Photoselection
of Nucleic Acid Ligands and Solution Selex

003274.1300
SECRET

SPECIFICATION IDENTIFICATION

the specification of which: (complete (a), (b) or (c))

- (a) ☐ is attached hereto.
(b) ☒ was filed on September 16, 1994 as PCT/US/94/10562 and nationalized on March 8, 1996 as ☒ Serial No. 08 / 612,895
or ☐ Express Mail No., as Serial No. not yet known _____
and was amended on _____ (if applicable).

NOTE: Amendments filed after the original papers are deposited with the PTO which contain new matter are not accorded a filing date by being referred to in the declaration. Accordingly, the amendments involved are those filed with the application papers or, in the case of a supplemental declaration, are those amendments claiming matter not encompassed in the original statement of invention or claims. See 37 CFR 1.57.

- (c) ☐ was described and claimed in PCT International Application No. _____ filed on _____ and as amended under PCT Article 19 on _____ (if any).

ACKNOWLEDGEMENT OF REVIEW OF PAPERS AND DUTY OF CANDOR

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information, which is material to patentability as defined in 37, Code of Federal Regulations, § 1.56

(also check the following items, if desired)

- ☒ and which is material to the examination of this application, namely, information where there is a substantial likelihood that a reasonable examiner would consider it important in deciding whether to allow the application to issue as a patent, and
☐ In compliance with this duty there is attached an information disclosure statement in accordance with 37 CFR 1.98.

PRIORITY CLAIM (35 U.S.C. § 119(a)-(d))

I hereby claim foreign priority benefits under Title 35, United States Code, § 119(a)-(d) of any foreign application(s) for patent or inventor's certificate or of any PCT international application(s) designating at least one country other than the United States of America listed below and have also identified below any foreign application(s) for patent or inventor's certificate or any PCT international application(s) designating at least one country other than the United States of America filed by me on the same subject matter having a filing date before that of the application(s) of which priority is claimed.

(complete (d) or (e))

- (d) ☒ no such applications have been filed.
(e) ☐ such applications have been filed as follows.

NOTE: Where item (c) is entered above and the International Application which designated the U.S. itself claimed priority check item (e), enter the details below and make the priority claim.

**PRIOR FOR US/PCT APPLICATION(S) FILED IN 12 MONTHS
(6 MONTHS FOR DESIGN) PRIOR TO THIS APPLICATION
AND ANY PRIORITY CLAIMS UNDER 35 U.S.C. § 119(a)-(d)**

COUNTRY (OR INDICATE IF PCT)	APPLICATION NUMBER	DATE OF FILING (day, month, year)	PRIORITY CLAIMED UNDER 37 USC 119
			<input type="checkbox"/> YES NO <input type="checkbox"/>
			<input type="checkbox"/> YES NO <input type="checkbox"/>
			<input type="checkbox"/> YES NO <input type="checkbox"/>
			<input type="checkbox"/> YES NO <input type="checkbox"/>
			<input type="checkbox"/> YES NO <input type="checkbox"/>

**CLAIM FOR BENEFIT OF PRIOR U.S. PROVISIONAL APPLICATION(S)
(34 U.S.C. § 119(e))**

I hereby claim the benefit under Title 35, United States Code, § 119(e) of any United States provisional application(s) listed below:

PROVISIONAL APPLICATION NUMBER

FILING DATE

_____ / _____	_____
_____ / _____	_____
_____ / _____	_____

**CLAIM FOR BENEFIT OF EARLIER US/PCT APPLICATION(S)
UNDER 35 U.S.C. 120**

- ☒ The claim for the benefit of any such applications are set forth in the attached ADDED PAGES TO COMBINED DECLARATION AND POWER OF ATTORNEY FOR DIVISIONAL, CONTINUATION OR CONTINUATION-IN-PART (C-I-P) APPLICATION.

(Declaration and Power of Attorney [1-1]—page 3 of 6)

008271872260

POWER OF ATTORNEY

Barry J. Swanson #33,215	Vanessa Appleby, #33,223
Thomas D. Bratschun #32,966	Diane Cruz, #33,960
Rosemary P. Kellogg #39,726	
Margaret M. Wall #33,462	

☐ Attached as part of this declaration and power of attorney is the authorization of the above-named attorney(s) to accept and follow instructions from my representative(s).

DIRECT TELEPHONE CALLS TO:
(Name and telephone number)

Barry J. Swanson
(303) 793-3333

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Full name of sole or first inventor

Inventor's signature

Date 5-17-96 Country of Citizenship US

Residence Boulder, Colorado

Post Office Address 1033 5th Street
Boulder, Colorado 80302

Michael	Willis
(GIVEN NAME)	(FAMILY (OR LAST NAME))

Inventor's signature

Date 5-8-96 Country of Citizenship US

Residence Louisville, Colorado

Post Office Address 786 W. Fir Court
Louisville, Colorado 80027

Tad Koch
(GIVEN NAME) (MIDDLE INITIAL OR NAME) FAMILY (OR LAST NAME)

Inventor's signature

Date May 21, 1996 Country of Citizenship US

Residence Boulder, Colorado

Post Office Address 2497 Powderhorn Lane
Boulder, Colorado 80303

(check proper) for any of the following added pages that form a part of this declaration)

☒ Signature for fourth and subsequent joint inventors. Number of pages added 1

. . .

☐ Signature by administrator(trix), executor(trix) or legal representative for deceased or incapacitated inventor. Number of pages added _____

. . .

☐ Signature for inventor who refuses to sign or cannot be reached by person authorized under 37 CFR 1.47. Number of pages added _____

. . .

☐ Added page for signature by one joint inventor on behalf of deceased inventor(s) where legal representative cannot be appointed in time (37 CFR 1.47).

. . .

☒ Added pages to combined declaration and power of attorney for divisional, continuation, or continuation-in-part (C-I-P) application.

☒ Number of pages added 3

. . .

☐ Authorization of attorney(s) to accept and follow instructions from representative.


. . .

(if no further pages form a part of this Declaration, then end this Declaration with this page and check the following item)

☐ This declaration ends with this page.

003211" 87/22/60

[illegible]

Steven _____ Ringquist _____
(GIVEN NAME) (MIDDLE INITIAL OR NAME) FAMILY (OR LAST NAME)
Inventor's signature 
Date 5/9/96 Country of Citizenship US
Residence Lyons, Colorado
Post Office Address 221 5th Avenue
Lyons, Colorado 80540

Kirk _____ Jensen _____
(GIVEN NAME) (MIDDLE INITIAL OR NAME) FAMILY (OR LAST NAME)

Inventor's signature _____
Date 5/15/96 Country of Citizenship US
Residence Boulder, Colorado
Post Office Address 1855 Athens Street, #204
Boulder, CO 80302

Brent
(GIVEN NAME)

Atkinson
(MIDDLE INITIAL OR NAME)

Atkinson
(FAMILY (OR LAST NAME))

Inventor's signature [Signature]

Date JUNE 3, 1996 Country of Citizenship US

Residence Boulder, Colorado WINTERTHUR, SWITZERLAND

Post Office Address ~~2711 Mapleton, #21~~ BRUEHLBARTENSTRASSE 7
~~Boulder, CO 80302~~ CH-8400
WINTERTHUR, SWITZERLAND

(Added Page to Combined Declaration and Power of Attorney for Signature by Fourth and Subsequent Inventors (1-2))

(complete this part only if this is a divisional, continuation or C-I-P application)

I hereby claim the benefit, under Title 35, United States Code, § 120, of any United States application(s) or PCT international application(s) designating the United States of America that is/are listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in that/those prior application(s) in the manner provided by the first paragraph of Title 35, United States Code, § 112, I acknowledge the duty to disclose information

(also check the following item, if desired)

(also check the following item, if desired)

☐ In compliance with this duty, there is attached an information disclosure statement in accordance with 37 C.F.R. 1.98.

Abstract

PRIOR U.S. APPLICATIONS OR PCT INTERNATIONAL APPLICATIONS DESIGNATING THE U.S. FOR BENEFIT UNDER 35 USC 120:					
U.S. APPLICATIONS			Status (check one)		
U.S. APPLICATIONS	U.S. FILING DATE	Patented	Pending	Abandoned	
1. 08 / 123,935	9/17/93		X		
2. 08 / 143,564	10/25/93		X		
3. 07 / 714,131	06/10/91	X			
PCT APPLICATIONS DESIGNATING THE U.S.					
PCT APPLI- CATION NO.	PCT FILING DATE	U.S. APPLICATION NOS. ASSIGNED (if any)			
4. _____	_____	0 / _____			
5. _____	_____	0 / _____			
6. _____	_____	0 / _____			

PRIOR U.S. APPLICATIONS OR PCT INTERNATIONAL APPLICATIONS DESIGNATING THE U.S. FOR BENEFIT UNDER 35 USC 120:					
U.S. APPLICATIONS			Status (check one)		
U.S. APPLICATIONS	U.S. FILING DATE	Patented	Pending	Abandoned	
1. 0 7/ 536,428	6/11/90			X	
2. 0 7/ 931,473	8/17/92	X			
3. 0 /					
PCT APPLICATIONS DESIGNATING THE U.S.					
PCT APPLI- CATION NO.	PCT FILING DATE	U.S. APPLICATION NOS. ASSIGNED (if any)			
4. _____		0 / _____			
5. _____		0 / _____			
6. _____		0 / _____			